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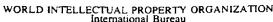
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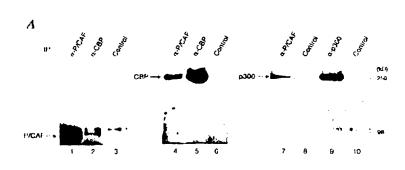
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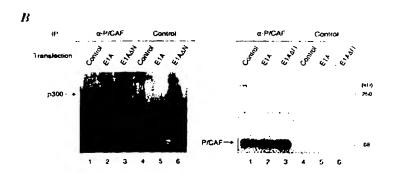
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### (54) Title: P300/CBP-ASSOCIATED TRANSCRIPTIONAL CO-FACTOR P/CAF AND USES THEREOF

#### (57) Abstract

The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones and which also binds to the p300/CBP cellular protein. The present invention further provides a nucleic acid encoding the P/CAF protein as well as a vector containing the nucleic acid and a host for the vector. A purified antibody which specifically binds the P/CAF protein is also provided. Also provided are methods of screening for compounds that inhibit or stimulate the transcription modulating and histone acetyltransferase activity of P/CAF and p300/CBP.





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# P300/CBP-ASSOCIATED TRANSCRIPTIONAL CO-FACTOR P/CAF AND USES THEREOF

#### BACKGROUND OF THE INVENTION

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#### Field of the Invention

The present invention provides a transcriptional co-factor, p300/CBP-associated factor (P/CAF), which modulates transcription through binding to the cellular transcription co-factors p300 and CBP and through acetylation of histones. Also provided are methods for screening for the presence of P/CAF and for substances which alter the transcription modulating effect and growth regulatory activity of P/CAF.

### Background Art

Cellular proteins p300 and CBP are global transcriptional coactivators that are involved in the regulation of various DNA-binding transcriptional factors (Janknecht and Hunter, 1996). Recently, p300 was found to be very closely related to CBP, a factor that binds selectively to the protein kinase A-phosphorylated form of CREB (3-5). Cellular factors p300 and CBP exhibit strong amino acid sequence similarity and share the capacity to bind both CREB and E1A (6-8). Although neither p300 nor CBP by itself binds to DNA, each can be recruited to promoter elements via interaction with sequence-specific activators and functions to be a transcriptional adaptor. For simplicity, p300 and CBP will be termed p300/CBP in the context of discussing their shared functional properties.

p300/CBP is a large protein consisting of over 2,400 amino acids, known to interact with a variety of DNA-binding transcriptional factors including nuclear hormone receptors (13,57), CREB (3,4, 7), c-Jun/v-Jun (9,11), YY1 (10), c-Myb/v-Myb (12,58), Sap-1a (59), c-Fos (11) and MyoD (60). DNA-binding factors recruit p300/CBP not only by direct but also indirect interactions through cofactors; for example, nuclear hormone receptors recruit p300/CBP directly as well as through indirect interactions, via SRC-1, which stimulates transcription by binding to various nuclear hormone receptors (13,61).



The transforming proteins encoded by adenovirus and several other small DNA tumor viruses disturb host cell growth control by interacting with cellular factors that normally function to repress cell proliferation. One of the most intensively studied of these viral proteins, the product of the adenovirus E1A gene, is itself sufficient for transformation (1). E1A transforming activity resides in two distinct domains, the targets of which include p300/CBP and products of the retinoblastoma (RB) susceptibility gene family (1,2). Interactions of E1A with p300/CBP and RB are thought to influence functionally distinct growth regulatory pathways, allowing the two domains to contribute additively to transformation (1).

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The paradigm for how E1A and functionally related viral proteins perturb cell growth regulation derives in large part from studies on their interactions with RB (1,2). The molecular function of E1A is based on its capacity to interfere with cellular protein-protein interactions. Since both E1A and various cellular targets bind to a site in RB termed the pocket domain (2), E1A can competitively disrupt the complex formation between RB and its cellular targets.

The second cellular factor implicated in E1A-dependent transformation, p300, is believed to inhibit G0/G1 exit, to activate certain enhancers, and to stimulate differentiation (1,2). E1A inhibits the p300/CBP-mediated transcriptional activation of many promoters (14). In one case that has been examined, the complex of p300 and YY1, E1A inhibits transcription without disrupting the complex (10).

The present invention provides a cellular protein designated P/CAF which binds to p300/CBP and plays an important role in both transcription and cell cycle regulation associated with a histone acetyltransferase activity. The present invention also provides a histone acetyltransferase activity in the p300/CBP cellular protein, thus providing targets for modulating transcription and cell cycle regulation in cells.

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### SUMMARY OF THE INVENTION

The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones and which also binds to the p300/CBP cellular protein.

The present invention further provides a nucleic acid encoding the P/CAF protein as well as a vector containing the nucleic acid and a host for the vector. A purified antibody which specifically binds the P/CAF protein is also provided.

In addition, also provided is a bioassay for screening substances for the ability to inhibit the transcription modulating activity of P/CAF and/or histone acetyltransferase activity, comprising contacting the substance with a system in which histone acetylation by P/CAF can be determined; determining the amount of histone acetylation by P/CAF in the presence of the substance, and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased amount of histone acetylation by -P/CAF in the presence of the substance indicating a substance that can inhibit the transcription modulating activity and/or histone acetyltransferase activity of P/CAF.

Furthermore, the present invention provides a bioassay for screening substances for the ability to inhibit the transcription modulating activity and/or histone acetyltransferase activity of P/CAF comprising contacting the substance with a system in which the p300 binding of P/CAF can be determined; determining the amount of p300 binding of P/CAF in the presence of the substance, and comparing the amount of p300 binding of P/CAF in the presence of the substance with the amount of p300 binding of P/CAF in the absence of the substance, a decreased amount of p300 binding of P/CAF in the presence of the substance indicating a substance that can inhibit the transcription modulating activity and/or histone acetyltransferase activity of P/CAF.

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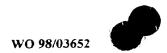
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Also provided is a method for determining the amount of P/CAF in a biological sample comprising contacting the biological sample with a polypeptide comprising the amino acid sequence of SEQ ID NO:3 under conditions whereby a P/CAF/p300 complex can be formed, and determining the amount of the P/CAF/p300 complex, the amount of the complex indicating the amount of P/CAF in the sample.

The present invention additionally provides a method for determining the amount of P/CAF in a biological sample comprising contacting the biological sample with an antibody which specifically binds P/CAF under conditions whereby a P/CAF/antibody complex can be formed; and determining the amount of the P/CAF/antibody complex, the amount of the complex indicating the amount of P/CAF in the sample.

Also provided herein is an assay for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of P/CAF, comprising: contacting the substance with a system in which histone acetylation by P/CAF can be determined; determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased or increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of P/CAF.

The present invention further provides an assay for screening substances for the ability to inhibit binding of P/CAF to p300/CBP comprising: contacting the substance with a system in which the P/CAF binding of P300/CBP can be determined; determining the amount of P/CAF binding of p300/CBP in the presence of the substance, and comparing the amount of binding of P/CAF to p300/CBP in the presence of the substance with the amount of binding of P/CAF to p300/CBP in the absence of the substance, a decreased amount of binding of P/CAF to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of P/CAF to p300/CBP.



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In addition, an assay is provided for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of p300/CBP, comprising: contacting the substance with a system in which histone acetylation by p300/CBP can be determined; determining the amount of histone acetylation by p300/CBP in the presence of the substance; and comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, a decreased or increased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of p300/CBP

Furthermore, the present invention provides an assay for screening substances for the ability to inhibit binding of a DNA-binding transcription factor to p300/CBP comprising: contacting the substance with a system in which the DNA-binding transcription factor binding of P300/CBP can be determined; determining the amount of DNA-binding transcription factor binding of p300/CBP in the presence of the substance; and comparing the amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance with the amount of binding of DNA-binding transcription factor to p300/CBP in the absence of the substance, a decreased amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of DNA-binding transcription factor to p300/CBP.

A method is also provided for inhibiting the transcription modulating activity of
P/CAF in a subject, comprising administering to the subject a transcription modulating
activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.

Also provided in the present invention is a method for stimulating the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity stimulating amount of a substance in a pharmaceutically acceptable carrier.

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Furthermore, the present invention provides a method for inhibiting the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.

Finally, the present invention additionally provides a method for stimulating the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity stimulating amount of a substance in a pharmaceutically acceptable carrier.

# BRIEF DESCRIPTION OF THE FIGURES

Figs. 1A-B. Fig 1A: P/CAF-p300/CBP interaction in vivo. Cell extract was immunoprecipitated with rabbit anti-P/CAF (lanes 1, 4, and 7), rabbit anti-CBP (lanes 2 15 and 5), and mouse anti-p300 (lane 9) antibodies. For controls, cell extract was precipitated with rabbit control IgG (lanes 3, 6, and 8) or mouse anti-HA monoclonal antibody (lane 10). The precipitates were analyzed by immunoblotting with anti-P/CAF (lanes 1-3), anti-CBP (lanes 4-6), and anti-p300 (lanes 7-10) antibodies. The positions of non-specific bands are indicated by asterisks. Fig. 1B: E1A inhibits the P/CAF-p300 20 interaction in vivo. Osteosarcoma cells were transfected with either control vector (lanes 1 and 4) or E1A- (lanes 2 and 5) or E1AAN- (lanes 3 and 6) expression vectors. Extract from the transfected subpopulation was immunoprecipitated with anti-P/CAF (lanes 1-3) or control (lanes 4-6) IgG. The precipitates were analyzed by immunoblotting with anti-p300 and anti-P/CAF. 25

Figs. 2A-F. P/CAF and E1A mediate antagonistic effects on cell cycle progression. HeLa cells (ATCC accession number CCL 2) were transfected by electroporation with 7  $\mu$ g of P/CAF-expression plasmid and/or 3  $\mu$ g of the full-length or the N-terminally deleted ( $\Delta 2$ -36) E1A 12S-expression plasmid as indicated in the figure These plasmids were constructed by subcloning FLAG-P/CAF and E1A cDNAs into



pCX (34) and pcDNAI (Invitrogen), respectively. All samples, in addition, contained 1  $\mu$ g of sorting plasmid (pCMV-IL2R) (31) and carrier plasmid (pCX) to normalize the total amount of DNA to 11  $\mu$ g. After transfection, cells were incubated in Dulbecco's modified Eagle's medium with 10% fetal bovine calf serum for 12 hours and subsequently labeled in medium containing 10  $\mu$ M bromo-deoxyuridine (BrdU) for 30 min. Subsequently, the transfected subpopulation was purified by magnetic affinity cell sorting and nuclei were analyzed by dual parameter flow cytometry as described (32). Histograms show percentages of cells in G1 and S phases. Abscissa values represent fluorescence intensity of bound anti-BrdU antibodies in log scale.

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Fig. 3. Histone acetyltransferase activity of P/CAF. Activity of hGCN5 (lanes 1 and 4) and P/CAF (lanes 2 and 5) that acetylates free histones (lanes 1-3) or histones in the nucleosome core particle (35) (lanes 4-6) was measured as described (36). Each reaction contains 0.3 pmol of affinity purified FLAG-hGCN5 or FLAG-P/CAF, 4 pmol of the histone octamer or the nucleosome core particle and 10 pmol of [1-14C]acetyl-CoA. Note that the histone octamer dissociates into dimers or tetramers under assay conditions. Acetylated histones were detected by autoradiography after separation by SDS-PAGE. The bands corresponding to acetylated histones H3 and H4 are indicated by arrows.

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# DETAILED DESCRIPTION OF THE INVENTION

As used in the specification and in the claims, "a" can mean one or more, depending upon the context in which it is used.

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#### P/CAF protein and fragments

The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones. The P/CAF protein can also bind to the amino acid region of SEQ ID NO 3

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(amino acid (aa) residues 1753 - 1966) of the cellular transcriptional factor, p300 (which has the complete amino acid sequence of SEQ ID NO:6 and the nucleotide sequence of SEQ ID NO:12), and the amino acid region of SEQ ID NO:6 (amino acid residues 1805 - 1854) of the cellular transcriptional factor, CBP (which has the complete amino acid sequence of SEQ ID NO:7 and the nucleotide sequence of SEQ ID NO:13). The P/CAF protein can be defined by any one or more of the typically used parameters. Examples of these parameters include, but are not limited to molecular weight (calculated or empirically determined), isoelectric focusing point, specific epitope(s), complete amino acid sequence, sequence of a specific region (e.g., N-terminus) of the amino acid sequence and the like

For example, The P/CAF protein can consist of the amino acid sequence of SEQ ID NO:1 or the P/CAF protein can comprise the amino acid sequence of SEQ ID NO:2 which represents the carboxy terminal end of the P/CAF protein and contains the histone acetyltransferase activity, or the amino acid sequence of SEQ ID NO:4, which represents the amino terminal end of the P/CAF protein, containing the binding site for p300/CBP. Because the amino-terminal region is specific for P/CAF it can be used to define and identify P/CAF.

As used herein, "purified" refers to a protein (polypeptide, peptide, etc.) that is sufficiently free of contaminants or cell components with which it normally occurs to distinguish it from the contaminants or other components of its natural environment. The purified protein need not be homogeneous, but must be sufficiently free of contaminants to be useful in a clinical or research setting, for example, in an assay for detecting antibodies to the protein. Greater levels of purity can be obtained using methods derived from well known protocols. Specific methods for purifying P/CAF proteins are known in the art.

As will be appreciated by those skilled in the art, the invention also includes those P/CAF polypeptides having slight variations in amino acid sequence which yield polypeptides equivalent to the P/CAF protein defined herein. Such variations may arise



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naturally as allelic variations (e.g., due to genetic polymorphism) or may be produced by human intervention (e.g., by mutagenesis of cloned DNA sequences), such as induced point, deletion, insertion and substitution mutants. Minor changes in amino acid sequence are generally preferred, such as conservative amino acid replacements, small internal deletions or insertions, and additions or deletions at the ends of the molecules. Substitutions may be designed based on, for example, the model of Dayhoff, et al. (37). These modifications can result in changes in the amino acid sequence, provide silent mutations, modify a restriction site, or provide other specific mutations.

Modifications to any of the P/CAF proteins or fragments can be made, while preserving the specificity and activity (function) of the native protein or fragment thereof. As used herein, "native" describes a protein that occurs in nature. The modifications contemplated herein can be conservative amino acid substitutions, for example, the substitution of a basic amino acid for a different basic amino acid. Modifications can also include creation of fusion proteins with epitope tags or known recombinant proteins or genes encoding them created by subcloning into commercial or non-commercial vectors (e.g., polyhistidine tags, flag tags, myc tag, glutathione-Stransferase [GST] fusion protein, xylE fusion reporter construct). Furthermore, the modifications can be such as do not affect the function of the protein or the way the protein accomplishes that function (e.g., its secondary structure or the ultimate result of the protein's activity). These products are equivalent to the P/CAF protein. The means for determining the function, way and result parameters are well known.

Having provided an example of a purified P/CAF protein, the invention also enables the purification of P/CAF homologs from other species and allelic variants from individuals within a species. For example, an antibody raised against the exemplary human P/CAF protein can be used routinely to screen preparations from different humans for allelic variants of the P/CAF protein that react with the P/CAF protein-specific antibody. Similarly, an antibody raised against an epitope, for example, from a conserved amino acid region of the human P/CAF protein can be used to routinely screen for homologs of the P/CAF protein in other species. A P/CAF protein can be

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routinely identified in and obtained from other species and from individuals within a species using the methods taught herein and others known in the art. For example, given the present sequence, the DNA encoding a conserved amino acid sequence can be used to probe genomic DNA or DNA libraries of an organism to predictably obtain the P/CAF gene for that organism. The gene can then be cloned and expressed as the

P/CAF protein and purified according to any of a number of routine, predictable methods. An example of the routine protein purification methods available in the art can be found in Pei et al. (38).

A purified polypeptide fragment of the P/CAF protein is also provided. The term "fragment" as used herein regarding a P/CAF protein, means a molecule of at least five contiguous amino acids of P/CAF protein that has at least one function shared by P/CAF protein or a region thereof. These functions can include antigenicity, binding capacity, acetyltransferase activity and structural roles, among others. The P/CAF fragment can be specific for a recited source. As used herein to describe an amino acid sequence (protein, polypeptide, peptide, etc.), "specific" means that the amino acid sequence is not found identically in any other source. The determination of specificity is made routine by the availability of computerized amino acid sequence databases and sequence comparison programs, wherein an amino acid sequence of almost any length can be quickly and reliably checked for the existence of identical sequences. If an identical sequence is not found, the protein is "specific" for the recited source. For example, a P/CAF fragment can be species-specific (e.g., found in the P/CAF protein of humans, but not of other species).

A fragment of the P/CAF protein having histone acetyltransferase activity can consist of the amino acid sequence of SEQ ID NO:2. A fragment of the P/CAF protein which binds to the amino acid sequence of SEQ ID NO:3 on p300 and the amino acid sequence of SEQ ID NO:9 on CBP can consist of the amino acid sequence of SEQ ID NO:4. To the extent that these fragments are specific for P/CAF, they can be used to identify and define P/CAF.

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An antigenic fragment of P/CAF protein is provided. An antigenic fragment has an amino acid sequence of at least about five consecutive amino acids of a P/CAF protein amino acid sequence and binds an antibody or elicits an immune response in an animal. An antigenic fragment can be selected by applying the routine technique of epitope mapping to P/CAF protein to determine the regions of the proteins that contain epitopes reactive with antibodies or are capable of eliciting an immune response in an animal. Once the epitope is selected, an antigenic polypeptide containing the epitope can be synthesized directly, or produced recombinantly by cloning nucleic acids encoding the antigenic polypeptide in an expression system, according to standard methods.

Alternatively, an antigenic fragment of the antigen can be isolated from the whole P/CAF protein or a larger fragment of the P/CAF protein by chemical or mechanical disruption. Fragments can also be randomly chosen from a known P/CAF protein sequence and synthesized. The purified fragments thus obtained can be tested to determine their antigenicity and specificity by routine methods.

#### Nucleic Acids Encoding P/CAF Protein

An isolated nucleic acid that encodes a P/CAF protein is also provided. As used herein, the term "isolated" means a nucleic acid separated or substantially free from at least some of the other components of the naturally occurring organism, for example, the cell structural components commonly found associated with nucleic acids in a cellular environment and/or other nucleic acids. The isolation of nucleic acids can therefore be accomplished by techniques such as cell lysis followed by phenol plus chloroform extraction, followed by ethanol precipitation of the nucleic acids (39). It is not contemplated that the isolated nucleic acids are necessarily totally free of all non-nucleic acid components or all other nucleic acids, but that the isolated nucleic acids are isolated to a degree of purification to be useful in clinical, diagnostic, experimental, or other procedures such as, for example, gel electrophoresis, Southern, Northern or dot blot hybridization, or polymerase chain reaction (PCR).

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A skilled artisan in the field will readily appreciate that there are a multitude of procedures which may be used to isolate the nucleic acids prior to their use in other procedures. These include, but are not limited to, lysis of the cell followed by gel filtration or anion exchange chromatography, binding DNA to silica in the form of glass beads, filters or diatoms in the presence of high concentrations of chaotropic salts, or ethanol precipitation of the nucleic acids.

The nucleic acids of the present invention can include positive and negative strand RNA as well as DNA and can include genomic and subgenomic nucleic acids found in the naturally occurring organism. The nucleic acids contemplated by the present invention include double stranded and single stranded DNA of the genome, complementary positive stranded cRNA and mRNA, and complementary cDNA produced therefrom and any nucleic acid which can selectively or specifically hybridize to the isolated nucleic acids provided herein. Stringent conditions (further described below) are used to distinguish selectively or specifically hybridizing nucleic acids from non-selectively and non-specifically hybridizing nucleic acids.

An isolated nucleic acid that encodes a P/CAF protein can be species-specific (i.e., does not encode the P/CAF protein of other species and does not occur in other species). Examples of the nucleic acids contemplated herein include the nucleic acid of SEQ ID NO:10 as well as the nucleic acids that encode each of the P/CAF proteins or fragments thereof described herein. P/CAF proteins and protein fragments can be routinely obtained as described herein and their structure (sequence) determined by routine means including the methods as used herein.

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P/CAF protein-encoding nucleic acids can be isolated from an organism in which they are normally found (e.g., humans), using any of the routine techniques. For example, a genomic DNA or cDNA library can be constructed and screened for the presence of the nucleic acid of interest using one of the present P/CAF protein-encoding nucleic acids as a probe. Methods of constructing and screening such libraries are well known in the art and kits for performing the construction and screening steps are

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commercially available (for example, Stratagene Cloning Systems, La Jolla, CA). Once isolated, the nucleic acid can be directly cloned into an appropriate vector, or if necessary, be modified to facilitate the subsequent cloning steps. Such modification steps are routine, an example of which is the addition of oligonucleotide linkers, which contain restriction sites, to the termini of the nucleic acid (See, for example, ref. 39).

P/CAF protein-encoding nucleic acids can also be synthesized. For example, a method of obtaining a DNA molecule encoding a specific P/CAF protein is to synthesize a recombinant DNA molecule which encodes the P/CAF protein. For example, nucleic acid synthesis procedures are routine in the art and oligonucleotides coding for a particular protein region are readily obtainable through automated DNA synthesis. A nucleic acid for one strand of a double-stranded molecule can be synthesized and hybridized to its complementary strand. One can design these oligonucleotides such that the resulting double-stranded molecule has either internal restriction sites or appropriate 5' or 3' overhangs at the termini for cloning into an appropriate vector.

Oligonucleotides complementary to or identical with the P/CAF proteinencoding nucleic acid sequence can be synthesized as primers for amplification reactions, such as PCR, or as probes to detect P/CAF protein encoding nucleic acids by various hybridization protocols (e.g., Northern blot, Southern blot, dot blot, colony screening, etc.).

Double-stranded molecules coding for relatively large proteins can readily be synthesized by first constructing several different double-stranded molecules that code for particular regions of the protein, followed by ligating these DNA molecules together. For example, Cunningham, et al. (40), have constructed a synthetic gene encoding the human growth hormone by first constructing overlapping and complementary synthetic oligonucleotides and ligating these fragments together. See also, Ferretti, et al. (41), wherein synthesis of a 1057 base pair synthetic bovine rhodopsin gene from synthetic oligonucleotides is disclosed.

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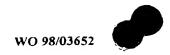
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By constructing a P/CAF protein-encoding nucleic acid in this manner, one skilled in the art can readily obtain any particular P/CAF protein with modifications at any particular position or positions. See also, U.S. Patent No. 5,503,995 which describes an enzyme template reaction method of making synthetic genes. Techniques such as this are routine in the art and are well documented. DNA encoding the P/CAF protein or P/CAF protein fragments can then be expressed *in vivo* or *in vitro*.

The nucleic acid encoding the P/CAF protein can be any nucleic acid that functionally encodes the P/CAF protein. To functionally encode the protein (i.e., allow the nucleic acid to be expressed), the nucleic acid can include, but is not limited to, expression control sequences, such as an origin of replication, a promoter, regions upstream or downstream of the promoter, such as enhancers that may regulate the transcriptional activity of the promoter, appropriate restriction sites to facilitate cloning of inserts adjacent to the promoter, antibiotic resistance genes or other markers which can serve to select for cells containing the vector or the vector containing the insert, and necessary information processing sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites and transcription termination sequences as well as any other sequence which may facilitate the expression of the inserted nucleic acid.

Preferred expression control sequences are promoters derived from metallothionine genes, actin genes, immunoglobulin genes, CMV, SV40, adenovirus, bovine papilloma virus, etc. A nucleic acid encoding a P/CAF protein can readily be determined based upon the genetic code for the amino acid sequence of the P/CAF protein and many nucleic acid sequences will encode a P/CAF protein. Modifications in the nucleic acid sequence encoding the P/CAF protein are also contemplated. Modifications that can be useful are modifications to the sequences controlling expression of the P/CAF protein to make production of P/CAF protein inducible or repressible as controlled by the appropriate inducer or repressor. Such means are standard in the art (see, e.g., ref. 39). The nucleic acids can be generated by means standard in the art, such as by recombinant nucleic acid techniques, as exemplified in the examples herein, and by synthetic nucleic acid synthesis or in vitro enzymatic synthesis.



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After a nucleic acid encoding a particular P/CAF protein of interest, or a region of that nucleic acid, is constructed, modified, or isolated, that nucleic acid can then be cloned into an appropriate vector, which can direct the *in vivo* or *in vitro* synthesis of that wild-type and/or modified P/CAF protein. The vector is contemplated to have the necessary functional elements that direct and regulate transcription of the inserted nucleic acid, as described above. The vector containing the P/CAF nucleic acid or nucleic acid fragment can be in a host (e.g., cell or transgenic animal) for expressing the nucleic acid. The P/CAF protein or fragment thereof can thus be produced in a host system containing the expression vector and its functional activity as described herein

can be demonstrated according to methods well known in the art.

There are numerous *E. coli* (*Escherichia coli*) expression vectors known to one of ordinary skill in the art useful for the expression of proteins. Other microbial hosts suitable for use include bacilli, such as *Bacillus subtilis*, and other enterobacteria, such as *Salmonella*, *Serratia*, as well as various *Pseudomonas* species. These prokaryotic hosts can support expression vectors which will typically contain expression control sequences compatible with the host cell (e.g., an origin of replication). In addition, any number of a variety of well-known promoters will be present, such as the lactose promoter system, a tryptophan (Trp) promoter system, a beta-lactamase promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence and have ribosome binding site sequences, for example, for initiating and completing transcription and translation. If necessary, an amino terminal methionine can be provided by insertion of a Met codon 5' and in-frame with the gene sequence. Also, the carboxy-terminal extension of the protein can be removed using standard oligonucleotide mutagenesis procedures.

Additionally, yeast expression can be used. There are several advantages to yeast expression systems. First, evidence exists that proteins produced in yeast secretion systems exhibit correct disulfide pairing. Second, post-translational glycosylation is efficiently carried out by yeast secretory systems. The Saccharomyces cerevisiae prepro-alpha-factor leader region (encoded by the  $MF\alpha-1$  gene) is routinely used to direct

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protein secretion from yeast (42). The leader region of pre-pro-alpha-factor contains a signal peptide and a pro-segment which includes a recognition sequence for a yeast protease encoded by the *KEX2* gene. This enzyme cleaves the precursor protein on the carboxyl side of a Lys-Arg dipeptide cleavage-signal sequence. The polypeptide coding sequence can be fused in-frame to the pre-pro-alpha-factor leader region. This construct is then put under the control of a strong transcription promoter, such as the alcohol dehydrogenase I promoter or a glycolytic promoter. The protein coding sequence is followed by a translation termination codon which is followed by transcription termination signals. Alternatively, the polypeptide encoding sequence of interest can be fused to a second protein coding sequence, such as Sj26 or  $\beta$ -galactosidase, used to facilitate purification of the resultant fusion protein by affinity chromatography. The insertion of protease cleavage sites to separate the components of the fusion protein is applicable to constructs used for expression in yeast.

Efficient post-translational glycosylation and expression of recombinant proteins can also be achieved in *Baculovirus* expression systems in insect cells.

Mammalian cells permit the expression of proteins in an environment that favors important post-translational modifications such as folding and cysteine pairing, addition of complex carbohydrate structures and secretion of active protein. Vectors useful for the expression of proteins in mammalian cells are characterized by insertion of the protein encoding sequence between a strong viral promoter and a polyadenylation signal. The vectors can contain genes conferring either gentamicin or methotrexate resistance for use as selectable markers. For example, the antigen and immunoreactive fragment coding sequence can be introduced into a Chinese hamster ovary (CHO) cell line using a methotrexate resistance-encoding vector. Presence of the vector RNA in transformed cells can be confirmed by Northern blot analysis and production of a cDNA or opposite strand RNA corresponding to the protein encoding sequence can be confirmed by Southern and Northern blot analysis, respectively. A number of other suitable host cell lines capable of secreting intact proteins have been developed in the art and include the CHO cell lines, HeLa cells, myeloma cell lines, Jurkat cells, and the like.



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Expression vectors for these cells can include expression control sequences, as described above. The vectors containing the nucleic acid sequences of interest can be transferred into the host cell by well-known methods, which vary depending on the type of cell host. For example, calcium chloride transfection is commonly utilized for prokaryotic cells, whereas calcium phosphate treatment or electroporation may be used for other cell hosts.

Alternative vectors for the expression of protein in mammalian cells, similar to those developed for the expression of human gamma-interferon, tissue plasminogen activator, clotting Factor VIII, hepatitis B virus surface antigen, protease Nexin I, and eosinophil major basic protein, can be employed. Further, the vector can include CMV promoter sequences and a polyadenylation signal available for expression of inserted nucleic acid in mammalian cells (such as COS7).

The nucleic acid sequences can be expressed in hosts after the sequences have been positioned to ensure the functioning of an expression control sequence. These expression vectors are typically replicable in the host organisms either as episomes or as an integral part of the host chromosomal DNA. Commonly, expression vectors can contain selection markers, e.g., tetracycline resistance or hygromycin resistance, to permit detection and/or selection of those cells transformed with the desired nucleic acid sequences (see, e.g., U.S. Patent 4,704,362).

The nucleic acids produced as described above can also be expressed in a host which is a non-human animal to create a transgenic animal, containing, in a germ or somatic cell, a nucleic acid comprising the coding sequence for all or a portion of the P/CAF protein, as well as all of the other regulatory elements required for expression of the P/CAF protein-encoding sequence. The animal will express the P/CAF gene or portion thereof to produce the P/CAF protein or protein fragment and such expression can be detected by determination of a particular phenotype unique to the transgenic animal expressing the transferred nucleic acid.

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The nucleic acid can be the nucleic acid of SEQ ID NO:10, a nucleic acid having a nucleotide sequence which encodes the P/CAF protein, a nucleic acid having a nucleotide sequence which encodes the protein of SEQ ID NO:1, as well as the nucleic acids that encode the proteins comprising the fragments of SEQ ID NOS:2 and 4.

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The nucleic acids of the invention can contain substitutions or deletions which provide a particular phenotype of interest. For example, various deletions or base substitutions can be introduced into the nucleic acid encoding the P/CAF protein for the purpose of studying the effects of these particular deletions or substitutions on the transcription modulation activity of the P/CAF protein. These effects can be monitored by observation of such characteristics as growth and development of the animal, the ability to develop tumors, survival rates and the like. The gene construct introduced into the animal cells to produce the transgenic animal can contain any of the regulatory elements described above to modulate expression of the foreign genes. As used herein, the term "phenotype" includes morphology, biochemical profiles, changes in tumor formation and other parameters that are affected by the presence of the P/CAF protein.

The transgenic animals of the invention can also be used in a method for determining the effectiveness of administering a nucleic acid encoding a functional P/CAF protein to a subject in need of a functional P/CAF protein. First, a nucleic acid encoding a nonfunctional P/CAF protein can be introduced into the animal's cells and expressed to yield a characteristic phenotype. Then, using standard gene therapy techniques, a nucleic acid encoding a functional P/CAF protein can be introduced into the animal's cells and the effects on the animal's phenotypic characteristics can be determined.

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Having provided and taught how to obtain a nucleic acid that encodes a P/CAF protein, an isolated nucleic acid that encodes a fragment of P/CAF protein is also provided. The nucleic acid encoding the fragment can be obtained using any of the methods applicable to the nucleic acid encoding the entire P/CAF protein. The nucleic acid fragment can encode a species-specific P/CAF protein fragment (e.g., found in the



P/CAF protein of humans, but not in the P/CAF proteins of other species). Nucleic acids encoding species-specific fragments of P/CAF protein are themselves species-specific or allele-specific fragments of the P/CAF gene.

Examples of fragments of a nucleic acid encoding a fragment of the P/CAF protein can include the nucleic acid sequences which encode the amino acid sequences of the fragments of SEQ ID NOS:2 or 4. The same routine computer analyses used to select these examples of fragments can be routinely used to obtain others. Fragments of P/CAF-encoding nucleic acids can be primers for PCR or probes, which can be species-specific, gene-specific or allele-specific. P/CAF-encoding nucleic acid fragments can encode antigenic or immunogenic fragments of P/CAF protein that can be used in therapeutic assays or screening protocols. P/CAF gene fragments can encode fragments of P/CAF protein having histone acetylase activity and/or p300/CBP binding activity as described above, as well as other uses that may become apparent.

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An isolated nucleic acid of at least ten nucleotides that selectively hybridizes with the nucleic acid of SEQ ID NO:10 under selected conditions is provided. For example, the conditions can be PCR amplification conditions and the hybridizing nucleic acid can be a primer consisting of a specific fragment of the reference sequence or a nearly identical nucleic acid that hybridizes only to the exemplified P/CAF-encoding nucleic acid or allelic variants thereof.

The invention provides an isolated nucleic acid that selectively hybridizes with the P/CAF-encoding nucleic acid sequence of SEQ ID NO:10 under stringent conditions. The hybridizing nucleic acid can be a probe that hybridizes only to the exemplified P/CAF-encoding nucleic acid sequence. Thus, the hybridizing nucleic acid can be a naturally occurring species-specific allelic variant of the exemplified P/CAF gene. The hybridizing nucleic acid can also include insubstantial base substitutions that do not prevent hybridization under the stated stringent conditions or affect either the function of the encoded protein, the way the protein accomplishes that function (e.g., its

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secondary structure) or the ultimate result of the protein's activity. The means for determining these parameters are well known.

As used herein to describe nucleic acids, the term "selectively hybridizes" excludes the occasional randomly hybridizing nucleic acids as well as nucleic acids that encode other known homologs of the P/CAF protein. The selectively hybridizing nucleic acids of the invention can have at least 70%, 73%, 78%, 80%, 85%, 88%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% complementarity with the segment and strand of the sequence to which it hybridizes. This list is not intended to exclude percent complementarity values between these values. The nucleic acids can be at least 10, 15, 16, 17, 18, 20, 21, 23, 24, 25, 30, 35, 40, 50, 100, 150, 200, 300, 500, 550, 750, 900, 950, or 1000 nucleotides in length or any intervening length, depending on whether the nucleic acid is to be used as a primer, probe or for protein expression. The hybridizing nucleic acid can comprise a region of at least ten nucleotides (up to full length) that is completely complementary to a unique region of the nucleic acid to which it hybridizes.

The nucleic acid can be an alternative coding sequence for the P/CAF protein, or can be used as a probe or primer for detecting the presence of or obtaining the P/CAF protein. If used as primers, the invention provides compositions including at least two nucleic acids which selectively hybridize with different regions of the nucleic acid so as to amplify a desired region. Depending on the length of the probe or primer, it can range between 70% complementary bases and full complementarity and still hybridize under stringent conditions.

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For example, for the purpose of obtaining or determining the presence of a nucleic acid encoding the P/CAF protein, the degree of complementarity between the hybridizing nucleic acid (probe or primer) and the sequence to which it hybridizes (P/CAF DNA in a sample) should be at least enough to exclude hybridization with a nucleic acid from another species. The invention provides examples of these nucleic acids of P/CAF, so that the degree of complementarity required to distinguish selectively



hybridizing from nonselectively hybridizing nucleic acids under stringent conditions can be clearly determined for each nucleic acid. It should also be clear that the hybridizing nucleic acids of the invention will not hybridize with nucleic acids encoding unrelated proteins (hybridization is selective) under stringent conditions.

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"Stringent conditions" refers to the washing conditions used in a hybridization protocol. In general, the washing conditions should be a combination of temperature and salt concentration chosen so that the denaturation temperature is approximately 5-20°C below the calculated  $T_m$  of the nucleic acid hybrid under study. The temperature and salt conditions are readily determined empirically in preliminary experiments in which samples of reference DNA immobilized on filters are hybridized to the probe or protein encoding nucleic acid of interest and then washed under conditions of different stringencies. For example, the nucleic acid sequence of SEQ ID NO:10 was used as a specific radiolabeled probe for the detection of messenger RNA transcribed from the P/CAF gene by performing hybridizations under stringent conditions. The  $T_m$  of such an oligonucleotide can be estimated by allowing 2°C for each A or T nucleotide, and 4°C for each G or C. For example, an 18 nucleotide probe of 50% G+C would, therefore, have an approximate  $T_m$  of 54°C.

The invention provides an isolated nucleic acid that selectively hybridizes with the P/CAF gene shown in the sequence set forth as SEQ ID NO:10 under stringent conditions. The invention further provides an isolated nucleic acid complementary to the nucleotide sequence set forth in SEQ ID NO:10.

#### 25 Antibodies to the P/CAF protein

A purified antibody and an antiserum containing polyclonal antibodies that specifically bind the P/CAF protein or antigenic fragment are also provided. The term "bind" means the well understood antigen/antibody binding as well as other nonrandom association with an antigen. "Specifically bind" as used herein describes an antibody or other ligand that does not cross react substantially with any antigen other than the one specified, in this case, an antigen of the P/CAF protein. Antibodies can be made as

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described in Harlow and Lane (33). Briefly, purified P/CAF protein or an antigenic fragment thereof can be injected into an animal in an amount and in intervals sufficient to elicit a humoral immune response. Serum polyclonal antibodies can be purified directly,

or spleen cells from the animal can be fused with an immortal cell line and screened for monoclonal antibody secretion, according to procedures well known in the art. Purified monospecific polyclonal antibodies that specifically bind the P/CAF antigen are also within the scope of the present invention. The antibodies of the present invention can

bind the protein of claim 1, the protein of claim 2, the protein of claim 3 and/or the

protein of claim 4, as well as any other proteins of the present invention.

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A ligand that specifically binds the antigen is also contemplated. The ligand can be a fragment of an antibody, such as, for example, an Fab fragment which retains P/CAF binding activity, or a smaller molecule designed to bind an epitope of the P/CAF antigen. The antibody or ligand can be bound to a substrate or labeled with a detectable moiety or both bound and labeled. The detectable moieties contemplated within the compositions of the present invention include those listed above in the description of the diagnostic methods, including fluorescent, enzymatic and radioactive markers.

The antibody can be bound to a solid support substrate or conjugated with a

detectable moiety or therapeutic compound or both bound and conjugated. Such
conjugation techniques are well known in the art. For example, conjugation of
fluorescent, radioactive or enzymatic moieties can be performed as described in the art
(33,43). The detectable moieties contemplated in the present invention can include
fluorescent, radioactive and enzymatic markers and the like. Therapeutic drugs
contemplated with the present invention can include cytotoxic moieties such as ricin A
chain, diphtheria toxin, pseudomonas exotoxin and other chemotherapeutic compounds

It is well understood by one of skill in the art that all of the above discussion regarding antibodies to P/CAF can also be applied with regard to production, characterization and use of antibodies which bind the p300/CBP protein or any of the DNA-binding transcription factors of this invention.



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The present invention also provides a method for determining the presence and thus the amount of P/CAF protein in a biological sample. As used herein, a biological sample includes any tissue or cell which would contain the P/CAF protein. Examples of cells include tissues taken from surgical biopsies, isolated from a body fluid or prepared in an *in vitro* tissue culture environment.

One example of determining the amount of P/CAF in a biological sample can comprise contacting the biological sample with a polypeptide comprising the amino acid sequence of SEQ ID NO 3 under conditions whereby a P/CAF/p300 complex can be formed; and determining the amount of the P/CAF/p300 complex, the amount of the complex indicating the amount of P/CAF in the sample. Determination of the amount of P/CAF/p300 complex can be accomplished through techniques standard in the art. For example, the complex may be precipitated out of a solution and detected by the addition of a detectable moiety conjugated to the p300 protein or by the detection of an antibody which binds p300 or the P/CAF protein, as taught in the Examples herein. Antibodies which bind p300 or the P/CAF protein can be either monoclonal or polyclonal antibodies and can be obtained as described herein. Detection of P/CAF/p300 complexes by the detection of the binding of antibodies reactive with p300 or the P/CAF protein can be accomplished using various immunoassays as are available in the art, as described below.

Alternatively, determination of the amount of P/CAF in a biological sample can comprise contacting the biological sample with a polypeptide comprising the amino acid sequence of SEQ ID NO:9 under conditions whereby a P/CAF/CBP complex can be formed; and determining the amount of the P/CAF/CBP complex, the amount of the complex indicating the amount of P/CAF in the sample. Determination of the amount of P/CAF/CBP complex can be accomplished through techniques standard in the art. For example, the complex may be precipitated out of a solution and detected by the addition of a detectable moiety conjugated to the CBP protein or by the detection of an antibody which binds either CBP or the P/CAF protein, as taught in the Examples

herein. Antibodies which bind CBP or the P/CAF protein can be either monoclonal or polyclonal antibodies and can be obtained as described herein. Detection of P/CAF/CBP complexes by the detection of the binding of antibodies reactive with CBP or the P/CAF protein can be accomplished using various immunoassays as are available in the art, as described below.

Another example of determining the amount of P/CAF in a biological sample comprises contacting the biological sample with an antibody which specifically binds P/CAF under conditions whereby a P/CAF/ antibody complex can be formed and determining the amount of the P/CAF/antibody complex, the amount of the complex indicating the amount of P/CAF in the sample. Antibodies which bind P/CAF can be either monoclonal or polyclonal antibodies and can be obtained as described herein. Determination of P/CAF/antibody complexes can be accomplished using various immunoassays as are available in the art, as described below.

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Immunoassays such as immunofluorescence assays, radioimmunoassays (RIA), immunoblotting and enzyme linked immunosorbent assays (ELISA) can be readily adapted for detection and measurement of P/CAF in a biological sample. Both polyclonal and monoclonal antibodies can be used in the assays. Available immunoassays are well known in the art and are extensively described in the patent scientific literature. See, for example, U.S. Patent Nos. 3,791,932, 3,839,153; 3,850,752; 3,850,578; 3,853,987; 3,867,517; 3,879,262; 3,901,654; 3,935,074; 3,984,533; 3,996,345; 4,034,074; and 4,098,876.

# 25 Screening assays for P/CAF

The present invention also provides a bioassay for screening substances for the ability to inhibit the histone acetyltransferase activity of P/CAF comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance under conditions whereby histone acetylation by P/CAF can occur; determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the

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amount of histone acetylation by P/CAF in the absence of the substance, a decreased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit the histone acetyltransferase activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to inhibit the histone acetyltransferase activity of P/CAF can be added to this system and assayed for inhibiting ability.

The present invention also provides a bioassay for screening substances for the ability to inhibit the transcription modulating activity of P/CAF, comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance under conditions whereby histone acetylation by P/CAF can occur; determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit the transcription modulating activity and cell cycle progression suppressing activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to inhibit the transcription modulating activity of P/CAF by interfering with the histone acetyltransferase activity of P/CAF can be added to this system and assayed for inhibiting ability.



Also provided in the present invention is a bioassay for screening substances for the ability to inhibit the binding of p300 to P/CAF, comprising contacting a system in which the binding of p300 to P/CAF can be determined, with the substance under conditions whereby the binding of p300 and P/CAF can occur; determining the amount of p300 binding to P/CAF in the presence of the substance, and comparing the amount of p300 binding to P/CAF in the presence of the substance with the amount of p300 binding to P/CAF in the absence of the substance, a decreased amount of p300 binding to P/CAF in the presence of the substance, a decreased amount of p300 binding to P/CAF in the presence of the substance indicating a substance that can inhibit the binding of p300 to P/CAF. The binding of p300 to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the p300 protein comprising the amino acid sequence of SEQ ID NO.3 and P/CAF. Alternatively, the system can comprise a cell extract produced from cells producing both p300 and P/CAF. Determination of the binding of p300 to P/CAF can be carried out as taught herein.

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Additionally provided in the present invention is a bioassay for screening substances for the ability to inhibit the binding of CBP to P/CAF, comprising contacting a system in which the binding of CBP to P/CAF can be determined, with the substance under conditions whereby the binding of CBP to P/CAF can occur, determining the amount of CBP binding to P/CAF in the presence of the substance, and comparing the amount of CBP binding to P/CAF in the presence of the substance with the amount of CBP binding to P/CAF in the absence of the substance, a decreased amount of CBP binding to P/CAF in the presence of the substance indicating a substance that can inhibit the binding of CBP to P/CAF. The binding of CBP to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the CBP protein comprising the amino acid sequence of SEQ ID NO 9 and P/CAF. Alternatively, the system can comprise a cell extract produced from cells producing both CBP and P/CAF. Determination of the binding of CBP to P/CAF can be carried out as taught herein.

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The present invention further contemplates a bioassay for screening substances for the ability to stimulate the histone acetyltransferase activity of P/CAF comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance; determining the amount of histone acetylation by P/CAF in the presence of the substance, and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, an increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can stimulate the histone acetyltransferase activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to stimulate the histone acetyltransferase activity of P/CAF can be added to this system and assayed for stimulating ability.

The present invention further contemplates a bioassay for screening substances for the ability to stimulate the transcription modulating activity of P/CAF comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance; determining the amount of histone acetylation by P/CAF in the presence of the substance, and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, an increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can stimulate the transcription modulating activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples.

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Thus, the compound to be tested for the ability to stimulate the transcription modulating activity of P/CAF by increasing the histone acetyltransferase activity of P/CAF can be added to this system and assayed for stimulating ability.

The present invention further provides a bioassay for screening substances for the ability to stimulate binding of p300 to P/CAF, comprising contacting a system in which the binding of p300 to P/CAF can be determined, with the substance under conditions whereby the binding of p300 to P/CAF can occur; determining the amount of p300 binding to P/CAF in the presence of the substance; and comparing the amount of p300 binding to P/CAF in the presence of the substance with the amount of p300 binding to P/CAF in the absence of the substance, an increased amount of p300 binding to P/CAF in the presence of the substance indicating a substance that can stimulate the binding of p300 to P/CAF. The binding of p300 to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the p300 protein comprising the amino acid sequence of SEQ ID NO:3 and P/CAF. Alternatively, the system can comprise a cell extract produced from cells producing both p300 and P/CAF. Determination of the binding of p300 to P/CAF can be carried out as taught herein.

Additionally provided in the present invention is a bioassay for screening substances for the ability to stimulate the binding of CBP to P/CAF, comprising contacting a system in which the binding of CBP to P/CAF can be determined, with the substance under conditions whereby the binding of CBP to P/CAF can occur; determining the amount of CBP binding to P/CAF in the presence of the substance, and comparing the amount of CBP binding to P/CAF in the presence of the substance with the amount of CBP binding to P/CAF in the absence of the substance, an increased amount of CBP binding to P/CAF in the presence of the substance indicating a substance that can stimulate the binding of CBP to P/CAF. The binding of CBP to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the CBP protein comprising the amino acid sequence of SEQ ID NO:9 and P/CAF. Alternatively, the system can comprise a cell

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extract produced from cells producing both CBP and P/CAF. Determination of the binding of CBP to P/CAF can be carried out as taught herein.

### Transcription modulating activity of P/CAF

The present invention contemplates a method for inhibiting the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity inhibiting amount of a substance in a pharmaceutically acceptable carrier. For example, the substance can be identified according to the protocols provided herein as one that can inhibit the transcription modulating activity of P/CAF by preventing the binding of P/CAF to p300/CBP or by inhibiting the histone acetyltransferase activity of P/CAF as well as by any other inhibitory mechanism as identified by the protocols provided herein. Inhibition of the transcription modulating activity of P/CAF in a subject is desirable, for example, to inhibit HIV TAT-mediated transcription and therefore, the method of the present invention can be used to treat HIV-infected subjects.

The substance can be in a pharmaceutically acceptable carrier. By "pharmaceutically acceptable" is meant a material that is not biologically or otherwise undesirable, i.e., the material may be administered to a subject, along with the substance, without causing any undesirable biological effects or interacting in a deleterious manner with any of the other components of the pharmaceutical composition in which it is contained. The carrier would naturally be selected to minimize any degradation of the active ingredient and to minimize any adverse side effects in the subject.

The transcription modulating activity and/or histone acetyltransferase activity of P/CAF can be inhibited in a subject by administering to the subject a substance which binds p300/CBP at the P/CAF binding site or a substance which binds the P/CAF protein at the p300/CBP binding site, the ultimate result being that P/CAF and p300/CBP do not bind with one another and P/CAF cannot exert its transcription modulating and/or histone acetyltransferase effect. The substance can be a protein, such as an antibody which binds the P/CAF protein binding site at or near the p300/CBP



binding site, thereby preventing its binding or an antibody which binds the p300/CBP protein at or near the P/CAF binding site, thereby preventing its binding. The substance can also bind the histone acetyltransferase site on P/CAF or at the acetylation site on the histone, thereby preventing acetylation by P/CAF.

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The substance which binds p300/CBP, the P/CAF protein or the histone and has the net effect of inhibiting the transcription modulating effect and or histone acetyltransferase activity of P/CAF in the cell can be delivered to a cell in the subject by mechanisms well known in the art.

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Alternatively, a nucleic acid encoding a protein which binds either to p300/CBP or the P/CAF protein and has the net effect of inhibiting the transcription modulating effect and/or histone acetyltransferase activity of P/CAF in the cell can be delivered to a cell in the subject by gene transduction mechanisms well known in the art. For example, nucleic acid can be introduced by liposomes as well as via retroviral or adeno-associated viral vectors, as described below.

The substance which inhibits the transcription modulating effect and/or histone acetyltransferase activity of P/CAF can be an antisense RNA or an antisense DNA which binds the RNA or DNA of P/CAF, thereby preventing translation or transcription of the RNA or DNA encoding P/CAF and having the net effect of inhibiting the transcription modulating effect and/or histone acetyltransferase activity of P/CAF by inhibiting P/CAF production. The antisense RNA of the present invention can be generated from the nucleic acid of SEQ ID NO:14 (human) or SEQ ID NO:15 (mouse). Furthermore, the antisense DNA can be a phosphorothioate oligodeoxyribonucleotide having the nucleotide sequence of SEQ ID NO:16 (human) or of SEQ ID NO:17 (mouse). The mouse antisense RNA can be used to inhibit the activity of mouse P/CAF, having the nucleotide sequence of SEQ ID NO:18 and the amino acid sequence of SEQ ID NO:8. The present invention also contemplates an antisense nucleic acid sequence which can bind the DNA or RNA of any of the transcription factors or other proteins now known or later identified to bind P/CAF, thereby inhibiting expression of the gene products of



these proteins and having the net effect of inhibiting the transcription modulating effect and/or histone acetyltransferase activity of P/CAF.

The antisense nucleic acid can comprise a typical nucleic acid, but the antisense nucleic acid can also be a modified nucleic acid or a derivative of a nucleic acid such as a phosphorothioate analogue of a nucleic acid. The composition can comprise, for example, an antisense RNA that specifically binds an RNA encoded by the gene encoding the serum protein. Antisense RNAs can be synthesized and used by standard methods (62).

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Antisense RNA can inhibit gene expression by forming an RNA/RNA duplex between the antisense RNA and the RNA transcribed from the target gene. The precise mechanism by which this duplex formation decreases the production of the protein encoded by the endogenous gene probably involves binding of complementary regions of the normal sense mRNA and the antisense RNA strand with duplex formation in a manner that blocks RNA processing and translation. Alternative mechanisms include the formation of a triplex between the antisense RNA and duplex DNA or the formation of an DNA-RNA duplex with subsequent degradation of DNA-RNA hybrids by RNAse H. Furthermore, an antigene effect can result from certain DNA-based oligonucleotides via triple-helix formation between the oligomer and double-stranded DNA which results in the repression of gene transcription. Regardless of the specific molecular mechanism, the present invention results in inhibition of expression of the P/CAF gene by the introduced and replicated DNA resulting in inhibition of the transcription modulating and/or histone acetyltransferase activity of P/CAF, by a reduction in the expression of the nucleic acid to which the antisense nucleic acid is hybridized, and therefore a reduction of the gene product from the targeted gene.

The antisense nucleic acid may be obtained by any number of techniques known to one skilled in the art. One method of constructing an antisense nucleic acid is to synthesize a recombinant antisense DNA molecule. For example, oligonucleotide synthesis procedures are routine in the art and oligonucleotides coding for a particular

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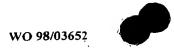
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protein or regulatory region are readily obtainable through automated DNA synthesis. A nucleic acid for one strand of a double-stranded molecule can be synthesized and hybridized to its complementary strand. One can design these oligonucleotides such that the resulting double-stranded molecule has either internal restriction sites or appropriate 5' or 3' overhangs at the termini for cloning into an appropriate vector. Double-stranded molecules coding for relatively large proteins or regulatory regions can be synthesized by first constructing several different double-stranded molecules that code for particular regions of the protein or regulatory region, followed by ligating these DNA molecules together. Once the appropriate DNA molecule is synthesized, this DNA can be cloned downstream of a promoter in an antisense orientation. Techniques such as this are routine in the art and are well documented.

An example of another method of obtaining an antisense nucleic acid is to isolate that nucleic acid from the organism in which it is found and clone it in an antisense orientation. For example, a DNA or cDNA library can be constructed and screened for the presence of the nucleic acid of interest. Methods of constructing and screening such libraries are well known in the art and kits for performing the construction and screening steps are commercially available (for example, Stratagene Cloning Systems, La Jolla, CA). Once isolated, the nucleic acid can be directly cloned into an appropriate vector in an antisense orientation, or if necessary, be modified to facilitate the subsequent cloning steps. Such modification steps are routine, an example of which is the addition of oligonucleotide linkers which contain restriction sites to the termini of the nucleic acid. General methods are set forth in Sambrook et al. (39).

The DNA that is introduced into the cell is in an expression orientation that is antisense to a corresponding endogenous DNA or RNA of the cells. For example, where an endogenous DNA comprises a gene which encodes for a particular protein, the introduced DNA is in an expression orientation opposite the expression of the endogenous DNA, that is the DNA operatively linked to a promoter is in an antisense expression orientation relative to the corresponding endogenous gene. The introduced DNA may be homologous to the entire transcribed gene or homologous to only part of



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the transcribed gene. Alternatively, the sequence of the introduced DNA may be divergent to that of the endogenous DNA but only divergent to the extent that hybridization of the nucleic acids occurs, thereby preventing transcription. One skilled in the art can determine the maximum extent of this divergence by routine screening of antisense DNAs corresponding to an endogenous DNA of the cell. In this manner, one skilled in the art can readily determine which fragments, or alternatively the extent of homology of the fragments or the entire gene that is necessary to inhibit gene expression.

The antisense nucleic acids of the present invention can be made according to protocols standard in the art, as well as described in the Examples provided herein. The antisense nucleic acids can be administered to a subject according to the gene transduction protocols standard in the art, as described below.

The present invention also contemplates a method for stimulating the transcription modulating activity and/or histone acetyltransferase activity of P/CAF in a subject comprising administering to the subject a substance, in a pharmaceutically acceptable carrier, determined according to the methods taught herein, to have a -stimulatory affect on the transcription modulating and/or histone acetyltransferase activity of P/CAF. The substance can be one which has been identified, according to the protocols provided herein, to stimulate histone acetyltransferase activity in P/CAF or promote binding of P/CAF to p300/CBP. The stimulation of the transcription modulation activity and/or histone acetyltransferase activity of P/CAF in a subject is desirable, for example, to activate tumor suppressor p53 (which promotes apoptosis) or to activate the muscle differentiation factor, MyoD. Thus, the method of the present invention can be employed to treat cancer and to promote muscle differentiation in conditions where muscle differentiation is desired. The substance can be delivered to a cell in the subject by mechanisms well known in the art.

Further contemplated in the present invention is a method for promoting binding of P/CAF to p300/CBP in a subject, comprising administering to the subject a substance

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identified by the methods provided herein to promote binding of P/CAF to either p300 or CBP.

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Additionally, a nucleic acid encoding a protein which stimulates the transcription modulating activity and/or histone acetyltransferase activity of P/CAF can be delivered to a cell in the subject by gene transduction mechanisms, as described below.

Also provided in the present invention is a method of inhibiting the cell cycle progression inducing effect of an oncoprotein which binds p300/CBP in a subject comprising transducing the cells of the subject with a vector comprising a nucleic acid encoding the P/CAF protein, inducing expression of the nucleic acid in the cell to produce the P/CAF in an amount which will allow the P/CAF gene product to replace the oncoprotein bound to p300/CBP, whereby the replacement of the oncoprotein bound to p300/CBP by the P/CAF gene product inhibits the cell cycle progression inducing effect of the oncoprotein. The oncoprotein which binds p300/CBP in the cell can be the adenovirus E1A oncoprotein.

A method for providing a functional P/CAF protein to a subject in need of the functional P/CAF protein is also provided, comprising transducing the cells of the subject with a vector comprising a nucleic acid encoding the P/CAF protein and inducing expression of the nucleic acid to produce the functional P/CAF protein in the cell, thereby providing the functional P/CAF protein to the subject. The transduction of the vector nucleic acid into the subject's cells can be carried out according to standard gene therapy protocols well known in the art (see, for example, U.S. Patent No. 5,339,346).

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# Screening assays for p300/CBP

The present invention also provides a bioassay for screening substances for the ability to inhibit the histone acetyltransferase activity of p300/CBP comprising contacting a system, in which histone acetylation by p300/CBP can be determined, with the substance under conditions whereby histone acetylation by p300/CBP can occur,



determining the amount of histone acetylation by p300/CBP in the presence of the substance; and comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, a decreased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can inhibit the histone acetyltransferase activity of p300/CBP. The acetylation of histones by p300/CBP can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P300/CBP protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to inhibit the histone acetyltransferase activity of p300/CBP can be added to this system and assayed for acetyltransferase inhibiting ability.

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Also provided in the present invention is a bioassay for screening substances for the ability to inhibit the binding of a transcriptional factor to p300/CBP, comprising contacting a system in which the binding of a transcriptional factor to p300/CBP can be determined, with the substance under conditions whereby the binding of the transcriptional factor and p300/CBP can occur, determining the amount of transcriptional factor binding to p300/CBP in the presence of the substance, and comparing the amount of transcriptional factor binding to p300/CBP in the presence of the substance with the amount of transcriptional factor binding to p300/CBP in the absence of the substance, a decreased amount of transcriptional factor binding to p300/CBP in the presence of the substance indicating a substance that can inhibit the binding of a transcriptional factor to p300/CBP. The binding of a transcriptional factor to p300/CBP can be determined in a system, for example, which can include a cell free reaction mixture comprising a transcriptional factor which binds p300/CBP and p300/CBP. Alternatively, the system can comprise a cell extract produced from cells producing both a transcriptional factor which binds p300/CBP and p300/CBP. The transcriptional factor which binds p300/CBP can be selected from, but is not limited to



the group consisting of nuclear hormone receptors, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YYI, Sap-1a, c-Fos, MyoD and SRC-1, as well as any other transcriptional factor now known or later identified to bind p300/CBP. The screening assay of the present invention can also be used to identify substances which inhibit the binding of p300/CBP to other components to which it is known to bind, for example, P/CAF, pp90<sub>RSK</sub>, TFIIB, E1A, SV40 large T antigen, as well as any other substances now known or later identified to bind p300/CBP. Determination of the binding of a transcriptional factor or other substance to p300/CBP can be carried out as taught in the Examples herein as well as by protocols described in the literature.

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The present invention further contemplates a bioassay for screening substances for the ability to stimulate the histone acetyltransferase activity of p300/CBP comprising contacting a system, in which histone acetylation by p300/CBP can be determined, with the substance; determining the amount of histone acetylation by p300/CBP in the presence of the substance; and comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, an increased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can stimulate the histone acetyltransferase activity of p300/CBP. The acetylation of histones by p300/CBP can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the p300/CBP protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to stimulate the histone acetyltransferase activity of p300/CBP can be added to this system and assayed for stimulating ability.

The present invention further provides a bioassay for screening substances for the ability to stimulate binding of a component, which binds p300/CBP, to p300/CBP, comprising contacting a system in which the binding of the component to p300/CBP can



be determined, with the substance under conditions whereby the binding of the component to p300/CBP can occur; determining the amount of component binding to p300/CBP in the presence of the substance; and comparing the amount of component binding to p300/CBP in the presence of the substance with the amount of component binding to p300/CBP in the absence of the substance, an increased amount of component binding to p300/CBP in the presence of the substance indicating a substance that can stimulate the binding of the component to p300/CBP. The binding of the component to p300/CBP can be determined in a system, for example, which can include a cell free reaction mixture comprising the component and p300/CBP. Alternatively, the system can comprise a cell extract produced from cells producing both the component and p300/CBP. The component which binds p300/CBP can be any of the transcriptional factors or other proteins which are known or are identified in the future to bind p300/CBP, as set forth above. Determination of the binding of the component to p300/CBP can be carried out as taught in the Examples provided herein and according 15 to protocols available in the literature.

# Histone acetyltransferase activity of p300/CBP

A method for inhibiting the histone acetyltransferase activity of p300/CBP in a subject is provided in the present invention, comprising administering to the subject a histone acetyltransferase activity inhibiting amount of a substance in a pharmaceutically acceptable carrier. The mechanism of the inhibitory action of the substance can be the inhibition of the binding of a DNA-binding transcription factor, such as, for example, a nuclear hormone receptor, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YY1, Sap-1a, c-Fos, MyoD or SRC-1, to p300/CBP.

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The histone acetyltransferase activity of p300/CBP can be inhibited in a subject by administering to the subject a substance which binds p300/CBP at the transcription factor binding site or a substance which binds the transcription factor protein at the p300/CBP binding site, the ultimate result being that the transcription factor and p300/CBP do not bind with one another and p300/CBP cannot acetylate histones.

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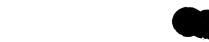


The substance which binds either to the transcription factor or the p300/CBP protein and has the net effect of inhibiting the histone acetyltransferase activity of p300/CBP in the cell can be identified according to the screening methods provided herein and delivered to a cell in the subject by mechanisms well known in the art. The substance can be a protein, such as an antibody which binds the p300/CBP protein binding site at or near the DNA-binding transcription factor binding site, thereby preventing its binding or an antibody which binds the DNA-binding transcription factor at or near the p300/CBP binding site, thereby preventing its binding. The substance can also bind the histone acetyltransferase site on p300/CBP (aa 1195-1673 on p300 or aa 1174-1850 on CBP) or at the acetylation site on the histone, thereby preventing acetylation by p300/CBP

Additionally, the substance can be a nucleic acid which can be expressed in the cell to produce a protein which inhibits the histone acetyltransferase activity of p300/CBP. For example, a nucleic acid encoding a protein which binds either to a transcription factor or the p300/CBP protein and has the net effect of inhibiting the histone acetyltransferase activity of p300/CBP in the cell can be delivered to a cell in the subject by gene transduction mechanisms well known in the art. For example, nucleic acid can be introduced by liposomes as well as via retroviral or adeno-associated viral vectors, as described below

The substance which inhibits the histone acetyltransferase activity of p300/CBP can be an antisense RNA or an antisense DNA which binds the RNA or DNA of p300/CBP thereby preventing translation or transcription of the RNA or DNA encoding p300/CBP and having the net effect of inhibiting the histone acetyltransferase activity of P/CAF by inhibiting p300/CBP production. The antisense RNA or DNA of the present invention can be produced and introduced into cells according to the same methods as set forth above for P/CAF antisense nucleic acids.

The present invention also contemplates a method for stimulating the histone acetyltransferase activity of p300/CBP in a subject comprising administering to the



subject a histone acetyltransferase activity stimulating amount of a substance, in a pharmaceutically acceptable carrier, determined according to the methods taught herein, to have a stimulatory affect on the histone acetyltransferase activity of p300/CBP. The substance can exert a stimulatory effect by promoting the binding of a DNA-binding transcription factor of the present invention to p300/CBP. The substance can be delivered to a cell in the subject by mechanisms well known in the art. A nucleic acid encoding a protein which stimulates the transcription modulating activity of p300/CBP can be delivered to a cell in the subject by gene transduction mechanisms, as described below.

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#### Gene transduction

In the methods described above which include gene transduction into cells (i.e., addition of exogenous DNA into cells), the nucleic acids of the present invention can be in a vector for delivering the nucleic acids to the site for expression of the P/CAF protein. The vector can be one of the commercially available preparations, such as the pGM plasmid (Promega). Vector delivery can be by liposome, using commercially available liposome preparations or newly developed liposomes having the features of the present liposomes. Additionally, vector delivery can be via a viral system, including, but not limited to, retroviral, adenoviral and adeno-associated viral systems. Other delivery methods can be adopted and routinely tested according to the methods taught herein.

The modes of administration of the liposome will vary predictably according to the disease being treated and the tissue being targeted. For example, for treating cancer in either the lung or the liver, which are both sinks for liposomes, intravenous delivery is reasonable. For other localized cancers, as well as precancerous conditions, catheterization of an artery upstream from the target organ is a preferred mode of delivery, because it avoids significant clearance of the liposome by the lung and liver. For cancerous lesions at a number of other sites (e.g., skin cancer, localized dysplasias), topical delivery is expected to be effective and may be preferred, because of its convenience.

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Leukemias and other disorders involving dysregulated proliferation of certain isolatable cell populations may be more readily treated by ex vivo administration of the nucleic acid.

The liposomes may be administered topically, parenterally (e.g., intravenously), by intramuscular injection, by intraperitoneal injection, transdermally, extracorporeally or the like, although intravenous or topical administration is typically preferred. The exact amount of the liposomes required will vary from subject to subject, depending on the species, age, weight and general condition of the subject, the severity of the disease being treated, the particular compound used, its mode of administration and the like. Thus, it is not possible to specify an exact amount. However, an appropriate amount may be determined by one of ordinary skill in the art using only routine experimentation given the teachings herein.

Parenteral administration, if used, is generally characterized by injection. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution of suspension in liquid prior to injection, or as emulsions. A more recently revised approach for parenteral administration involves use of a slow release or sustained release system such that a constant level of dosage is maintained. See, e.g., U.S. Patent No. 3,610,795, which is incorporated by reference herein.

Topical administration can be by creams, gels, suppositories and the like. Ex vivo (extracorporeal) delivery can be as typically used in other contexts.

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The present invention is more particularly described in the following examples which are intended as illustrative only since numerous modifications and variations therein will be apparent to those skilled in the art.

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#### **EXAMPLES**

#### I. P/CAF studies.

## 5 Cloning and characterization of P/CAF protein.

In human cells, CBP binds to c-Jun in a phosphorylation-dependent manner in association with stimulation of transcription (9). In yeast, GCN4 is believed to be a c-Jun counterpart on the basis of similarities in DNA recognition (15) as well as the participation of both proteins in UV signaling pathways (16). Yeast genetic screening has led to the isolation of various cofactors for GCN4, including GCN5 (yGCN5), ADA2 (yADA2) and ADA3 (yADA3) (17-19). These factors are considered to function as a complex (or in a common pathway) based on genetic and protein-protein interaction studies (18-22). Finally, p300/CBP and yADA2 exhibit significant sequence similarity within a 50 amino acid region including a Zn<sup>2+</sup> finger motif (3). Human counterparts to yGCN5, yADA2, or yADA3 that interact with p300/CBP to mediate transcriptional activation by c-Jun were searched for in various nucleotide sequence databases.

Comparison of the yGCN5 protein sequence with various databases (23) revealed significant similarities with the two randomly sequenced human cDNAs, ETS05039 (24) (P=4.0x10<sup>-15</sup>) and NIB2000-5R (P=6.5x10<sup>-9</sup>). Given that these cDNAs were truncated, human fetal liver and fetal brain cDNA libraries (Clontech) were screened with ETS05039 and NIB2000-5R, respectively and complete clones were isolated from the human fetal liver cDNA library. The complete sequences revealed that the ETS05039- and NIB2000-5R-derived clones are encoded by distinct genes but are highly related within the protein coding regions (68% identity at the DNA level, 75% identity and 86% similarity at the protein level). The former encodes an N-terminal region with no sequence similarity to any proteins in the databases besides the yGCN5-related C-terminal region, whereas the latter encodes only the yGCN5-related region. Given that p300/CBP-binding activity was observed in the former polypeptide as shown below, it was designated p300/CBP-associated factor (P/CAF), having the amino acid



sequence of SEQ ID NO:1 and the nucleotide sequence of SEQ ID NO:10 and the latter was named human GCN5 (hGCN5), having the amino acid sequence of SEQ ID NO:5 and the nucleotide sequence of SEQ ID NO:11.

Additionally, an RNA blot (Clontech) was hybridized with a random-primed probe made from the cDNA encoding P/CAF. RNA blotting indicated that transcripts detected by the P/CAF and hGCN5 cDNAs are ubiquitously expressed, but the former is most abundant in heart and skeletal muscle, whereas the latter is most abundant in pancreas and skeletal muscle.

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# P/CAF-p300/CBP interaction in vitro

The P/CAF binding site was presumed to reside in the C terminal one third of CBP (residues 1,678-2,442) because it was observed that this region, when fused to a DNA binding domain, activates transcription (4) in a manner repressed by coexpression of 12S E1A. This region was divided into 6 overlapping fragments and each was expressed in *E. coli* as a glutathione-S-transferase (GST) fusion protein. GST-CBP fusions were incubated with recombinant P/CAF protein and, subsequently, purified using glutathione-Sepharose. Co-purified P/CAF was detected by immunoblotting analysis.

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To construct GST-fusions, various regions of CBP and p300 were amplified by PCR. A series of deletions of the CBP segment B was created by site-directed *in vitro* mutagenesis (30). These fragments were subcloned into pGEX-2T (Pharmacia). GST-fusions were expressed in *E. coli* and extracted with buffer B [20 mM Tris-HCl (pH 8.0), 5 mM MgCl<sub>2</sub>, 10% glycerol, 1 mM AEBSF, 0.1% NP40, 10 μg/ml of aprotinin, 10 μg/ml of leupeptin, 1 μg/ml of pepstatin A, 1 mM DTT] containing 0.1 M KCl for these experiments. GST-CBP-segment B was purified by glutathione-Sepharose and phenyl-Sepharose chromatographic steps, P/CAF, hGCN5, and E1A were expressed as FLAG-fusions in Sf9 cells via baculovirus vectors and affinity-purified with M2-agarose (ref. 30, Kodak-IBI). For interaction, a crude *E. coli* extract containing 20 pmol of GST-fusion was incubated with 40-60 pmol of P/CAF or E1A in a total volume of 50 μl of

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buffer B with 0.1 M KCl on ice for 10 min. Samples were further incubated with 10 µl (packed volume) of glutathione-Sepharose at 4°C for 30 min, washed four times with 200 µl of buffer B containing 0.1 M KCl, and eluted with 20 µl of buffer E [50 mM Tris-HCl (pH 8.0), 0.2 M KCl, 20 mM glutathione] for 60 min. Interacting proteins were detected by anti-FLAG immunoblotting or silver staining.

For p300 interactions, the segment spanning residues 1763-1966 (segment B') of p300, which is analogous to the CBP segment-B, was used. Twenty percent of the P/CAF and hGCN5 inputs and 100% of the E1A input were also analyzed. In the GST precipitation assays, almost identical amounts of the GST fusions were recovered in all samples. Interaction between P/CAF and CBP (segment B) was determined in the absence and in the presence of E1A. Control reactions with GST-CBP alone and without GST-CBP were also performed. Input proteins were analyzed.

Two CBP segments, A and B, interacted specifically with P/CAF. The stronger interaction was observed in the latter segment, which does not include the yADA2-like Zn<sup>2+</sup> finger. Given that the CBP segment-B is well conserved in p300 (66% identity, 75% similarity), the binding of P/CAF to p300 *in vitro* was also analyzed. For this experiment, the p300 segment spanning residues 1763-1966, termed segment B', which is analogous to the CBP segment-B, was used. Like CBP, p300 interacted specifically with P/CAF. These studies demonstrated that P/CAF binds specifically to both p300 and CBP *in vitro*. In contrast to P/CAF, hGCN5 did not bind to CBP or p300.

These studies also demonstrated that the Zn<sup>2+</sup> finger region of p300/CBP, which shares sequence similarity with yADA2, is not essential for the interaction with P/CAF. Cloning of a human structural homolog of yADA2, termed hADA2 (25) has revealed that, unlike the sequence similarity between p300/CBP and yADA2, which is restricted to a 50 amino acid region, hADA2 shares extensive similarity (30% identity, 52% similarity) to yADA2 over the entire protein sequence. Moreover, a computer search of the complete genomic sequence of *Saccharomyces cerevisiae* revealed that yeast does

not have counterparts of p300/CBP or P/CAF. Thus, the p300/CBP-P/CAF pathway may have been acquired during metazoan evolution.

### 5 Action of E1A in vitro

Previous reports indicated that E1A binds to both the p300 segment spanning residues 1767-1816 and the CBP segment spanning residues 1805-1854 (7). These interactions were reconfirmed in the present system, thus, both p300 and CBP segments covering the previously identified regions interacted with E1A.

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For further mapping, a series of deletions was introduced within the CBP segment-B and tested for interactions with P/CAF and E1A. Deletions of residues 1801-1825 or 1824-1851 markedly reduced interactions with both P/CAF and E1A, whereas deletion of residues 1850-1878 did not affect these interactions. Furthermore, deletion of residues 1801-1851 completely abolished interactions with both P/CAF and E1A. These data indicate that residues 1801-1851 of CBP are critical for interaction with both P/CAF and E1A. Taken together with the evidence that CBP segment A (aa residues 1,678-1,880) also binds to these factors, the above findings demonstrate that P/CAF and E1A bind to the same or very closely spaced sites on CBP.

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Evidence that both P/CAF and E1A recognize the same p300/CBP segments raises the possibility of direct competition between P/CAF and E1A for binding to p300/CBP. To test this possibility, a competition experiment was performed with the use of affinity purified recombinant proteins. The interaction of P/CAF with the CBP-segment B was progressively inhibited by the addition of increasing amounts of E1A. In contrast, no inhibition was caused by an E1A mutant which does not bind to p300/CBP (E1AAN). Similar results were obtained with the p300-segment B', leading to the conclusion that P/CAF and E1A compete for the same binding sites in p300/CBP.

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### P/CAF-p300/CBP interaction in vivo

The *in vivo* interaction between P/CAF and p300/CBP was established by co-immunoprecipitation from a human osteosarcoma cell extract. Proteins in this extract were immunoprecipitated with rabbit anti-P/CAF, rabbit anti-CBP and anti-p300 antibodies. For controls, cell extract was precipitated with rabbit control IgG or mouse anti-HA monoclonal antibody. The precipitates were analyzed by immunoblotting with anti-P/CAF, anti-CBP and anti-p300 antibodies.

Osteosarcoma cells were transfected with either control vector or E1A- or

E1AAN-expression vectors. Extract from the transfected subpopulation was
immunoprecipitated with anti-P/CAF or control IgG. The precipitates were analyzed by
immunoblotting with anti-p300 and anti-P/CAF antibodies.

Rabbit anti-P/CAF antibody was raised to the P/CAF segment spanning residues 125-397 and purified by immunoaffinity chromatography (33). A mixture of 15 monoclonal antibodies raised to the human p300 segment spanning residues 1572-2371 (5) and rabbit polyclonal antibodies raised to the mouse CBP segment spanning residues 2-23 (for immunoprecipitation) and 1736-2179 (immunoblotting) were purchased from Upstate Biotechnology. Approximately 2 x 10<sup>7</sup> human osteosarcoma U-2 OS cells (ATCC accession number HTB 96) were extracted with 10 ml of lysis buffer [25 mM 20 HEPES-KOH (pH 7.2), 150 mM potassium acetate, 2 mM EDTA, 1 mM DTT, 1 mM AEBSF, 10 μg/ml of aprotinin, 10 μg/ml of leupeptin, 1 μg/ml of pepstatin A, 20 mM sodium fluoride, 0.1% NP40]. Two to 10 ml of extract were incubated with 2 μg of the respective antibody for four hours at 4°C. Fifty µl (packed volume) of protein-A 25 Trisacryl (Pierce) were added and incubation was continued for two hours. The matrix was washed four times with 1 ml of the lysis buffer, then boiled in 2x SDS sample buffer. Human osteosarcoma U-2 OS cells were transfected with 20 µg of the indicated plasmid and 1 µg of sorting plasmid (pCMV-IL2R) (31). The transfected subpopulation was purified by magnetic affinity cell sorting (32). Extract from approximately 2 x 10<sup>5</sup> sorted cells was immunoprecipitated as described. 30

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Anti-P/CAF antibody specifically detected a 95 kDa protein, which is very close to the calculated value for the full-length P/CAF, in the immunoprecipitates. Anti-P/CAF antibody co-immunoprecipitated both CBP and p300. Similarly, anti-CBP antibody also co-immunoprecipitated P/CAF. However, anti-p300 antibody did not co-immunoprecipitate P/CAF. This is most likely due to steric interference since the anti-p300 antibody was raised to the p300 segment spanning residues 1572-2371 which includes the P/CAF binding region. These data demonstrate that P/CAF forms complexes with both p300 and CBP in vivo.

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#### 10 Action of E1A in vivo

The *in vitro* experiments described herein indicate that P/CAF and E1A compete for the binding sites in p300/CBP. Thus, a study was conducted to determine whether E1A targets the endogenous interaction between P/CAF and p300. An E1A-expression vector was transiently transfected into human osteosarcoma cells and the transfected subpopulation was purified by cell sorting. Then, the interaction between P/CAF and p300 in transfected cells was examined by co-immunoprecipitation with anti-P/CAF antibody. The endogenous interaction of P/CAF with p300 was drastically inhibited by expression of E1A. On the other hand, no inhibition was observed by the E1A mutant lacking the p300 binding domain (E1AΔN), indicating that E1A disrupts the P/CAF-p300 complex *in vivo* through an interaction with p300.

### Cell cycle regulation by P/CAF

Given that binding of P/CAF to p300/CBP is inhibited by E1A, experiments were performed to evaluate whether P/CAF, by binding to and forming a functional complex with p300, is involved in the regulation of entry into S phase. This possibility was addressed by examining whether transient expression of P/CAF would affect the rate of G1/S transit in HeLa cells. P/CAF negatively affected the distribution of cells between G1 and S phases in this assay.

HeLa cells were transfected by electroporation with 7  $\mu$ g of P/CAF-expression plasmid and/or 3  $\mu$ g of the full-length or the N-terminally deleted ( $\Delta 2$ -36) E1A 12S-

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expression plasmid as indicated. These plasmids were constructed by subcloning FLAG-P/CAF and E1A cDNAs into pCX (34) and pcDNAI (Invitrogen), respectively. All samples, in addition, contained 1 µg of sorting plasmid (pCMV-IL2R) (31) and carrier plasmid (pCX) to normalize the total amount of DNA to 11 µg. After transfection, cells were incubated in Dulbecco's modified Eagle's medium with 10% fetal bovine calf serum for 12 h, and subsequently labeled in medium containing 10 µM bromo-deoxyuridine (BrdU) for 30 min. Subsequently, the transfected subpopulation was purified by magnetic affinity cell sorting and nuclei were analyzed by dual parameter flow cytometry as described (32).

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The fraction of cells accumulating in S phase in control cultures was 23%, compared to 15% in P/CAF-transfected cells. This effect was reproducible in multiple independent experiments. In parallel experiments to verify the utility of this experimental protocol, plasmids encoding E2F-1, simian virus 40 small t, cyclin A or cyclin E increased the accumulation of cells in S phase, whereas plasmids encoding the cyclin-dependent kinase inhibitors p21 or p27 reduced the number of S phase cells.

On the basis of evidence that E1A and P/CAF compete for binding sites on p300, it seemed possible that cotransfection of P/CAF with E1A would oppose the mitogenic effect caused by E1A. As shown by the data herein, this is indeed the case. E1A alone has mitogenic activity in this experimental setting, while the E1A mutant lacking the p300 binding domain (E1ADN) has very weak activity. Comparable expression levels between wild type and mutant E1A in the transfected cells were revealed by immunoblotting analysis with anti-E1A. Intriguingly, when P/CAF was cotransfected with E1A, the mitogenic activity of E1A was significantly counteracted by P/CAF. These results show that P/CAF and E1A mediate antagonistic effects on cell cycle progression.

In the course of assessing P/CAF activity, it was also revealed that p300 is able to inhibit cell cycle progression under the same assay conditions. These findings suggest

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that P/CAF and p300, perhaps by forming a complex, act in concert to suppress cell cycle progression.

# Histone acetyltransferase activity in P/CAF

Acetylation of the N-terminal histone tails has been considered to play a crucial role in accessibility of transcription factors to nucleosomal templates (26-27). Recently, yGCN5 has been identified as a histone acetyltransferase (28). On the basis of this information, intrinsic histone acetyltransferase activity in P/CAF and hGCN5 was examined. As substrates, the core histones (histones H2A, H2B, H3 and H4) and the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) were used.

Activity of hGCN5 and P/CAF that acetylates free histones or histones in the nucleosome core particle (35) was measured as described (36). Each reaction contained 0.3 pmol of affinity purified FLAG-hGCN5 or FLAG-P/CAF, 4 pmol of the histone octamer or the nucleosome core particle and 10 pmol of [1-14C]acetyl-CoA. The histone octamer dissociated into dimers or tetramers under assay conditions. Acetylated histones were detected by autoradiography after separation by SDS-PAGE.

P/CAF and hGCN5 acetylated the core histones with almost the same efficiency. Both factors acetylated histones H3 and H4, but preferentially H3. In contrast, very weak or no acetylation by hGCN5 was detected in the nucleosome core particles. Remarkably, significant acetylation by P/CAF was observed in a nucleosomal context. Although all core histones are acetylated in the nucleus, P/CAF and hGCN5 did not acetylate histones H2A and H2B *in vitro*.

Direct function of P/CAF is likely to involve its intrinsic histone acetyltransferase activity. Although exact molecular mechanisms by which acetylation of core histones contribute to transcription remains undefined, acetylation of the histones is considered to play an important role in transcriptional regulation (26-27). The positively charged N-terminal tails of core histones are believed to affect nucleosome structure by interacting



with DNA at or near the nucleosome-spacer junction. Acetylation of the histone tails presumably destabilizes the nucleosome and facilitates access by regulatory factors. Likewise, there is a general correlation between the level of acetylation and transcriptional activity of nucleosomal domains. The findings of the present invention provide insights into the mechanisms of targeted histone acetylation.

Cellular factor p300/CBP binds to various sequence-specific factors that are involved in cell growth and/or differentiation, including CREB (3,4), c-Jun (9), Fos (11), c-Myb (12) and nuclear receptors (13). P/CAF could stimulate the activation function of these factors via promoter-specific histone acetylation. The present invention demonstrates that E1A appears to perturb normal cellular regulation by disrupting the connection between p300/CBP and its associated histone acetyltransferase.

#### II. p300/CBP studies.

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## Purification of E1A associated histone acetyltransferase.

FLAG-epitope tagged E1A (or ΔE1A) was expressed in Sf9 cells (ATCC accession number CRL 1711) by infecting recombinant baculovirus (43). All purification steps were carried out at 4°C. Extract was prepared from infected cells by one cycle of freeze and thaw in buffer B (20 mM Tris-HCl, pH 8.0; 5 mM MgCl<sub>2</sub>, 10% glycerol; 1 mM PMSF; 10 mMβ-mercaptoethanol, 0.1% Tween 20) containing 0.1 M KCl and the complete protease inhibitor cocktail (Boehringer Mannheim). To prepare E1A-immobilized beads, the extract was incubated with M2 anti-FLAG antibody agarose (Kodak-IBI) for four hours with rotating and subsequently washed with the same buffer three times. The resulting beads were incubated with HeLa (ATCC accession number CCL 2) nuclear extract for four to eight hours and thereafter washed with the same buffer six times. Finally, FLAG-E1A was eluted from the beads along with associated polypeptides by incubating with the same buffer containing 0.1 mg/ml FLAG peptide.

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For further purification, eluted polypeptides were dialyzed in 0.05 M KCl-buffer B and subsequently loaded onto a SMART Mono Q column (Pharmacia) equilibrated with the same 0.05 M KCl-buffer B. After washing, the column was developed with a linear gradient of 0.05-1.0 M KCl in buffer B. Mono Q fractions were concentrated with a MICROCON spin-filter (Amicon) and consequently loaded onto a SMART Superdex 200 column (Pharmacia) equilibrated with 0.1 M KCl-buffer B.

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# Histone acetyltransferase assays

Filter binding assays were performed as described (80) with minor modifications. Samples were incubated at 30°C for 10-60 minutes in 30 ml of assay buffer containing 50 mM Tris-HCl, pH 8.0, 10% glycerol; 1 mM DTT, 1 mM PMSF; 10 mM sodium butyrate, 6 pmol of [3H]acetyl CoA (4.3 mCi/mmole, Amersham Life Science Inc.), and 33 mg/ml of calf thymus histones (Sigma Chemical Co.). In experiments where synthetic peptides were substituted for core histones, 50 pmol of each peptide were used. After incubation, the reaction mixture was spotted onto Whatman P-81 phosphocellulose filter 15 paper and washed for 30 minutes with 0.2 M sodium carbonate buffer pH 9.2 at room temperature with 2-3 changes of the buffer. The dried filters were counted in a liquid scintillation counter.

PAGE analysis was done as above except that 90 pmol of [14C]acetyl CoA (55 mCi/mmole, Amersham Life Science Inc.) and 9 pmol of core histones or mononucleosomes were used. Core histones and mononucleosomes were prepared as described (35). For trypsin digestion, reaction mixtures were further incubated with various amounts of trypsin on ice for 30 minutes. The samples were analyzed on one dimensional SDS-PAGE gels or two dimensional gels, where the first dimension was an acid-urea-PAGE gel (44) and the second dimension was an SDS-PAGE gel.

## Protein expression

For baculovirus expression, cDNAs corresponding to p300 portions of aa 1-670, aa 671-1194 and aa 1135-2414 were amplified by PCR (EXPAND High Fidelity PCR System; Boehringer Mannheim) as KpnI-NotI fragments. The resulting fragments were

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subcloned into a baculovirus transfer vector having the FLAG-tag sequence (43). The recombinant viruses were isolated using the BACULOGOLD system (Pharmingen), according to the manufacturer's protocol and were infected into Sf9 cells (ATCC accession number CRL 1711) to express FLAG-p300. Recombinant proteins were affinity purified with M2 anti-FLAG antibody-immobilized agarose (Kodak-IBI) according to the manufacturer's protocol.

For bacterial expression, cDNAs encoding the p300 portions and the CBP portion (aa 1174-1850) were first subcloned into the baculovirus transfer vector having the FLAG-tag as described above. Thereafter, the XhoI and NotI fragments encoding FLAG-p300 or FLAG-CBP fusions were resubcloned into the *E. coli* expression vector pET-28c (Novagene) digested with SalI and NotI. Recombinant proteins were expressed in *E. coli* BL21(DE3) and affinity purified with M2-antibody agarose.

#### 15 Histone acetyltransferases that associate with E1A

Although the adenovirus E1A 12S protein (E1A) inhibits transcription in a variety of genes via direct binding to p300/CBP (45), E1A also stimulates transcription in some contexts (46). Thus, p300/CBP-bound E1A was tested to determine whether it might recruit histone acetyltransferases or deacetylases to regulate transcription. In addition, experiments were conducted as described below to determine if p300/CBP per se is a histone acetyltransferase.

Initially, recombinant FLAG-epitope tagged E1A was immobilized on anti-FLAG antibody beads. Immobilized E1A was incubated with a HeLa nuclear extract for affinity purification of E1A-associated polypeptides. FLAG-E1A was then eluted from the beads, along with E1A-associated polypeptides, by incubating with FLAG-peptide. Although E1A per se has no histone acetyltransferase activity, E1A recruited significant amounts of histone acetyltransferase activity from the nuclear extract. It is very unlikely that this activity is derived from P/CAF given that E1A and P/CAF cannot bind to p300/CBP simultaneously (43). Consistent with this, no P/CAF was detected in these fractions by immunoblotting.



The E1A N-terminus, a region that is not highly conserved among the various adenovirus serotypes, is involved in p300/CBP binding *in vivo*. Mutations in the N-terminal region lead to loss of the ability for p300/CBP binding without affecting RB binding (1,47). Thus, the requirement of the E1A N-terminal region for the recruitment of histone acetyltransferase activity was tested. In contrast to the wild type, the N-terminal deleted form of E1A ( $\Delta$ N-E1A) recruited only a background level of acetyltransferase activity. In agreement with previous reports (47), the  $\Delta$ N-E1A showed no ability to interact with p300/CBP, although it still retained the ability to interact with a variety of other polypeptides, including RB

To define the relationship between p300/CBP and histone acetyltransferase activity, affinity purified E1A-binding polypeptides were separated by Mono Q ion-exchange column. Both p300/CBP and the acetyltransferase activity were coeluted at 140 mM KCl, while most of polypeptides were eluted at 260 mM KCl. The active fraction of Mono Q column (~140 mM KCl) was further separated by Superdex-200 gel filtration column. Both p300/CBP and the acetyltransferase activity coeluted after the void volume, indicating that p300/CBP is involved in the histone acetyltransferase activity.

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#### p300 is a histone acetyltransferase

The data provided herein indicate that p300 per se, or a polypeptide(s) associated with p300, possesses histone acetyltransferase activity. To test the former possibility, the acetyltransferase activity of recombinant p300 was measured. p300 was divided into three fragments, each of which was expressed in and purified from Sf9 cells via a baculovirus expression vector. Histone acetyltransferase activity was readily detected in the C-terminal fragment containing amino acids 1135-2414, whereas no activity was found in the other fragments, demonstrating conclusively that p300 per se is a histone acetyltransferase.

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### p300/CBP-histone acetyltransferase domain

To map the histone acetyltransferase domain of p300, a series of deletions was prepared. Given the poor conservation of the glutamine-rich region (aa 1815-2414) in the *C. elegans* p300/CBP homolog (6), the p300 fragment encoding aa 1135-1810 was expressed in and purified from *E. coli*. Importantly, this candidate region of p300 (aa 1135-1810) showed significant histone acetyltransferase activity. For further mapping within this region, a series of N-terminal deletions was constructed. Deletion of 60 residues, resulting in a fragment containing aa 1195-1810, had no effect on the acetyltransferase activity, whereas the deletion of 185 residues, yielding a fragment comprising aa residues 1320-1810, completely eliminated the acetyltransferase activity

Next, a series of C-terminal deletions was analyzed to determine the requirement of the P/CAF (or E1A) -binding domain. The p300 fragments lacking the E1A binding domain (aa 1195-1760, 1195-1706 and 1195-1673) still retained the acetyltransferase activity, whereas the further truncated mutant (aa 1195-1652) completely lost the acetyltransferase activity. Consistent with these results, the internal deletion of residues 1418-1720 showed no acetyltransferase activity. These data demonstrate that the histone acetyltransferase domain is located between the bromodomain and the -E1A-binding domain. Given that the histone acetyltransferase domain is highly conserved between p300 and CBP (91% similarity), the corresponding region of CBP, aa residues 1174-1850, was expressed to confirm the acetyltransferase activity. As expected, comparable activity was detected, indicating that both p300 and CBP are histone acetyltransferases.

Among various acetyltransferases including histone acetyltransferases GCN5 and P/CAF, putative acetyl-CoA binding sites are conserved (48). However, multiple alignment analysis (49) showed that the p300/CBP histone acetyltransferase domain does not belong to this group. Moreover, comparison of the p300/CBP histone acetyltransferase domain with peptide sequence databases (23) showed no sequence similarity to any other proteins. Accordingly, this invention shows that p300/CBP represents a novel class of acetyltransferases in that it does not have the conserved motif found among previously described acetyltransferases (48).



# p300 acetylates all core histones in mononucleosomes

Substrate specificity for acetylation by p300 was also examined. As substrates, histone octamers and mononucleosomes (146 base pairs of DNA wrapped around the octamer of core histones) were used. Given that the histone octamer dissociates into dimers or tetramers under physiological conditions, the histone octamer is referred to here as core histones. When core histones were used, p300 acetylated all four proteins, but preferentially H3 and H4. More importantly, in a nucleosomal context, p300 acetylated all four core histones nearly stoichiometrically. In contrast, p300 acetylated neither BSA nor lysozyme.

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Hyperacetylated histones are believed to be linked with transcriptionally active chromatin (26,27,50,51). Hyperacetylated forms are found in histones H4, H3 and H2B, which have multiple acetylation sites *in vivo*. Thus, the level of acetylation by p300 was also tested.

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Mononucleosomes treated with p300 were analyzed by two-dimensional gel electrophoresis. A Coomassie blue-stained gel and the corresponding autoradiogram showed that a significant amount of histones, especially H4, were hyperacetylated. Importantly, acetylation levels by p300 were very close to those of hyperacetylated histones prepared from HeLa nuclei treated with sodium butyrate, a histone deacetylase inhibitor. In contrast, no acetylated forms were detected in the reaction without p300. These results indicate that p300 acetylates histones in mononucleosomes to the hyperacetylated state by targeting multiple lysine residues.

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p300 acetylates the four lysines in the histone H4 N-terminal tail in vitro which are acetylated in vivo

Lysines at positions 5, 8, 12 and 16 of histone H4 are acetylated in vivo (51). Recent studies with yeast histone acetyltransferases demonstrate the position-specific acetylation by distinct acetyltransferases, i.e., while cytoplasmic acetyltransferases for histone deposition and chromatin assembly modify



positions 5 and 12, GCN5 modifies positions 8 and 16 (52). Accordingly, the positions of acetylation by p300 were also determined. A series of synthetic peptides containing acetylated lysines at various positions was used to determine the acetylation site-specificity of p300. Consistent with the two-dimensional gel electrophoresis analysis, the experiments with peptide substrates showed that p300 acetylates all four lysines in the histone H4 that are acetylated *in vivo*. These results are consistent with the view that deposition-related diacetylated histones are deacetylated during maturation of chromatin (53).

## p300 preferentially acetylates the N-terminal histone tail

Histone acetyltransferases modify specific lysine residues in the N-terminal tail of core histones but not the C-terminal globular domain in vivo (26,27,50,51). Structural models of nucleosomes (54,55,56) suggest that most of the lysine residues in the C-terminal globular domain are buried. Therefore, experiments were conducted to examine whether restricted acetylation of the N-terminal tail resulted from the substrate specificity of the enzyme or inaccessibility of the enzyme to the core domain in nucleosomes. The globular domains of all core histones contain a long helix flanked on either side by a loop segment and short helix, termed the "histone fold" (54,55,56). The histone fold is involved in formation of the stable H2A-H2B and H3-H4 hetero-dimers, consisting of extensive hydrophobic contacts between the paired molecules. Therefore, it is likely that a histone monomer cannot fold properly, thereby increasing access of the histone acetyltransferase to the core domain. Based on these considerations, experiments were conducted to determine whether p300 acetylates free histone H4 in a N-terminal-specific manner.

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Histone H4 was acetylated with p300 and subsequently the histone tail was removed by partial digestion with trypsin. The distributions of radioactivity between intact and core histones were compared. While the globular core histone domain was predominant at the higher trypsin concentrations, radioactivity was detected mostly in the intact histone. These data demonstrate that p300 preferentially acetylates the N-terminal tail of histone H4.

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# III. P/CAF interaction with MyoD

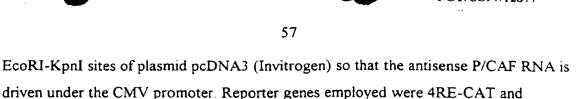
# Tissue culture and transfection experiments

C<sub>2</sub>C<sub>12</sub> mouse cells (ATCC accession number CRL 1772) were grown in Dulbecco's modified Eagle medium (DMEM) supplemented with 20% fetal bovine serum (FBS) until they reached confluence. Differentiation was induced by switching medium to differentiation medium (DM), consisting of DMEM containing 2% horse serum. C<sub>3</sub>H/10T1/2 fibroblasts (ATCC accession number CCL 226) were grown in DMEM supplemented with 10% FBS. Cells were transfected by the calcium phosphate precipitation method. Total amounts of transfected DNA were equalized by empty vector DNA. After 12 h incubation in medium containing the precipitated DNA, the cells were washed and incubated in fresh DMEM containing 10% FBS for an additional 24 h. Afterwards, differentiation was induced by incubating in DM for 36 to 72 h. Chloramphenicol acetyltransferase (CAT) assays were performed as previously described (64,69). The quantities of cell extracts used for CAT assays were normalized  $to\beta$ -galactosidase activity by cotransfection of 1 mg of the  $\beta$ -galactosidase expression vector, pON260.

Expression vectors used for transfection experiments are as follows: pCX-P/CAF for P/CAF (43), pCMV-bp300 for p300 (65), pCMV-p300 (1869-2414) (64) and pCMV-p300 (1514-1922) (60) for p300 wild type and mutants; pE1A12S, pE1A12S R2G, pE1A12S D2-36 and pE1A12S D121-130 for E1A wild type and mutants (66,67,68), and pEMSV-MyoD for MyoD (64)

The antisense P/CAF RNA expression vector, pcDNA3 P/CAF-AS, was created as follows. The 2.5 Kb EcoRI-KpnI fragment containing the entire P/CAF open reading frame was isolated from pCX-P/CAF (43). This fragment was subcloned into the





MCK-CAT (69). 4RE-CAT is driven by a synthetic promoter containing 4 copies of the E-box, whereas MCK-CAT is driven by the native MCK promoter (nucleotides -1256 to

5 +7).

### Microinjection and immunofluorescence

Cells were grown on small glass slides, subdivided into numbered squares of 2 mm x 2 mm and microinjected with purified and concentrated antibodies, as previously described (70). For immunofluorescence, cells were fixed in either 2% 10 paraformaldehyde or 1:2 methanol/acetone solution, preincubated with 5% BSA/PBS and incubated with the primary antibodies for 30 min at 37° C. Subsequently, antibody was visualized by incubating with either rhodamine- or fluorescein-conjugated secondary antibody for 30 min at 37° C. Injected antibodies were stained with a rhodamine-conjugated secondary antibody and nuclei were counter-stained by DAPI as previously described (69).

Antibodies employed are as follows; rabbit polyclonal affinity purified anti-P/CAF antibody (43), rabbit polyclonal anti-p300/CBP antiserum (71), mouse monoclonal anti-MyoD antibody (clone 5.8A, kindly provided by P. Houghton), goat polyclonal anti-c-Jun affinity purified antibody (Santa Cruz) and rabbit pre-immune serum.

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### Immunoprecipitation and DNA affinity purification

Cells were resuspended in lysis buffer (20 mM NaPO<sub>4</sub>, 150 mM NaCl, 5mM MgCl<sub>2</sub>, 0.1% NP40, 1 mM DTT, 10 mM sodium fluoride, 0.1 mM sodium vanadate, 1 mM phenylmethylsulfonyl-fluoride and 10 mg/ml each of leupeptin, aprotinin and pepstatin). After 30 min incubation on ice, samples were centrifuged at 12,000 x g for 30 min and supernatants were used as cell extracts. Extracts were pre-cleared by



incubating with rabbit pre-immune serum and protein A/G Plus-Agarose (Santa Cruz) for 2 h at 4 C. For immunoprecipitation, the supernatants were incubated with the respective antibodies for 3 h at 4 C. Protein A/G Plus-Agarose was added, and incubation continued for 3 h. The matrix was washed with lysis buffer, then boiled in 2 X SDS sample buffer. Immunoblotting was performed by using the ECL chemiluminescent detection kit (Amersham) according to the manufacturer's protocol.

Affinity purification of E-box-bound complexes was done as previously described (69). Briefly, 100 ng of the biotinylated double stranded DNA containing the E-box were immobilized on streptavidin-conjugated magnetic beads and incubated with 500 mg of cell extracts in the presence of poly dI-dC. After extensive washing, bound proteins were eluted with SDS sample buffer and analyzed by immunoblotting.

# In vitro protein-protein interaction assays

The CBP-B fragment and its deletion derivatives were expressed as 15 GST-fusions described previously (43). MyoD and E1A (43) were expressed as FLAG-fusion proteins in Sf9 cells via a baculovirus expression system and affinity-purified on M2 anti-FLAG antibody-agarose (Kodak-IBI). Crude E. coli extracts containing GST-fusions were incubated with various amounts of MyoD and/or E1A in 50 ml of buffer B (20 mM Tris-HCl, pH 8 0, 0 1 M KCl, 5 mM MgCl<sub>2</sub>, 10% 20 glycerol, and 0.1% Nonidet P-40) on ice for 10 min. GST-precipitation was performed as described (43). MyoD and E1A were detected by immunoblotting with anti-FLAG M2 antibody. For the interaction between P/CAF and MyoD, 1.5 pmol of FLAG-P/CAF and 15 pmol of FLAG-MyoD were incubated in 50 ml of buffer B on ice for 10 min. The mixture was further incubated with 2 mg of anti-P/CAF (43) or anti-hADA2 antibody for 60 min. The immunocomplexes were precipitated by incubation with 10 ml of protein A-Trisacryl (Pierce) and rotated for 1-4 hr at 4oC. The matrix was washed 4 times with 200 ml of buffer B and boiled in 10 ml of 2 X SDS sample buffer. The proteins were resolved on a 4%-20% gradient SDS-PAGE and subjected to immunoblotting with the anti-FLAG M2 antibody. The blot was developed 30 with the SUPERSIGNAL chemiluminescent substrates (Pierce).

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### P/CAF coactivates muscle-specific transcription

P/CAF and MyoD were co-transfected into mouse C3H10T1/2 fibroblasts, and MyoD-mediated transcription was determined from reporter activity driven by the artificial (4RE) and the naturally-occurring muscle creatine kinase (MCK) promoters. Overexpression of P/CAF stimulated MyoD-dependent transcription several folds in both promoters. Similar results were obtained for the myoD activated myogenin promoter Transcriptional activation was further stimulated by co-transfecting with MyoD, P/CAF and p300 expression vectors, suggesting that P/CAF may function by forming a complex with p300/CBP. Consistent with the lack of DNA binding capacity in P/CAF, overexpression of P/CAF alone did not increase the basal transcriptional activity of either enhancer. To test whether P/CAF and p300/CBP function in the same pathway, two dominant negative forms of p300 were employed which specifically inhibit p300/CBP-mediated transcription (60,64). The p300 segment spanning residues 1514-1922 inhibits the MyoD-dependent activation via direct interaction with MyoD (60), whereas the p300 segment spanning residues 1869-2414 inhibit it without direct interaction (64). Both dominant negative mutants inhibited MyoD-coactivation by P/CAF), suggesting that P/CAF and p300/CBP function in the same pathway.

For further elucidation of the activation mechanism by P/CAF, the effect of E1A, which inhibits MyoD-dependent transcription and differentiation (66,72,73) via direct interaction with p300/CBP (65,78), was tested. Expression of E1A in C3H10T1/2 fibroblasts inhibited stimulation of MyoD-directed transcription by P/CAF overexpression. E1A mutants lacking p300/CBP-binding activity, E1A D2-36 and E1A R2G (67,79), had almost no effect. On the other hand, an E1A mutant retaining p300/CBP-binding activity, E1A D121-130, behaved like the wild type. Since E1A associates with p300/CBP, but not with P/CAF, these results suggest that P/CAF functions in MyoD-directed transcription via interaction with p300/CBP.

To address the role of P/CAF as a myogenic coactivator in a more relevant environment, P/CAF was overexpressed in proliferating C2C12 myoblasts which express

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endogenous myogenic bHLH factors. As observed in fibroblasts, overexpression of P/CAF stimulated muscle specific transcription. Concomitant expression of exogenous p300 increased P/CAF-mediated coactivation. The repression exerted by wild type E1A, but not mutant E1A D2-36, on P/CAF coactivation of MyoD was also observed in muscle cells.

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Similar experiments were performed with myogenic cell lines that were stably transformed with wild type or mutant E1A-expressing vectors (66). Coactivation by P/CAF was inhibited by wild type E1A or the E1A mutant that retains

10 p300/CBP-binding activity (E1A\Delta121-130). In contrast, E1A mutants that lack p300/CBP-binding (E1A \Delta2-36 and E1A R2G) allowed transcriptional coactivation by P/CAF. Taken together, these experiments show that P/CAF coactivates MyoD-directed transcription via interaction with p300/CBP.

# 15 P/CAF stimulates myogenic differentiation

Given that P/CAF potentiates MyoD-directed transcription, the ability of P/CAF to assist MyoD in promoting myogenic differentiation was investigated. To this aim, C3H10T1/2 fibroblasts were transiently transfected with P/CAF and MyoD expression vectors. An expression vector for the green fluorescent protein (GFP) was co-transfected to identify transfected cells. After incubation in differentiation medium, the myogenic conversion of transfected cells was determined by simultaneous expression of the GFP and the differentiation-specific marker myosin heavy chain (MHC). Forced expression of MyoD in fibroblasts caused muscle differentiation in 12% of the transfected fibroblasts. This myogenic conversion was 20% by co-expressing MyoD and P/CAF. As observed in transcription experiments, stimulation of differentiation by P/CAF was counteracted by co-transfection with the p300 dominant negative mutant, p300 (1869-2414). Consistent with a general role for coactivators, overexpression of P/CAF alone was unable to differentiate fibroblasts.

Similar experiments were done using proliferating C2C12 myoblasts in which the differentiation program is already committed. Most of the myoblasts differentiated into

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myotubes by overexpressing P/CAF, whereas only a modest effect was observed by overexpressing p300. In contrast, differentiation was inhibited slightly by overexpressing c-Jun. This inhibitory effect presumably was caused by titration of p300/CBP, which associates directly with c-Jun (74). A similar inhibition was observed in the p300 dominant negative mutant. Consistent with the transcriptional effect, E1A almost completely inhibited differentiation. The E1A mutant RG2, lacking p300/CBP-binding capability but retaining the retinoblastoma protein (Rb)-binding capability, only partially inhibited differentiation, although this same mutant inhibited transcription as severely as the wild type. Taken together, these data show that P/CAF stimulates muscle differentiation by coactivating MyoD function via p300/CBP association.

### P/CAF is essential for myogenic transcription and differentiation

To test the necessity of P/CAF for myogenic transcription, experiments were conducted whereby P/CAF synthesis was inhibited by expressing antisense P/CAF RNA. A vector from which the P/CAF mRNA is transcribed in the antisense orientation (P/CAF-AS) was transfected with P/CAF and MyoD expression vectors into fibroblasts and MyoD-dependent transcription was examined. Cotransfection of the antisense expression vector strongly inhibited MyoD-dependent transcription below the level of induction elucidated by MyoD alone, demonstrating that expression of P/CAF antisense RNA inhibits not only the coactivation exerted by exogenous P/CAF but also that of endogenous P/CAF. These results indicate that P/CAF is essential for MyoD-dependent transcription.

Studies were also carried out to determine whether expression of P/CAF antisense RNA inhibits myogenic differentiation. C3H10T1/2 fibroblasts were transiently transfected with various expression vectors with or without the P/CAF antisense RNA expression vector. Expression of P/CAF antisense RNA reduced MyoD-mediated myogenic conversion of fibroblasts. Expression of P/CAF antisense RNA also counteracted the stimulatory effect of both P/CAF and p300 on myogenic differentiation. These data support the view that P/CAF and p300/CBP coactivate

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MyoD-dependent transcription in the same pathway. More drastic inhibition was observed in C2C12 myoblasts in similar experiments. Therefore, it can be concluded that P/CAF is essential for transcription of muscle specific genes and hence differentiation into myotubes.

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To further confirm the essential role of P/CAF for myogenic differentiation, we blockage experiments by antibody microinjection were performed. Antibodies were injected into the cytoplasm of proliferating C2C12 myoblasts to prevent the nuclear transport of newly synthesized target proteins. After incubating in the differentiation medium, the degree of differentiation was determined. Microinjection of an anti-P/CAF antibody almost completely inhibited differentiation. Similar results were obtained by microinjecting anti-p300/CBP antibodies. Although microinjection of either anti-p300/CBP or P/CAF antibody was sufficient to inhibit differentiation, an even greater inhibition was observed by coinjecting both of them. Microinjection of anti-P/CAF or anti-p300/CBP antibody did not interfere with induction of p53 by DNA damaging agents, showing specificity of the inhibition by the antibodies. In contrast to anti-P/CAF or anti-p300/CBP antibodies, the injection of anti-MyoD antibody only partially inhibited differentiation, supporting the view of functional redundancy between MyoD and Myf-5 (75,76). Injection of anti-c-Jun antibody or control antibody did not interfere with muscle differentiation.

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Similar experiments were performed with C3H10T1/2 fibroblasts stably expressing MyoD. In these cells, either anti-p300/CBP or anti-P/CAF antibody completely inhibited muscle differentiation. In contrast to myoblasts, anti-MyoD antibody completely blocked differentiation in the fibroblasts expressing MyoD. Anti-c-Jun and control antibodies did not interfere with differentiation. Taken together, these results demonstrate that P/CAF and p300/CBP are indispensable for activation of the myogenic program.

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# p300/CBP, P/CAF and MyoD form a multimeric complex in vivo

The data described above indicate that P/CAF stimulates MyoD-directed transcription via association with p300/CBP. Thus, experiments were conducted to investigate whether P/CAF, p300/CBP and MyoD could associate in a complex.

First, cellular extracts derived from C2C12 myotubes were subjected to immunoprecipitation. Both anti-MyoD and anti-p300/CBP antibodies co-precipitated P/CAF. In a complementary experiment, both anti-p300/CBP and anti-P/CAF antibodies also co-precipitated MyoD, suggesting that these factors form a multimeric protein complex in myotubes.

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Next, attempts were made to detect this complex on the E-box, the DNA binding site for MyoD. Immobilized DNA containing an E-box sequence was incubated with myotube extracts. After extensive washing, P/CAF, p300/CBP and MyoD were analyzed by immunoblotting. P/CAF, p300/CBP and MyoD were all affinity purified on the immobilized DNA, whereas they were not purified on the control DNA lacking the E-box. Given that P/CAF and p300/CBP per se cannot bind to DNA, these observations indicate that P/CAF and p300/CBP are recruited through MyoD at the E-box sites to form a multi-protein complex.

## 20 Complex formation is inhibited by viral transforming factors

Since the oncoviral proteins E1A and large T antigen inhibit myogenic transcription and differentiation, the effect of these factors on the formation of complexes on the E-box was tested. Importantly, very small amouts of P/CAF and p300/CBP were co-purified on the E-box from myocyte extracts which stably express E1A or large T antigen, although MyoD was detected under these conditions. The lower recovery of MyoD from E1A-expressing muscle cells could reflect the low level of MyoD in the extracts (66). These results indicate that E1A and large T antigen dissociate P/CAF and p300/CBP from MyoD without altering MyoD binding to DNA.

Consistent with the previous observations that transiently expressed E1A prevents interaction between P/CAF and p300/CBP in vivo (43), the association

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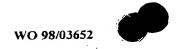
between p300/CBP and P/CAF was abolished in myoblasts stably transformed by wild type E1A but not in those clones transformed with the E1A mutant R2G unable to bind p300/CBP. Similarly, the interaction between p300/CBP and P/CAF was abolished by large T antigen but not by the mutant protein that localizes into the cytoplasm (77).

Interaction between MyoD, P/CAF and CBP in vitro

Previous interaction experiments *in vitro* indicate that the CBP region spanning residues 1801 to 1850 is crucial for interaction with both P/CAF and E1A (43). While most sequence-specific factors bind to CBP sites distinct from the P/CAF/E1A binding sites, MyoD interacts with an overlapping CBP fragment called the CH3 region (60,64,65). To understand how P/CAF, p300/CBP and MyoD associate, the CBP sites important for MyoD binding were mapped more precisely. Consistent with previous reports (60,64,65), the CBP fragment spanning residues 1801-2000 (fragment B) bound MyoD. Moreover, deletion of residues 1801 to 1850 within fragment B completely abolished interaction with MyoD, which is similar to the results obtained with P/CAF and E1A. Importantly, an internal deletion of residues 1850-1878 abolished the MyoD interaction with CBP, while it did not affect binding of E1A or P/CAF (43). These results suggest that MyoD and P/CAF bind to distinct sites of p300/CBP, albeit the binding sites may overlap. Moreover, a direct interaction was observed between MyoD and P/CAF, which may contribute to stabilization of the multimeric complex.

These data show that E1A prevents not only p300/CBP-interaction with P/CAF but also that with MyoD in vivo. To obtain evidence that this inhibition is due to the direct action by E1A, competition experiments were performed in vitro. Importantly, the interaction between CBP and MyoD was strongly inhibited by addition of E1A, implicating that E1A inhibits myogenic transcription by disrupting multiple interactions.

Although the present process has been described with reference to specific details of certain embodiments thereof, it is not intended that such details should be





regarded as limitations upon the scope of the invention except as and to the extent that they are included in the accompanying claims.

Throughout this application various publications are referenced by numbers

within parentheses. Full citations for these publications are as follows. The disclosures of these publications in their entireties are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this invention pertains.

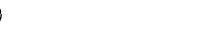
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#### SEQUENCE LISTING

## (1) GENERAL INFORMATION.

- (i) APPLICANT: The United States of America, as repesented by the Secretary, Department of Health and Human Services, c/o National Institutes of Health, Office of Technology Transfer, 6011 Executive Boulevard, Suite 325, Rockville, Maryland 20842
- (ii) TITLE OF THE INVENTION: METHODS AND COMPOSITIONS FOR p300/CBP-ASSOCIATED TRANSCRIPTIONAL CO-FACTOR P/CAF
- (iii) NUMBER OF SEQUENCES: 18
- (iv) CORRESPONDENCE ADDRESS:
  - (A) ADDRESSEE: NEEDLE & ROSENBERG, P.C.
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  - (C) CITY: Atlanta
  - (D) STATE: GA
  - (E) COUNTRY: USA
  - (F) ZIP: 30303
- (V) COMPUTER READABLE FORM:
  - (A) MEDIUM TYPE: Diskette
  - (B) COMPUTER: IBM Compatible
  - (C) OPERATING SYSTEM: DOS
  - (D) SOFTWARE: FastSEQ for Windows Version 2.0
- (vi) CURRENT APPLICATION DATA:
  - (A) APPLICATION NUMBER:
  - (B) FILING DATE: 23-JUL-1997
- (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
  - (A) APPLICATION NUMBER: Corresponding U.S. Serial No. 60/022,273
  - (B) FILING DATE: 23-July-1996
- (viii) ATTORNEY/AGENT INFORMATION:
  - (A) NAME: Miller, Mary L
  - (B) REGISTRATION NUMBER: 39,303
  - (C) REFERENCE/DOCKET NUMBER: 14014.0238/P
- (ix) TELECOMMUNICATION INFORMATION:
  - (A) TELEPHONE: 404/688-0770
  - (B) TELEFAX: 404/688-9880
  - (C) TELEX:
- (2) INFORMATION FOR SEQ ID NO:1:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 832 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
- . (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: None





# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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## (2) INFORMATION FOR SEQ ID NO:2:

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  - (A) LENGTH: 481 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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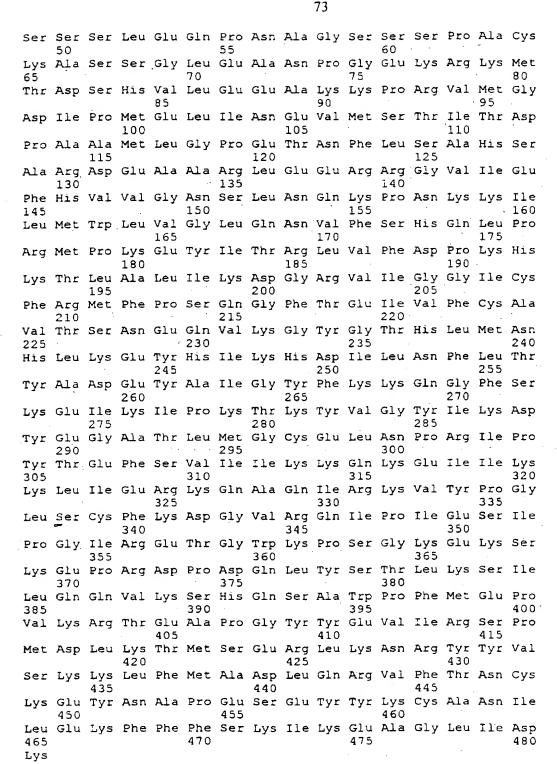
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#### (2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 203 amino acids
  - (B) TYPE: amino acid
- (C) STRANDEDNESS: single



#### (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: None

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## (2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 351 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

### (ii) MOLECULE TYPE: None

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Ser Glu Ala Gly Gly Ala Gly Pro Gly Gly Cys Gly Ala Gly Ala 10 Gly Ala Gly Ala Gly Pro Gly Ala Leu Pro Pro Gln Pro Ala Ala Leu 25 Pro Pro Ala Pro Pro Gln Gly Ser Pro Cys Ala Ala Ala Ala Gly Gly 40 Ser Gly Ala Cys Gly Pro Ala Thr Ala Val Ala Ala Ala Gly Thr Ala 55 Glu Gly Pro Gly Gly Gly Ser Ala Arg Ile Ala Val Lys Lys Ala 75 70 Gln Leu Arg Ser Ala Pro Arg Ala Lys Lys Leu Glu Lys Leu Gly Val 95 90 85 Tyr Ser Ala Cys Lys Ala Glu Glu Ser Cys Lys Cys Asn Gly Trp Lys 110 105 . 100 Asn Pro Asn Pro Ser Pro Thr Pro Pro Arg Ala Asp Leu Gln Gln Ile 125 -120 Ile Val Ser Leu Thr Glu Ser Cys Arg Ser Cys Ser His Ala Leu Ala





Ala His Val Ser His Leu Glu Asn Val Ser Glu Glu Glu Met Asn Arg 150 155 Leu Leu Gly Ile Val Leu Asp Val Glu Tyr Leu Phe Thr Cys Val His 165 170. Lvs Glu Glu Asp Ala Asp Thr Lys Gln Val Tyr Phe Tyr Leu Phe Lys 180 185 Leu Leu Arg Lys Ser Ile Leu Gln Arg Gly Lys Pro Val Val Glu Gly 200 205 195 Ser Leu Glu Lys Lys Pro Pro Phe Glu Lys Pro Ser Ile Glu Gln Gly 215 220 Val Asn Asn Phe Val Gln Tyr Lys Phe Ser His Leu Pro Ala Lys Glu 235 230 Arg Gln Thr Ile Val Glu Leu Ala Lys Met Phe Leu Asn Arg Ile Asn 250 255 245 Tyr Trp His Leu Glu Ala Pro Ser Gln Arg Arg Leu Arg Ser Pro Asn 260 265 Asp Asp Ile Ser Gly Tyr Lys Glu Asn Tyr Thr Arg Trp Leu Cys Tyr 285 280 275 Cys Asn Val Pro Gln Phe Cys Asp Ser Leu Pro Arg Tyr Glu Thr Thr 295 Gln Val Phe Gly Arg Thr Leu Leu Arg Ser Val Phe Thr Val Met Arg 310 315 320 Arg Gln Leu Leu Glu Gln Ala Arg Gln Glu Lys Asp Lys Leu Pro Leu 335 325 Glu Lys Arg Thr Leu Ile Leu Thr His Phe Pro Lys Phe Leu Ser 345 350 340

#### (2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 476 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: None

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Leu Glu Glu Glu Ile Tyr Gly Ala Asn Ser Pro Ile Trp Glu Ser 10 Gly Phe Thr Met Pro Pro Ser Glu Gly Thr Gln Leu Val Pro Arg Pro 25 Ala Ser Val Ser Ala Ala Val Val Pro Ser Thr Pro Ile Phe Ser Pro 40 45 Ser Met Gly Gly Gly Ser Asn Ser Ser Leu Ser Leu Asp Ser Ala Gly 55 Ala Glu Pro Met Pro Gly Glu Lys Arg Thr Leu Pro Glu Asn Leu Thr 7.5 7.0 Leu Glu Asp Ala Lys Arg Leu Arg Val Met Gly Asp Ile Pro Met Glu 90 95 Leu Val Asn Glu Val Met Leu Thr Ile Thr Asp Pro Ala Ala Met Leu 105 110 100 Gly Pro Glu Thr Ser Leu Leu Ser Ala Asn Ala Ala Arg Asp Glu Thr 120 125 Ala Arg Leu Glu Glu Arg Arg Gly Ile Ile Glu Phe His Val Ile Gly 135 140 Asn Ser Leu Thr Pro Lys Ala Asn Arg Arg Val Leu Leu Trp Leu Val 150 155 Gly Leu Gln Asn Val Phe Ser His Gln Leu Pro Arg Met Pro Lys Glu 165 170 175 Tyr Ile Ala Arg Leu Val Phe Asp Pro Lys His Lys Thr Leu Ala Leu 185



Ile Lys Asp Gly Arg Val Ile Gly Gly Ile Cys Phe Arg Met Phe Pro 200 .195 Thr Gln Gly Phe Thr Glu Ile Val Phe Cys Ala Val Thr Ser Asn Glu 220 215 Gln Val Lys Gly Tyr Gly Thr His Leu Met Asn His Leu Lys Glu Tyr 235 -230 His Ile Lys His Asn Ile Leu Tyr Phe Leu Thr Tyr Ala Asp Glu Tyr 250 245 Ala Ile Gly Tyr Phe Lys Lys Gln Gly Phe Ser Lys Asp Ile Lys Val 260 265 Pro Lys Ser Arg Tyr Leu Gly Tyr Ile Lys Asp Tyr Glu Gly Ala Thr 280 275 Leu Met Glu Cys Glu Leu Asn Pro Arg Ile Pro Tyr Thr Glu Leu Ser 300 .295 His Ile Ile Lys Lys Gln Lys Glu Ile Ile Lys Lys Leu Ile Glu Arg 315 310 Lys Gln Ala Gln Ile Arg Lys Val Tyr Pro Gly Leu Ser Cys Phe Lys 335 330 325 Glu Gly Val Arg Gln Ile Pro Val Glu Ser Val Pro Gly Ile Arg Glu 345 Thr Gly Trp Lys Pro Leu Gly Lys Glu Lys Gly Lys Glu Leu Lys Asp 355 360 Pro Asp Gln Leu Tyr Thr Thr Leu Lys Asn Leu Leu Ala Gln Ile Lys 380 375 370 Ser His Pro Ser Ala Trp Pro Phe Met Glu Pro Val Lys Lys Ser Glu 395 390 Ala Pro Asp Tyr Tyr Glu Val Ile Arg Phe Pro Ile Asp Leu Lys Thr 410 405 Met Thr Glu Arg Leu Arg Ser Arg Tyr Tyr Val Thr Arg Lys Leu Phe 425 420 Val Ala Asp Leu Gln Arg Val Ile Ala Asn Cys Arg Glu Tyr Asn Pro 440 Pro Asp Ser Glu Tyr Cys Arg Cys Ala Ser Ala Leu Glu Lys Phe Phe 455 450 Tyr Phe Lys Leu Lys Glu Gly Gly Leu Ile Asp Lys 470

## (2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2414 amino acids
- . (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

 Met
 Ala
 Glu
 Asn
 Val
 Val
 Glu
 Pro
 Gly
 Pro
 Pro
 Ser
 Ala
 Lys
 Arg
 Pro

 Lys
 Leu
 Ser
 Pro
 Ala
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 Glu
 Leu
 Ile
 Ile
 Ile
 Ile
 Ala
 Ile
 Ala





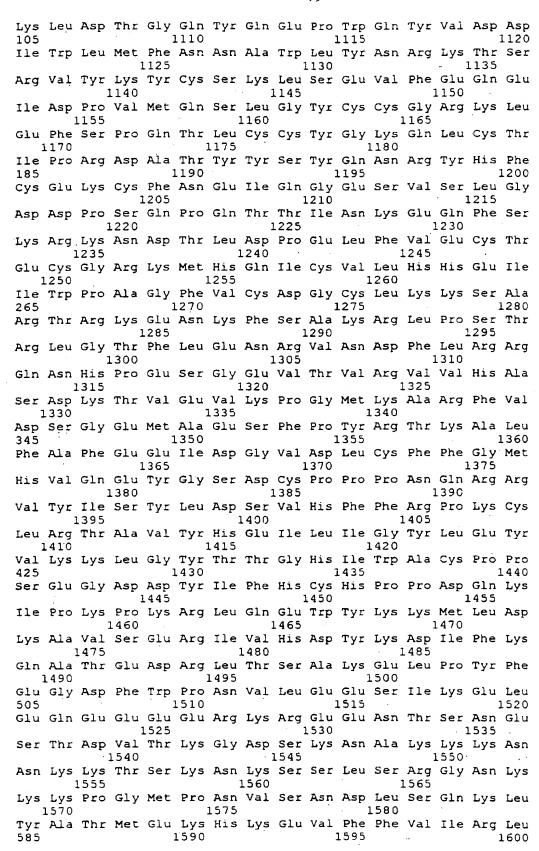
Pro Gly Leu Gly Leu Ile Asn Ser Met Val Lys Ser Pro Met Thr Gln 120 115 Ala Gly Leu Thr Ser Pro Asn Met Gly Met Gly Thr Ser Gly Pro Asn 140 135 Gln Gly Pro Thr Gln Ser Thr Gly Met Met Asn Ser Pro Val Asn Gln 150 . 155 Pro Ala Met Gly Met Asn Thr Gly Thr Asn Ala Gly Met Asn Pro Gly 170 165 Met Leu Ala Ala Gly Asn Gly Gln Gly Ile Met Pro Asn Gln Val Met 185 Asn Gly Ser Ile Gly Ala Gly Arg Gly Arg Gln Asp Met Gln Tyr Pro 200 Asn Pro Gly Met Gly Ser Ala Gly Asn Leu Leu Thr Glu Pro Leu Gln 215 220 Gln Gly Ser Pro Gln Met Gly Gly Gln Thr Gly Leu Arg Gly Pro Gln 230 235 Pro Leu Lys Met Gly Met Met Asn Asn Pro Asn Pro Tyr Gly Ser Pro 250 245 Tyr Thr Gln Asn Pro Gly Gln Gln Ile Gly Ala Ser Gly Leu Gly Leu 265 Gln Ile Gln Thr Lys Thr Val Leu Ser Asn Asn Leu Ser Pro Phe Ala 285 280 Met Asp Lys Lys Ala Val Pro Gly Gly Gly Met Pro Asn Met Gly Gln 295 300 Gln Pro Ala Pro Gln Val Gln Gln Pro Gly Leu Val Thr Pro Val Ala 315 310 Gln Gly Met Gly Ser Gly Ala His Thr Ala Asp Pro Glu Lys Arg Lys 330 . 325 Leu Ile Gln Gln Gln Leu Val Leu Leu His Ala His Lys Cys Gln 345 Arg Arg Glu Gln Ala Asn Gly Glu Val Arg Gln Cys Asn Leu Pro His 365 360 355 Cys Arg Thr Met Lys Asn Val Leu Asn His Met Thr His Cys Gln Ser 375 380 Gly Lys Ser Cys Gln Val Ala His Cys Ala Ser Ser Arg Gln Ile Ile 395 390 Ser His Trp Lys Asn Cys Thr Arg His Asp Cys Pro Val Cys Leu Pro 405 410 Leu Lys Asn Ala Gly Asp Lys Arg Asn Gln Gln Pro Ile Leu Thr Gly 420 425 Ala Pro Val Gly Leu Gly Asn Pro Ser Ser Leu Gly Val Giy Gln Gln 440 Ser Ala Pro Asn Leu Ser Thr Val Ser Gln Ile Asp Pro Ser Ser Ile 455 4.60 Glu Arg Ala Tyr Ala Ala Leu Gly Leu Pro Tyr Gln Val Asn Gln Met 475 470 Pro Thr Gln Pro Gln Val Gln Ala Lys Asn Gln Gln Asn Gln Gln Pro 495 485 490 Gly Gln Ser Pro Gln Gly Met Arg Pro Met Ser Asn Met Ser Ala Ser 505 Pro Met Gly Val Asn Gly Gly Val Gly Val Gln Thr Pro Ser Leu Leu 520 515 Ser Asp Ser Met Leu His Ser Ala Ile Asn Ser Gln Asn Pro Met Met 535 Ser Glu Asn Ala Ser Val Pro Ser Leu Gly Pro Met Pro Thr Ala Ala 550 555 Gln Pro Ser Thr Thr Gly Ile Arg Lys Gln Trp His Glu Asp Ile Thr 570 565 Gln Asp Leu Arg Asn His Leu Val His Lys Leu Val Gln Ala Ile Phe 580 585 Pro Thr Pro Asp Pro Ala Ala Leu Lys Asp Arg Arg Met Glu Asn Leu



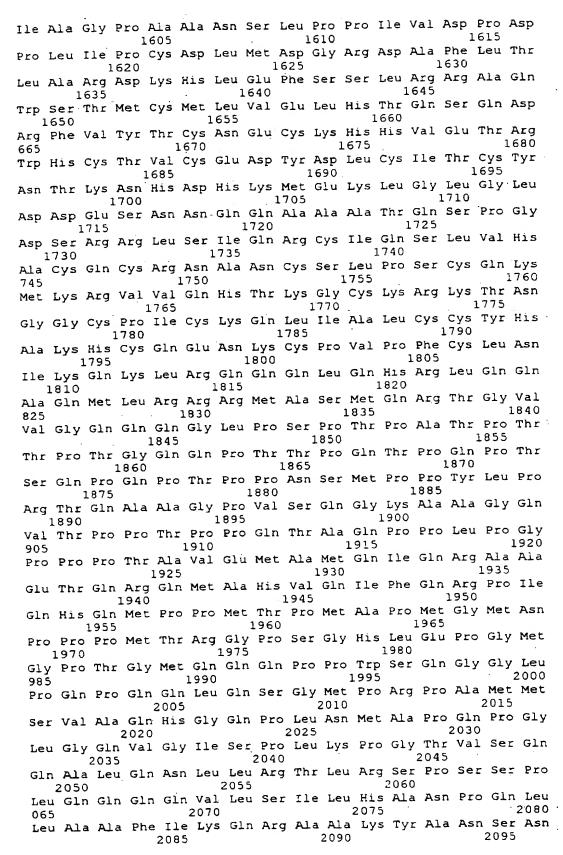
Val Ala Tyr Ala Arg Lys Val Glu Gly Asp Met Tyr Glu Ser Ala Asn 615 Asn Arg Ala Glu Tyr Tyr His Leu Leu Ala Glu Lys Ile Tyr Lys Ile 635 630 Gln Lys Glu Leu Glu Glu Lys Arg Arg Thr Arg Leu Gln Lys Gln Asn 650 645 Met Leu Pro Asn Ala Ala Gly Met Val Pro Val Ser Met Asn Pro Gly 665 660 Pro Asn Met Gly Gln Pro Gln Pro Gly Met Thr Ser Asn Gly Pro Leu 685 680 Pro Asp Pro Ser Met Ile Arg Gly Ser Val Pro Asn Gln Met Met Pro 700 695 Arg Ile Thr Pro Gln Ser Gly Leu Asn Gln Phe Gly Gln Met Ser Met 710 715 Ala Gln Pro Pro Ile Val Pro Arg Gln Thr Pro Pro Leu Gln His His 725 730 Gly Gln Leu Ala Gln Pro Gly Ala Leu Asn Pro Pro Met Gly Tyr Gly 745 750 740 Pro Arg Met Gln Gln Pro Ser Asn Gln Gly Gln Phe Leu Pro Gln Thr 765. 760 Gln Phe Pro Ser Gln Gly Met Asn Val Thr Asn Ile Pro Leu Ala Pro 775 . 780 Ser Ser Gly Gln Ala Pro Val Ser Gln Ala Gln Met Ser Ser Ser 795 790 Cys Pro Val Asn Ser Pro Ile Met Pro Pro Gly Ser Gln Gly Ser His 810 805 Ile His Cys Pro Gln Leu Pro Gln Pro Ala Leu His Gln Asn Ser. Pro 830 825 820 Ser Pro Val Pro Ser Arg Thr Pro Thr Pro His His Thr Pro Pro Ser 840 Ile Gly Ala Gln Gln Pro Pro Ala Thr Thr Ile Pro Ala Pro Val Pro 855 Thr Pro Pro Ala Met Pro Pro Gly Pro Gln Ser Gln Ala Leu His Pro 875 . 870 Pro Pro Arg Gln Thr Pro Thr Pro Pro Thr Thr Gln Leu Pro Gln Gln 890 885 Val Gln Pro Ser Leu Pro Ala Ala Pro Ser Ala Asp Gln Pro Gln Gln 905 Gln Pro Arg Ser Gln Gln Ser Thr Ala Ala Ser Val Pro Thr Pro Ash 925 915 920 Ala Pro Leu Leu Pro Pro Gln Pro Ala Thr Pro Leu Ser Gln Pro Ala 935 940 Val Ser Ile Glu Gly Gln Val Ser Asn Pro Pro Ser Thr Ser Ser Thr 950 955 Glu Val Asn Ser Gln Ala Ile Ala Glu Lys Gln Pro Ser Gln Glu Val 970 965 Lys Met Glu Ala Lys Met Glu Val Asp Gln Pro Glu Pro Ala Asp Thr 985 Gln Pro Glu Asp Ile Ser Glu Ser Lys Val Glu Asp Cys Lys Met Glu 995 1000 1005 Ser Thr Glu Thr Glu Glu Arg Ser Thr Glu Leu Lys Thr Glu Ile Lys . 1015 1020 Glu Glu Glu Asp Gln Pro Ser Thr Ser Ala Thr Gln Ser Ser Pro Ala . 1030 1035 Pro Gly Gln Ser Lys Lys Ile Phe Lys Pro Glu Glu Leu Arg Gln 1050 1045 Ala Leu Met Pro Thr Leu Glu Ala Leu Tyr Arg Gln Asp Pro Glu Ser . \* .1060 1065 Leu Pro Phe Arg Gln Pro Val Asp Pro Gln Leu Leu Gly Ile Pro Asp 1075 1080 1085 Tyr Phe Asp Ile Val Lys Ser Pro Met Asp Leu Ser Thr Ile Lys Arg 1095













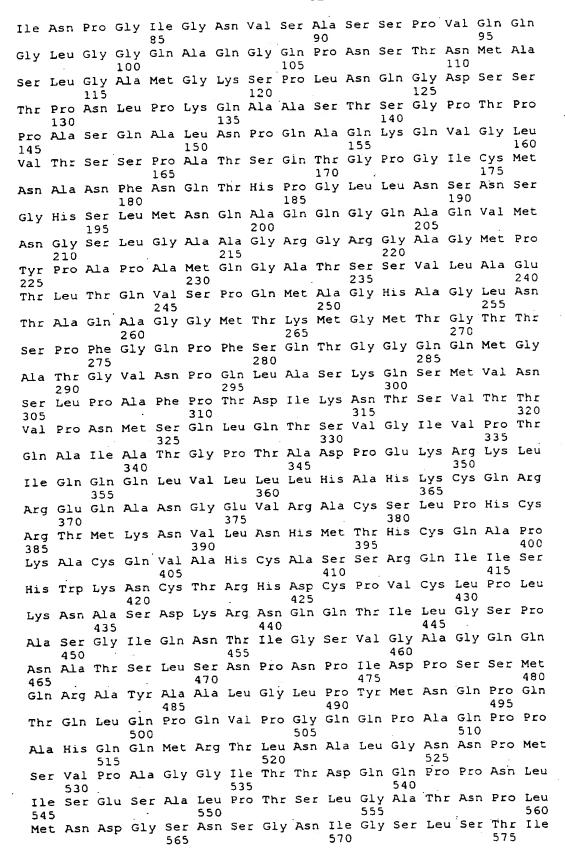
Pro Gln Pro Ile Pro Gly Gln Pro Gly Met Pro Gln Gly Gln Pro Gly . 2110 2105 Leu Gln Pro Pro Thr Met Pro Gly Gln Gln Gly Val His Ser Asn Pro 2120 2115 2125 Ala Met Gln Asn Met Asn Pro Met Gln Ala Gly Val Gln Arg Ala Gly 2135 2140 Leu Pro Gln Gln Gln Pro Gln Gln Gln Leu Gln Pro Pro Met Gly Gly 2150 2155 2160 Met Ser Pro Gln Ala Gln Gln Met Asn Met Asn His Asn Thr Met Pro 2170 2165 Ser Gln Phe Arg Asp Ile Leu Arg Arg Gln Gln Met Met Gln Gln 2180 2185 2190 Gln Gln Gly Ala Gly Pro Gly Ile Gly Pro Gly Met Ala Asn His 2200 2205 2195 Asn Gln Phe Gln Gln Pro Gln Gly Val Gly Tyr Pro Pro Gln Pro Gln 2210 2215 2220 Gln Arg Met Gln His His Met Gln Gln Met Gln Gly Asn Met Gly 2230 2235 Gln Ile Gly Gln Leu Pro Gln Ala Leu Gly Ala Glu Ala Gly Ala Ser 2245 2250 Leu Gln Ala Tyr Gln Gln Arg Leu Leu Gln Gln Met Gly Ser Pro 2260 2265 2270 Val Gln Pro Asn Pro Met Ser Pro Gln Gln His Met Leu Pro Asn Gln 2275 2280 2285 Ala Gln Ser Pro His Leu Gln Gly Gln Gln Ile Pro Asn Ser Leu Ser 2290 2295 2300 Asn Gln Val Arg Ser Pro Gln Pro Val Pro Ser Pro Arg Pro Gln Ser 2310 2315 Gln Pro Pro His Ser Ser Pro Ser Pro Arg Met Gln Pro Gln Pro Ser 2330 2335 2325 Pro His His Val Ser Pro Gln Thr Ser Ser Pro His Pro Gly Leu Val 2345 2350 2340 Ala Ala Gln Ala Asn Pro Met Glu Gln Gly His Phe Ala Ser Pro Asp 2360 2365 2355 Gln Asn Ser Met Leu Ser Gln Leu Ala Ser Asn Pro Gly Met Ala Asn 2375 2380 Leu His Gly Ala Ser Ala Thr Asp Leu Gly Leu Ser Thr Asp Asn Ser 2390 2395 Asp Leu Asn Ser Asn Leu Ser Gln Ser Thr Leu Asp Ile His 2410

#### (2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2441 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Met Ala Glu Asn Leu Leu Asp Gly Pro Pro Asn Pro Lys Arg Ala Lys 10 Leu Ser Ser Pro Gly Phe Ser Ala Asn Asp Asn Thr Asp Phe Gly Ser 20 25 30 Leu Phe Asp Leu Glu Asn Asp Leu Pro Asp Glu Leu Ile Pro Asn Gly 40 45 35 Glu Leu Ser Leu Leu Asn Ser Gly Asn Leu Val Pro Asp Ala Ala Ser 55 60 Lys His Lys Gln Leu Ser Glu Leu Leu Arg Gly Gly Ser Gly Ser Ser 🕟 70 75



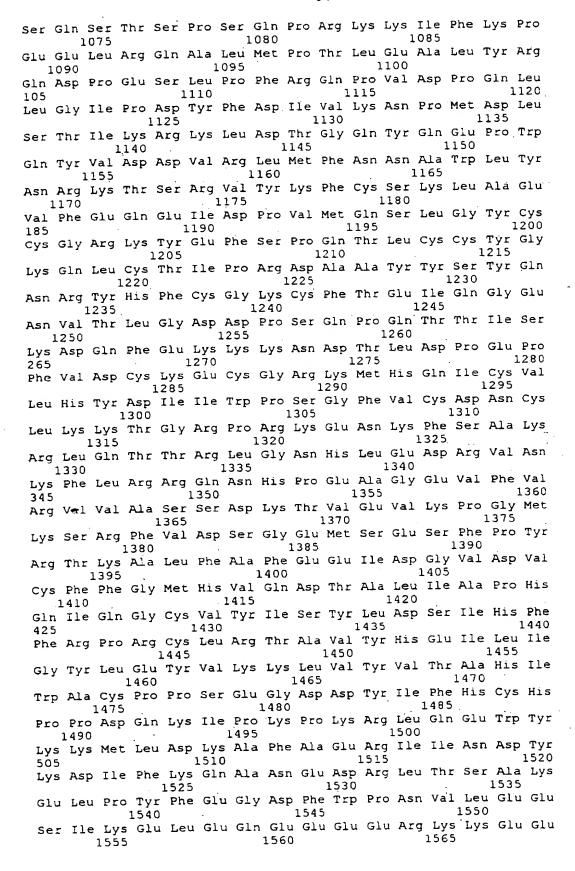






Pro Thr Ala Ala Pro Pro Ser Ser Thr Gly Val Arg Lys Gly Trp His Glu His Val Thr Gln Asp Leu Arg Ser His Leu Val His Lys Leu Val Gln Ala Ile Phe Pro Thr Pro Asp Pro Ala Ala Leu Lys Asp Arg Arg Met Glu Asn Leu Val Ala Tyr Ala Lys Lys Val Glu Gly Asp Met Tyr . 640 Glu Ser Ala Asn Ser Arg Asp Glu Tyr Tyr His Leu Leu Ala Glu Lys Ile Tyr Lys Ile Gln Lys Glu Leu Glu Glu Lys Arg Arg Thr Arg Leu His Lys Gln Gly Ile Leu Gly Asn Gln Pro Ala Leu Pro Ala Ser Gly Ala Gln Pro Pro Val Ile Pro Pro Ala Gln Ser Val Arg Pro Pro Asn Gly Pro Leu Pro Leu Pro Val Asn Arg Met Gln Val Ser Gln Gly Met Asn Ser Phe Asn Pro Met Ser Leu Gly Asn Val Gln Leu Pro Gln Ala Pro Met Gly Pro Arg Ala Ala Ser Pro Met Asn His Ser Val Gin Met Asn Ser Met Ala Ser Val Pro Gly Met Ala Ile Ser Pro Ser Arg Met Pro Gln Pro Pro Asn Met Met Gly Thr His Ala Asn Asn Ile Met Ala Gln Ala Pro Thr Gln Asn Gln Phe Leu Pro Gln Asn Gln Phe Pro Ser Ser Ser Gly Ala Met Ser Val Asn Ser Val Gly Met Gly Gln Pro Ala Ala Gln Ala Gly Val Ser Gln Gly Gln Glu Pro Gly Ala Ala Leu Pro Asn Pro Leu Asn Met Leu Ala Pro Gln Ala Ser Gln Leu Pro Cys Pro Pro Val Thr Gln Ser Pro Leu His Pro Thr Pro Pro Pro Ala Ser Thr Ala Ala Gly Met Pro Ser Leu Gln His Pro Thr Ala Pro Gly Met Thr Pro Pro Gln Pro Ala Ala Pro Thr Gln Pro Ser Thr Pro Val Ser Ser Gly Gln Thr Pro Thr Pro Thr Pro Gly Ser Val Pro Ser Ala Ala Gln Thr Gln Ser Thr Pro Thr Val Gln Ala Ala Ala Gln Ala Gln Val Thr Pro Gln Pro Gln Thr Pro Val Gln Pro Pro Ser Val Ala Thr Pro Gln Ser Ser Gln Gln Pro Thr Pro Val His Thr Gln Pro Pro Gly Thr Pro Leu Ser Gln Ala Ala Ala Ser Ile Asp Asn Arg Val Pro Thr Pro Ser Thr Val Thr Ser Ala Glu Thr Ser Ser Gln Gln Pro Gly Pro Asp Val Pro Met Leu Glu Met Lys Thr Glu Val Gln Thr Asp Asp Ala Glu 1000 1005 Pro Glu Pro Thr Glu Ser Lys Gly Glu Pro Arg Ser Glu Met Met Glu Glu Asp Leu Gln Gly Ser Ser Gln Val Lys Glu Glu Thr Asp Thr Thr 1030 1035 1040 Glu Gln Lys Ser Glu Pro Met Glu Val Glu Glu Lys Lys Pro Glu Val 1050 1055 Lys Val Glu Ala Lys Glu Glu Glu Asn Ser Ser Asn Asp Thr Ala









Ser Thr Ala Ala Ser Glu Thr Pro Glu Gly Ser Gln Gly Asp Ser Lys 1575 1580 Asn Ala Lys Lys Lys Asn Asn Lys Lys Thr Asn Lys Asn Lys Ser Ser 1590 1595 · 1600 Ile Ser Arg Ala Asn Lys Lys Lys Pro Ser Met Pro Asn Val Ser Asn 1605 1610 1615 Asp Leu Ser Gln Lys Leu Tyr Ala Thr Met Glu Lys His Lys Glu Val 1620 1625 1630 Phe Phe Val Ile His Leu His Ala Gly Pro Val Ile Ser Thr Gln Pro 1640 1645 Pro Ile Val Asp Pro Asp Pro Leu Leu Ser Cys Asp Leu Met Asp Gly 1655 1660 Arg Asp Ala Phe Leu Thr Leu Ala Arg Asp Lys His Trp Glu Phe Ser 1670 1675 1680 Ser Leu Arg Arg Ser Lys Trp Ser Thr Leu Cys Met Leu Val Glu Leu 1685 1690 1695 His Thr Gln Gly Gln Asp Arg Phe Val Tyr Thr Cys Asn Glu Cys Lys 1700 1705 1710 His His Val Glu Thr Arg Trp His Cys Thr Val Cys Glu Asp Tyr Asp 1715 1720 1725 Leu Cys Ile Asn Cys Tyr Asn Thr Lys Ser His Thr His Lys Met Val 1730 1735 1740 Lys Trp Gly Leu Gly Leu Asp Asp Glu Gly Ser Ser Gln Gly Glu Pro 1750 1755 1760 745 Gln Ser Lys Ser Pro Gln Glu Ser Arg Arg Leu Ser Ile Gln Arg Cys 1765 1770 1775 Ile Gln Ser Leu Val His Ala Cys Gln Cys Arg Asn Ala Asn Cys Ser 1780 1785 1790 Leu Pro Ser Cys Gln Lys Met Lys Arg Val Val Gln His Thr Lys Gly 1800 1805 Cys Lys Arg Lys Thr Asn Gly Gly Cys Pro Val Cys Lys Gln Leu Ile 1820 1810 1815 Ala Leu Cys Cys Tyr His Ala Lys His Cys Gln Glu Asn Lys Cys Pro 1830 1835 Val Pro Phe Cys Leu Asn Ile Lys His Asn Val Arg Gln Gln Ile 1850 1855 1845 Gln His Cys Leu Gln Gln Ala Gln Leu Met Arg Arg Met Ala Thr 1865 1870 1860 Met Asn Thr Arg Asn Val Pro Gln Gln Ser Leu Pro Ser Pro Thr Ser 1880 1885 Ala Pro Pro Gly Thr Pro Thr Gln Gln Pro Ser Thr Pro Gln Thr Pro 1895 1900 Gln Pro Pro Ala Gln Pro Gln Pro Ser Pro Val Asn Met Ser Pro Ala 1910 1915 Gly Phe Pro Asn Val Ala Arg Thr Gln Pro Pro Thr Ile Val Ser Ala 1930 1925 Gly Lys Pro Thr Asn Gln Val Pro Ala Pro Pro Pro Pro Ala Gln Pro 1945 1950 Pro Pro Ala Ala Val Glu Ala Ala Arg Gln Ile Glu Arg Glu Ala Gln 1960 1965 . 1955 Gln Gln Gln His Leu Tyr Arg Ala Asn Ile Asn Asn Gly Met Pro Pro 1975 1980 Gly Arg Asp Gly Met Gly Thr Pro Gly Ser Gln Met Thr Pro Val Gly 1990 1995 2000 985 Leu Asn Val Pro Arg Pro Asn Gln Val Ser Gly Pro Val Met Ser Ser 2005 2010 2015 Met Pro Pro Gly Gln Trp Gln Gln Ala Pro Ile Pro Gln Gln Gln Pro 2020 2025 2030 Met Pro Gly Met Pro Arg Pro Val Met Ser Met Gln Ala Gln Ala Ala 2040 2045 Val Ala Gly Pro Arg Met Pro Asn Val Gln Pro Asn Arg Ser Ile Ser 2055 -2060



Pro Ser Ala Leu Gln Asp Leu Leu Arg Thr Leu Lys Ser Pro Ser Ser 2070 2075 2080 065 Pro Gln Gln Gln Gln Val Leu Asn Ile Leu Lys Ser Asn Pro Gln 2095 2085 2090 Leu Met Ala Ala Phe Ile Lys Gln Arg Thr Ala Lys Tyr Val Ala Asn 2100 2105 2110 Gln Pro Gly Met Gln Pro Gln Pro Gly Leu Gln Ser Gln Pro Gly Met 2115 2120 2125 Gln Pro Gln Pro Gly Met His Gln Gln Pro Ser Leu Gln Asn Leu Asn 2130 2135 2140 Ala Met Gln Ala Gly Val Pro Arg Pro Gly Val Pro Pro Pro Gln Pro 145 2150 2155 2160 Ala Met Gly Gly Leu Asn Pro Gln Gly Gln Ala Leu Asn Ile Met Asn 2165 2170 2175 Pro Gly His Asn Pro Asn Met Thr Asn Met Asn Pro Gln Tyr Arg Glu 2180 2185 2190 Met Val Arg Arg Gln Leu Leu Gln His Gln Gln Gln Gln Gln Gln 2195 2200 Gln Gln Gln Gln Gln Gln Gln Asn Ser Ala Ser Leu Ala Gly Gly 2210 2215 2220 Met Ala Gly His Ser Gln Phe Gln Gln Pro Gln Gly Pro Gly Gly Tyr 225 2230 2235 Ala Pro Ala Met Gln Gln Gln Arg Met Gln Gln His Leu Pro Ile Gln 2245 2250 . 2255 Gly Ser Ser Met Gly Gln Met Ala Ala Pro Met Gly Gln Leu Gly Gln 2260 2265 2270 Met Gly Gln Pro Gly Leu Gly Ala Asp Ser Thr Pro Asn Ile Gln Gln 2275 2280 2285 Ala Leu Gln Gln Arg Ile Leu Gln Gln Gln Gln Met Lys Gln Gln Ile 2290 2295 2300 Gly Ser Pro Gly Gln Pro Asn Pro Met Ser Pro Gln Gln His Met Leu 305 2310 2315 2320 Ser Gly Gln Pro Gln Ala Ser His Leu Pro Gly Gln Gln Ile Ala Thr 2325 2330 2335 Ser Leu Ser Asn Gln Val Arg Ser Pro Ala Pro Val Gln Ser Pro Arg 2340 2345 2350 Pro Gln Ser Gln Pro Pro His Ser Ser Pro Ser Pro Arg Ile Gln Pro 2355 2360 2365
Gln Pro Ser Pro His His Val Ser Pro Gln Thr Gly Thr Pro His Pro 2375 2380 Gly Leu Ala Val Thr Met Ala Ser Ser Met Asp Gln Gly His Leu Gly 385 2390 2395 2400 Asn Pro Glu Gln Ser Ala Met Leu Pro Gln Leu Asn Thr Pro Asn Arg 2415 2405 2410 Ser Ala Leu Ser Ser Glu Leu Ser Leu Val Gly Asp Thr Thr Gly Asp 2430 2420 . 2425 Thr Leu Glu Lys Phe Val Glu Gly Leu 2435 2440

## (2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 813 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear ...
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met Ala Glu Ala Gly Gly Ala Gly Ser Pro Ala Leu Pro Pro Ala Pro 1 5 10 15



Pro His Gly Ser Pro Arg Thr Leu Ala Thr Ala Ala Gly Ser Ser Ala Ser Cys Gly Pro Ala Thr Pro Val Ala Ala Ala Gly Thr Ala Glu Gly Pro Gly Gly Gly Ser Ala Arg Ile Ala Val Lys Lys Ala Gln Leu Arg Ser Ala Pro Arg Ala Lys Lys Leu Glu Lys Leu Gly Val Tyr Ser Ala Cys Lys Ala Glu Glu Ser Cys Lys Cys Asn Gly Trp Lys Asn Pro 8.5 Asn Pro Ser Pro Thr Pro Pro Arg Gly Asp Leu Gln Gln Ile Ile Val Ser Leu Thr Glu Ser Cys Arg Ser Cys Ser His Ala Leu Ala Ala His Val Ser His Leu Glu Asn Val Ser Glu Glu Glu Met Asp Arg Leu Leu Gly Ile Val Leu Asp Val Glu Tyr Leu Phe Thr Cys Val His Lys Glu . 150 Glu Asp Ala Asp Thr Lys Gln Val Tyr Phe Tyr Leu Phe Lys Leu Leu Arg Lys Ser Ile Leu Gln Arg Gly Lys Pro Val Val Glu Gly Ser Leu Glu Lys Lys Pro Pro Phe Glu Lys Pro Ser Ile Glu Gln Gly Val Asn Asn Phe Val Gln Tyr Lys Phe Ser His Leu Pro Ser Lys Glu Arg Gln Thr Thr Ile Glu Leu Ala Lys Met Phe Leu Asn Arg Ile Asn Tyr Trp His Leu Glu Ala Pro Ser Gln Arg Arg Leu Arg Ser Pro Asn Asp Asp Ile Ser Gly Tyr Lys Glu Asn Tyr Thr Arg Trp Leu Cys Tyr Cys Asn Val Pro Gln Phe Cys Asp Ser Leu Pro Arg Tyr Glu Thr Thr Lys Val Phe Gly Arg Thr Leu Leu Arg Ser Val Phe Thr Ile Met Arg Arg Gln Leu Leu Glu Gln Ala Arg Gln Lys Lys Asp Lys Leu Pro Leu Glu Lys Arg Thr Leu Ile Leu Thr His Phe Pro Lys Phe Leu Ser Met Leu Glu Glu Glu Val Tyr Ser Gln Asn Ser Pro Ile Trp Asp Gln Asp Phe Leu Ser Ala Ser Ser Arg Thr Ser Pro Leu Gly Ile Gln Thr Val Ile Ser Pro Pro Val Thr Gly Thr Ala Leu Phe Ser Ser Asn Ser Thr Ser His Glu Gln Ile Asn Gly Gly Arg Thr Ser Pro Gly Cys Arg Gly Ser Ser Gly Leu Glu Ala Asn Pro Gly Glu Lys Arg Lys Met Asn Asn Ser His Ala Pro Glu Glu Ala Lys Arg Ser Arg Val Met Gly Asp Ile Pro Val Glu Leu Ile Asn Glu Val Met Ser Thr Ile Thr Asp Pro Ala Gly Met Leu Gly Pro Glu Thr Asn Phe Leu Ser Ala His Ser Ala Arg Asp Glu Ala Ala Arg Leu Glu Glu Arg Arg Gly Val Ile Glu Phe His Val Val Gly Asr. Ser Leu Asn Gln Lys Pro Asn Lys Lys Ile Leu Met Trp Leu Val Gly Leu Gln Asn Val Phe Ser His Gln Leu Pro Arg Met Pro Lys 



Glu	Tyr	Ile 515	Thr	Arg	Leu	Val	Phe 520	Asp	Pro	Lys	His	Lys 525	Thr	Leu	Ala
Leu	11e 530	Lys	Asp	Gly	Arg	Val 535	Ile	Gly	Gly	Ile	Cys 540	Phe	Arg	Met	Phe
Pro 545	Ser	Gln	Gly	Phe	Thr 550	Glu	Ile	Val	Phe	Cys 555	Ala	Val	Thr	Ser	Asn 560
Glu	Gln	Val	Lys	G1 y 565		Gly	Thr	His	Leu 570	Met	Asn	His	Leu	Lys 575	Glu
-			580	His				Asn 585					590	•	
_		595	Gly	Tyr	Phe	2-	600	Gln				605			
	610	Lys	Thr			615		Tyr			620				
625	Leu				630			Pro		635					640
ser				645				Glu	650					622	
_			660					Val 665					6/0		
-		675					680					685			•
	690					695		Lys			700				
705					710					715					Val 720
_				725					730					/35	Thr
			740	) ·				745					750		Lys
	•	755	· ·				760	)				765			Leu
	770	<b>,</b>		•		775	<b>,</b>				780				Asn
785					790	)				795	)			Lys	Phe 800
Phe	Ph∈	se:	c Lys	805		s Glu	ı Ala	a Gly	810	l Ile	: Asp	Lys	•		

## (2) INFORMATION FOR SEQ ID NO:9:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 50 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

His Thr Lys Gly Cys Lys Arg Lys Thr Asn Gly Gly Cys Pro Val Cys 1 10 15

Lys Gln Leu Ile Ala Leu Cys Cys Tyr His Ala Lys His Cys Gln Glu 20 25 30.

Asn Lys Cys Pro Val Pro Phe Cys Leu Asn Ile Lys His Asn Val Arg 35

Gln Gln 50

## (2) INFORMATION FOR SEQ ID NO:10:

.(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2204 base pairs





- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

ACCCACTCCC	CCCAGAGCCG				AATCCTGTCG	. 60
GAGTTGTAGC			TTCCCACCTG		CAGAGGAAGA	120
AATGAACAGA	CTCCTGGGAA			CTCTTTACCT	GTGTCCACAA	180
GGAAGAAGAT		AACAAGTTTA		TTTAAGCTCT	TGAGAAAGTC	240
TATTTTACAA	AGAGGAAAAC	CTGTGGTTGG	AAGGCTCTTT	GGAAAAGAAA	CCCCCATTTG	300
AAAAACCTAG	CATTGAACAG	GGTGTGAATA	ACTTTGTGCA		AGTCACCTGC	360
CAGCAAAAAG	AAAGGCAAAC	CAATAGTTGA	GTTGGCAAAA	ATGTTCCTAA	·ACCGCATCAC	420
CTATTGGCAT	CTGGAGGCAC	CATCTCAACG	AGACTGCGAT	CTCCAATGAT	GATATTCTGG	480
ATACAAAGAG	AACTACACAA	GGTGGCTGTG	TTACTGCAAC	GTGCCACAGT	TCTGCGACAG	540
TCTACCTCGG	TACGAAACCA		TGGGAGAACA		GGTCTTCACT	600
GTTATGAGGC	GACAACTCCT		AGACAGGAAA		GCCTCTTGAA	660
AAACGAACTC	TAATCCTCAC	TCATTTCCCA			AGAAGAAGTA	720
TATAGTCAAA	ACTCTCCCAT	CTGGGATCAC	CATTTTCTCT	CAGCCTCTTC	CAGAACCAGC	780
CAGCTAGGCA	TCCAAACAGT	TATCAATCAC	CTCCTGTGGC	TGGGACAATT	TCATACAATT	840
CAACCTCATC	TTCCCTTGAG	CAGCCAAACG	••••	CAGTCCTGCC	TGCAAAGCCT	900
CTTCTGGACT	TGAGGCAAAC	CCAGGAGAAA		GACTGATTCT	CATGTTCTGG	960
AGGAGGCCAA'	GAAACCCCGA	GTTATGGGGG		GGAATTAATC	AACGAGGTTA	1020
TGTCTACCAT	CACGGACCCT	GCAGCAATGC	TTGGACCAGA		CTGTCAGCAC	1080
ACTCGGCCAG	GGATGAGGCG	GCAAGGTTGG	AAGAGCGCAG	GGGTGTAATT	GAATTTCACG	1140
TGGTTGGCAA		CAGAAACCAA		CCTGATGTGG	CTGGTTGGCC	1200
TACAGAACGT		CAGCTGCCCC	•••	AGAATACATC	<del>-</del>	1260
TCTTTGACCC	GAAACACAAA		TAATTAAAGA		ATTGGTGGTA	1320
TCTGTTTCCG			TCACAGAGAT	TGTCTTCTGT	GCTGTAACCT	1380
	AGTCAAGGGC		ACCTGATGAA		GAATATCACA	1440
TAAAGCATGA	CATCCTGAAC	TTCCTCACAT	ATGCAGATGA		GGATACTTTA	1500
AGAAACAGGG		GAAATTAAAA		CAAATATGTT	GGCTATATCA	1560
AGGATTATGA	AGGAGCCACT		GTGAGCTAAA		CCGTACACAG	1620
AATTTTCTGT	CATCATTAAA	AAGCAGAAGG			GAAAGAAAAC	1680
AGGCACAAAT	TCGAAAAGTT	TACCCTGGAC	TTTCATGTTT		GTTCGACAGA	1740
TTCCTATAGA	AAGCATTCCT		AGACAGGCTG		GGAAAAGAGA	1800
AAAGTAAAGA		CCTGACCAGC	TTTACAGCAC	GCTCAAGAGC		1860
AGGTGAAGAG		GCTTGGCCCT	TCATGGAACC	TGTGAAGAGA		1920
CAGGATATTA		AGGTCCCCCA			GAACGCCTCA	1980
		AAGAAATTAT	• • • • • • • • • • • • • • • • • • • •	CTTACAGCGA		2040
	GTACAACGCC			ATGTGCCAAT		2100
AATTCTTCTT	CAGTAAAATT	AAGGAAGCTG			TTTTTCCCCC	2160
TCTGCTTCTT	AGAAACTCAC	CAAGCAGTGT	GCCTAAAGCA	AGGT		2204

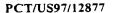
## (2) INFORMATION FOR SEQ ID NO:11:

#### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2093 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

GAATTCCGGC	GAAACCACTC	ATGTCTTTGG	GCGAAGCCTT	CTCCGGTCCA	TTTTCACCGT	60
TACCCGCCGG	CAGCTGCTGG	AAAAGTTCCG	AGTGGAGAAG	GACAAATTGG	TGCCCGAGAA	120
GAGGACCCTC	ATCCTCACTC	ACTTCCCCAA	GTAAGGCTCC	TTCTGGCCTA	CCAGGATTTG	180
GCCCCAAGTT	CACATCCTCC	CTGTTGTCCC	CTTTTTTCCA	GGAAGGCTTC	CTGGATTGGT	240
CCCTCCTCTC	CCTCCATGGG	CCTTTTGGGA	TCTGGGCGTC	TACCTGGCAG	ACTTGCCCAT	300
GGCCCAGAAG	CAACTTGCTA	GTACTAGTCT	GGGGATGGCA	GATTCCTGTC	CATGCTGGAG	360
GAGGAGATCT	ATGGGGCAAA	CTCTCCAATC	TGGGAGTCAG	GCTTCACCAT	GCCACCCTCA	420









GAGGGGACAC AGCTGGTTCC CCGGCCAGCT TCAGTCAGTG CAGCGGTTGT TCCCAGCACC CCCATCTCA GCCCCAGCAT GGGTGGGGGC AGCAACAGCT CCCTGAGTCT GGATTCTGCA GGGGCCGAGC CTATGCCAGG CGAGAAGAGG ACGCTCCCAG AGAACCTGAC CCTGGAGGAT GCCAAGCGGC TCCGTGTGAT GGGTGACATC CCCATGGAGC TGGTCAATGA GGTCATGCTG ACCATCACTG ACCCTGCTGC CATGCTGGGG CCTGAGACGA GCCTGCTTTC GGCCAATGCG 720 GCCCGGGATG AGACAGCCCG CCTGGAGGAG CGCCGCGGCA TCATCGAGTT CCATGTCATC GGCAACTCAC TGACGCCCAA GGCCAACCGG CGGGTGTTGC TGTGGCTCGT GGGGCTGCAG AATGTCTTTT CCCACCAGCT GCCGCGCATG CCTAAGGAGT ATATCGCCCG CCTCGTCTTT 900 GACCCGAAGC ACAAGACTCT GGCCTTGATC AAGGATGGGC GGGTCATCGG TGGCATCTGC 960 TTCCGCATGT TTCCCACCCA GGGCTTCACG GAGATTGTCT TCTGTGCTGT CACCTCGAAT 1020 GAGCAGGTCA AGGGTTATGG GACCCACCTG ATGAACCACC TGAAGGAGTA TCACATCAAG CACAACATTC TCTACTTCCT CACCTACGCC GACGAGTACG CCATCGGCTA CTTCAAAAAG 1080 1140 CAGGGTTTCT CCAAGGACAT CAAGGTGCCC AAGAGCCGCT ACCTGGGCTA CATCAAGGAC TACGAGGGAG CGACGCTGAT GGAGTGTGAG CTGAATCCCC GCATCCCCTA CACGGAGCTG 1260 TCCCACATCA TCAAGAAGCA GAAAGAGATC ATCAAGAAGC TGATTGAGCG CAAACAGGCC 1320 CAGATCCGCA AGGTCTACCC GGGGCTCAGC TGCTTCAAGG AGGGCGTGAG GCAGATCCCT 1380 GTGGAGAGCG TTCCTGGCAT TCGAGAGACA GGCTGGAAGC CATTGGGGAA GGAGAAGGGG 1440 AAGGAGCTGA AGGACCCCGA CCAGCTCTAC ACAACCCTCA AAAACCTGCT GGCCCAAATC 1500 AAGTCTCACC CCAGTGCCTG GCCCTTCATG GAGCCTGTGA AGAAGTCGGA GGCCCCTGAC 1560 TACTACGAGG TCATCCGCTT CCCCATTGAC CTGAAGACCA TGACTGAGCG GCTGCGAAGC 1620 CGCTACTACG TGACCCGGAA GCTCTTTGTG GCCGACCTGC AGCGGGTCAT CGCCAACTGT 1680 CGCGAGTACA ACCCCCGGA CAGCGAGTAC TGCCGCTGTG CCAGCGCCCT GGAGAAGTTC 1740 TTCTACTTCA AGCTCAAGGA GGGAGGCCTC ATTGACAAGT AGGCCCATCT TTGGGCCGCA 1800 AGCTCTTTCT GGACCTTCAG GCACCCCCAA GCGTGCAGCT CTGTCCCAGC CTTCACTGTG 1980 TGTGAGAGGT CTCCTGGGTT GGGGCCCAGC CCCTCTAGAG TAGCTGGTGG CCAGGGATGA ACCTTGCCCA GCCGTGGTGG CCCCCAGGCC TGGTCCCCAA GAGCCCGGAA TTC 2093

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#### (2) INFORMATION FOR SEQ ID NO:12:

# (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 9046 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

CCTTGTTTGT	GTGCTAGGCT	GGGGGGAGA	GAGGGCGAGA	GAGAGCGGGC	GAGAGTGGGC	60
	CCGGGCTGAG			GAGTGCGGAG	GGGAGTCGGG	120
AAGCAGGACG	GCGGCAGGGG		GGCAGGGGGC	CCGGGGCGCA	CGGGCTGAGG	180
TCGGAGAGAG	CCCCCTCCCG	TCCGCACACA	CCCCCACCGC	GGTCCAGCAG	CCGGGCCGGC	240
CGACCCCCAG	GGGGGGACCA	TTACATAACC	CGCGCCCCGG	CCGTCTTCTC	CCGCCGCCGC	300
GTCGACGCTA		GGCGGGCGCT	CCAGCACTGG	CCGCCGGCGT	GGGGCGTAGC	360
GGCGCCCGAA	CTGAGCCCGG	GCGGAAAGGA	AGGCGAAGGA	GGGGAGCGCC	GGCGCGAGGA	420
AGCGGCCGTA	TTATTATTTC		GCCTCCTCGG			480
GGGGCCGCCT	ececcecce	CCGGAGCGGG	-	CTGGTCCGGC	CAGCTCCGCT	540
GCGTGCGGGC	GGCGCTGCTC	GGCCCGGCCC	CCTCGGCCCT	GTGAGGCGGC	GGCGCCAGCC	600
CCCGGCGTCC	TTGCCGCGCC	TCCGCCGGCC	GCCGCGCGAT		GGCGCCAGCC	660
TGGCTCTCGG	CTCGGGCGAG	TTCTCTGCGG	CCATTAGGGG	CCGGTGCGGC		720
AGCGCGGCGG	CAGGAGGAGG		TGGGGGCGCA		GGGGCACCGG	780
GAGGAGGTGA	GTGTCTCTTG	TCGCCTCCTC	CTCTCCCCCC	TTTTCGCCCC	CGCCTCCTTG	_
TGGCGATGAG	AAGGAGGAGG	ACAGCGCCGA		GTTGATGGCG		840
TCCGAGAGAC	CTCGGCTGGG	CAGGGGCCGG	CCGTGGCGGG	CCGGGGACTG	CGCCTCTAGA	900
GCCGCGAGTT		CGCCGCAGCG	GACCGGCCTC	GGCGAATTTG		960
CCCTCCTCCG			CTCGCACTTG	CCCTTACCTT	TTCTATCGAG	1020
TCCGCATCCC			GCGAAGAGAA	AAAGGAACTT	CCCCCACCCC	1080
CTCGGGTGCC			CCCTGGGTGC	GGCGCGGGGA	CCCCGGGCCG	1140
AAGAAGAGAT				ATCTCCGAAA	GAATTAAAAA	1200
TGGCCGAGAA				GCGGCCTAAA	CTCTCATCTC	1260
•			<del>-</del>			1320
CGGCCCTCTC						1380
ACGACTTACO	, MCMICHAIIP	WICHWOLDIN				





TTAATCAGCT TCAGACAAGT CTTGGCATGG TACAAGATGC AGCTTCTAAA CATAAACAGC 1440 TGTCAGAATT GCTGCGATCT GGTAGTTCCC CTAACCTCAA TATGGGAGTT GGTGGCCCAG GTCAAGTCAT GGCCAGCCAG GCCCAACAGA GCAGTCCTGG ATTAGGTTTG ATAAATAGCA 1560 TGGTCAAAAG CCCAATGACA CAGGCAGGCT TGACTTCTCC CAACATGGGG ATGGGCACTA GTGGACCAAA TCAGGGTCCT ACGCAGTCAA CAGGTATGAT GAACAGTCCA GTAAATCAGC CTGCCATGGG AATGAACACA GGGACGAATG CGGGCATGAA TCCTGGAATG TTGGCTGCAG 1740 GCAATGGACA AGGGATAATG CCTAATCAAG TCATGAACGG TTCAATTGGA GCAGGCCGAG 1800 GGCGACAGGA TATGCAGTAC CCAAACCCAG GCATGGGAAG TGCTGGCAAC TTACTGACTG 1860 AGCCTCTTCA GCAGGGCTCT CCCCAGATGG GAGGACAAAC AGGATTGAGA GGCCCCCAGC 1920 CTCTTAAGAT GGGAATGATG AACAACCCCA ATCCTTATGG TTCACCATAT ACTCAGAATC CTGGACAGCA GATTGGAGCC AGTGGCCTTG GTCTCCAGAT TCAGACAAAA ACTGTACTAT 2040 CAAATAACTT ATCTCCATTT GCTATGGACA AAAAGGCAGT TCCTGGTGGA GGAATGCCCA 2100 ACATGGGTCA ACAGCCAGCC CCGCAGGTCC AGCAGCCAGG TCTGGTGACT CCAGTTGCCC 2160 AAGGGATGGG TTCTGGAGCA CATACAGCTG ATCCAGAGAA GCGCAAGCTC ATCCAGCAGC AGCTTGTTCT CCTTTTGCAT GCTCACAAGT GCCAGCGCCG GGAACAGGCC AATGGGGAAG TGAGGCAGTG CAACCTTCCC CACTGTCGCA CAATGAAGAA TGTCCTAAAC CACATGACAC ACTGCCAGTC AGGCAAGTCT TGCCAAGTGG CACACTGTGC ATCTTCTCGA CAAATCATTT 2340 2400 CACACTGGAA GAATTGTACA AGACATGATT GTCCTGTGTG TCTCCCCCTC AAAAATGCTG 2460 GTGATAAGAG AAATCAACAG CCAATTTTGA CTGGAGCACC CGTTGGACTT GGAAATCCTA 2520 GCTCTCTAGG GGTGGGTCAA CAGTCTGCCC CCAACCTAAG CACTGTTAGT CAGATTGATC 2580 CCAGCTCCAT AGAAAGAGCC TATGCAGCTC TTGGACTACC CTATCAAGTA AATCAGATGC 2640 CGACACAACC CCAGGTGCAA GCAAAGAACC AGCAGAATCA GCAGCCTGGG CAGTCTCCCC 2700 AAGGCATGCG GCCCATGAGC AACATGAGTG CTAGTCCTAT GGGAGTAAAT GGAGGTGTAG 2760 GAGTTCAAAC GCCGAGTCTT CTTTCTGACT CAATGTTGCA TTCAGCCATA AATTCTCAAA ACCCAATGAT GAGTGAAAAT GCCAGTGTGC CCTCCCTGGG TCCTATGCCA ACAGCAGCTC 2880 AACCATCCAC TACTGGAATT CGGAAACAGT GGCACGAAGA TATTACTCAG GATCTTCGAA ATCATCTTGT TCACAAACTC GTCCAAGCCA TATTTCCTAC GCCGGATCCT GCTGCTTTAA 2940 3000 AAGACAGACG GATGGAAAAC CTAGTTGCAT ATGCTCGGAA AGTTGAAGGG GACATGTATG AATCTGCAAA CAATCGAGCG GAATACTACC ACCTTCTAGC TGAGAAAATC TATAAGATCC AGAAAGAACT AGAAGAAAAA CGAAGGACCA GACTACAGAA GCAGAACATG CTACCAAATG CTGCAGGCAT GGTTCCAGTT TCCATGAATC CAGGGCCTAA CATGGGACAG CCGCAACCAG GAATGACTTC TAATGGCCCT CTACCTGACC CAAGTATGAT CCGTGGCAGT GTGCCAAACC 3300 AGATGATGCC TCGAATAACT CCACAATCTG GTTTGAATCA ATTTGGCCAG ATGAGCATGG CCCAGCCCC TATTGTACCC CGGCAAACCC CTCCTCTTCA GCACCATGGA CAGTTGGCTC 3420 AACCTGGAGC TCTCAACCCG CCTATGGGCT ATGGGCCTCG TATGCAACAG CCTTCCAACC AGGGCCAGTT CCTTCCTCAG ACTCAGTTCC CATCACAGGG AATGAATGTA ACAAATATCC 3480 3540 CTTTGGCTCC GTCCAGCGGT CAAGCTCCAG TGTCTCAAGC ACAAATGTCT AGTTCTTCCT 3600 GCCCGGTGAA CTCTCCTATA ATGCCTCCAG GGTCTCAGGG GAGCCACATT CACTGTCCCC AGCTTCCTCA ACCAGCTCTT CATCAGAATT CACCCTCGCC TGTACCTAGT CGTACCCCCA CCCCTCACCA TACTCCCCCA AGCATAGGGG CTCAGCAGCC ACCAGCAACA ACAATTCCAG 3780 CCCCTGTTCC TACACCACCA GCCATGCCAC CTGGGCCACA GTCCCAGGCT CTACATCCCC 3840 CTCCAAGGCA GACACCTACA CCACCAACAA CACAACTTCC CCAACAAGTG CAGCCTTCAC 3900
TTCCTGCTGC ACCTTCTGCT GACCAGCCCC AGCAGCAGCC TCGCTCACAG CAGAGCACAG 3960
CAGCGTCTGT TCCTACCCCA AACGCACCGC TGCTTCCTCC GCAGCCTGCA ACTCCACTTT 4020
CCCAGCCAGC TGTAAGCATT GAAGGACAGG TATCAAATCC TCCATCTACT AGTAGCACAG 4080 AAGTGAATTC TCAGGCCATT GCTGAGAAGC AGCCTTCCCA GGAAGTGAAG ATGGAGGCCA 4140 AAATGGAAGT GGATCAACCA GAACCAGCAG ATACGCAGCC GGAGGATATT TCAGAGTCTA AAGTGGAAGA CTGTAAAATG GAATCTACCG AAACAGAAGA GAGAAGCACT GAGTTAAAAA 4260 CTGAAATAAA AGAGGAGGAA GACCAGCCAA GTACTTCAGC TACCCAGTCA TCTCCGGCTC 4320 CAGGACAGTC AAAGAAAAAG ATTTTCAAAC CAGAAGAACT ACGACAGGCA CTGATGCCAA 4380 CATTGGAGGC ACTTTACCGT CAGGATCCAG AATCCCTTCC CTTTCGTCAA CCTGTGGACC CTCAGCTTTT AGGAATCCCT GATTACTTTG ATATTGTGAA GAGCCCCATG GATCTTTCTA 4500 CCATTAAGAG GAAGTTAGAC ACTGGACAGT ATCAGGAGCC CTGGCAGTAT GTCGATGATA 4560 TTTGGCTTAT GTTCAATAAT GCCTGGTTAT ATAACCGGAA AACATCACGG GTATACAAAT 4620 ACTGCTCCAA GCTCTCTGAG GTCTTTGAAC AAGAAATTGA CCCAGTGATG CAAAGCCTTG 4680 GATACTGTTG TGGCAGAAG TTGGAGTTCT CTCCACAGAC ACTGTGTTGC TACGGCAAAC AGTTGTGCAC AATACCTCGT GATGCCACTT ATTACAGTTA CCAGAACAGG TATCATTTCT GTGAGAAGTG TTTCAATGAG ATCCAAGGGG AGAGCGTTTC TTTGGGGGAT GACCCTTCCC 4860 AGCCTCAAAC TACAATAAAT AAAGAACAAT TTTCCAAGAG AAAAAATGAC ACACTGGATC 4920 CTGAACTGTT TGTTGAATGT ACAGAGTGCG GAAGAAAGAT GCATCAGATC TGTGTCCTTC 4980 ACCATGAGAT CATCTGGCCT GCTGGATTCG TCTGTGATGG CTGTTTAAAG AAAAGTGCAC GAACTAGGAA AGAAAATAAG TTTTCTGCTA AAAGGTTGCC ATCTACCAGA CTTGGCACCT 5100 TTCTAGAGAA TCGTGTGAAT GACTTTCTGA GGCGACAGAA TCACCCTGAG TCAGGAGAGG 5160





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тедететта	AGTAGTTCAT	GCTTCTGACA	AAACCGTGGA	AGTAAAACCA	GGCATGAAAG	5220
CDDGGTTTGT	GGACAGTGGA	GAGATGGCAG	AATCCTTTCC	ATACCGAACC	AAAGCCCTCT	5280
TTGCCTTTGA	AGAAATTGAT	GGTGTTGACC	TGTGCTTCTT	TGGCATGCAT	GTTCAAGAGT	5340
ATGGCTCTGA	CTGCCCTCCA	CCCAACCAGA	GGAGAGTATA	CATATCTTAC	CTCGATAGTG	5400
<b>ጥተር እጥጥጥርጥጥ</b>	CCGTCCTAAA	TGCTTGAGGA	CTGCAGTCTA	TCATGAAATC	CTAATTGGAT	5460
DTTTDGDDTA	TGTCAAGAAA	TTAGGTTACA	CAACAGGGCA	TATTTGGGCA	TGTCCACCAA	5520
GTGAGGGAGA	TGATTATATC	TTCCATTGCC	ATCCTCCTGA	CCAGAAGATA	CCCAAGCCCA	5580
DECEDETECA	GGAATGGTAC	AAAAAAATGC	TTGACAAGGC	TGTATCAGAG	CGTATTGTCC	5640
AGCGACIGCA	GGATATTTT	AAACAAGCTA	CTGAAGATAG	ATTAACAAGT	GCAAAGGAAT	5700
MCCCTTTTTTT	CGAGGGTGAT	TTCTGGCCCA	ATGTTCTGGA	AGAAAGCATT	AAGGAACTGG	5760
ANCHOCAGGA	AGAAGAGAGA	AAACGAGAGG	AAAACACCAG	CAATGAAAGC	ACAGATGTGA	5820
CCDDGGGAGA	CAGCAAAAAT	GCTAAAAAGA	AGAATAATAA	GAAAACCAGC	AAAAATAAGA	5880
CCAAGGGAGA	TAGGGGCAAC	AAGAAGAAAC	CCGGGATGCC	CAATGTATCT	AACGACCTCT	5940
CACACAAAACT	ATATGCCACC	ATGGAGAAGC	ATAAAGAGGT	CTTCTTTGTG	ATCCGCCTCA	6000
THECHE	TGCTGCCAAC	TCCCTGCCTC	CCATTGTTGA	TCCTGATCCT	CTCATCCCCT	6060
CCCDTCTCDT	GGATGGTCGG	GATGCGTTTC	TCACGCTGGC	AAGGGACAAG	CACCTGGAGT	6120
CCGATCTGAT	CCGAAGAGCC	CAGTGGTCCA	CCATGTGCAT	GCTGGTGGAG	CTGCACACGC	6180
TCTCTTCACT	CCGCTTTGTC	TACACCTGCA	ATGAATGCAA	GCACCATGTG	GAGACACGCT	6240
AGAGCCAGGA	TGTCTGTGAG	CATTATCACT	TGTGTATCAC	CTGCTATAAC	ACTAAAAACC	6300
BECACTGIAC	AATGGAGAAA	CTAGGCCTTG	GCTTAGATGA	TGAGAGCAAC	AACCAGCAGG	6360
ATGACCACAA	CCAGAGCCCA	GGCGATTCTC	GCCGCCTGAG	TATCCAGCGC	TGCATCCAGT	6420
CTGCAGCCAC	TGCTTGCCAG	TETCEGAATG	CCAATTGCTC	ACTGCCATCC	TGCCAGAAGA	6480
CTCTGGTCCA	TGTGCAGCAT	ACCAAGGGTT	GCAAACGGAA	AACCAATGGC	GGGTGCCCCA	6540
TGAAGCGGGI	GCTCATTGCC	CTCTCCTCCT	ACCATGCCAA	GCACTGCCAG	GAGAACAAAT	6600
TCTGCAAGCA	GTTCTGCCTA	DACATCAAGC	AGAAGCTCCG	GCAGCAACAG	CTGCAGCACC	6660
GCCCGGIGCC	GGCCCAAATG	CTTCGCAGGA	GGATGGCCAG	CATGCAGCGG	ACTGGTGTGG	6720
GACTACAGCA	ACAGGGCCTC	CCTTCCCCCA	CTCCTGCCAC	TCCAACGACA	CCAACTGGCC	6780
TIGGGCAGCA	CACCCGCAG	ACGCCCCAGC	CCACTTCTCA	GCCTCAGCCT	ACCCCTCCCA	6840
AACAGCCAAC	ACCCTACTTG	CCCAGGACTC	AAGCTGCTGG	CCCTGTGTCC	CAGGGTAAGG	6900
ATAGCATGCC	GGTGACCCCT	CCARCCCTC	CTCAGACTGC	TCAGCCACCC	CTTCCAGGGC	6960
CAGCAGGCCA	AGCAGTGGAA	DTGGCDDTGC	AGATTCAGAG	AGCAGCGGAG	ACGCAGCGCC	7020
CCCCACCTAC	CGTGCAAATT	TTTCDDDGGC	CARTCCARCA	CCAGATGCCC	CCGATGACTC	7080
AGATGGCCCA	CATGGGTATG	AACCCACCTC	CCATGACCAG	AGGTCCCAGT	GGGCATTTGG	7140
CCATGGCCCC	GGGACCGACA	GGGATGCAGC	DACAGCCACC	CTGGAGCCAA	GGAGGATTGC	7200
AGCCAGGGAT	GGGACCGACA	TCTCCCATCC	CAAGGCCAGC	CATGATGTCA	GTGGCCCAGC	7260
CTCAGCCCCA	TTTGAACATG	GCTCCACAAC	CAGGATTGGG	CCAGGTAGGT	ATCAGCCCAC	7320
ATGGTCAACC	CACTGTGTCT	CDDCDAGCCT	TACAAAACCT	TTTGCGGACT	CTCAGGTCTC	7380
TCAAACCAGG	CACIGIGICI	CAACAGGTGC	TTAGTATCCT	TCACGCCAAC	CCCCAGCTGT	7440
TCAGCICICC	CATCAAGCAG	CGGGCTGCCA	AGTATGCCAA	CTCTAATCCA	CAACCCATCC	7500
TGGCTGCALT	TGGCATGCCC	CAGGGGCAGC	CAGGGCTACA	GCCACCTACC	ATGCCAGGTC	7560
TIGGGCAGCC	CCNCTCCNAT	CCAGCCATGC	AGAACATGAA	TCCAATGCAG	GCGGGCGTTC	7620
AGCAGGGGG	CCACICCAAI	CAGCAACCAC	AGCAGCAACT	CCAGCCACCC	ATGGGAGGGA	7680
MCAGGGGC FGG	CCTCACCAC	ATGAACATGA	ACCACAACAC	CATGCCTTCA	CAATTCCGAG	7740
A CAMCCOCCA	ACCACACCA	ATGATGCAAC	AGCAGCAGCA	ACAGGGAGCA	GGGCCAGGAA	7800
MUNICITIES	AATEGCCAAC	CATAACCAGT	TCCAGCAACC	CCAAGGAGTT	GGCTACCCAC	7860
ChChGCCGC	CCACCGGATG	CAGCATCACA	TGCAACAGAT	GCAACAAGGA	AATATGGGAC	7920
ACAMB GGCCI	CCTTCCCCA	GCCTTGGGAG	CAGAGGCAG	TGCCAGTCTA	CAGGCCTATC	7980
AGATAGGCCA	CCTTCAGCAE	CAGATGGGGT	CCCCTGTTCA	GCCCAACCCC	ATGAGCCCCC	8040
AGCAGCGAC A	r correctabal	CAGGCCCAGT	CCCCACACC"	ACAAGGCCAG	CAGATCCCTA	8100
AGCAGCAIA	CARTCARGE		AGCCTGTCC	TTCTCCACGO	CCACAGTCCC	8160
ACCCCCCCC	CARICAROIC	TCCCCAAGGA	TGCAGCCTCA	GCCTTCTCC	CACCACGTTT	.8220
AGCCACACA	T ANGTITUTE	CATCCTGGA	TGGTAGCTG	CCAGGCCAAC	CCCATGGAAC	8280
T T C C C T T T T	TECCAECCC	GACCAGAATT	CAATGCTTT	TCAGCTTGCT	r AGCAATCCAG	8340
AAGGGCAII	CCTCCATGG	r GCAAGCGCC	A CGGACCTGG	ACTCAGCAC	GATAACTCAG	8400
ACMAGA VALA	C DEFICITION	CAGAGTACA	TAGACATAC	A CTAGAGACA	C CTTGTATTTT	8460
MCT I GWWT I	ייידיים ייים מיים מיים מיים	r CTCTTAACA	A GACTTTTTG'	T ACTGAAAAC	A ATTTTTTGA	8520
A TOUR TOUR COM	D CCCTDDDDC	A CAATTTTCC	T TGGAACACA'	T AAGAACTGT	G CAGTAGCCGT	8580
ATCTTTCGT	U GCCIMANG	T GCAAGATGA	A CCTGAGGGA	T GATAGAATA	CAAAGAATATA	8640
TIGIGGITI.	~ ~~~~~~~~~ ~ ~~~~~~~~~~~~~~~~~~~~~~~~	A CCACCAGCC	T TTCTTCCCC	T TTGTGTGTG	r ggttcaagtg	8700
TECNETIC	D GGAGGGCTGA	G GCCTGTGAA	G CCAAACAAT	A TGCTCCTGC	C TTGCACCTCC	8760
AATAGGTTT	T TTTTTTTTT	T TTAAATTAA	T GAACATATG	T AATATTAAT	G AACATATGTA	8820
מיים מייים מיי	G TTATTATT	A CTGGTGCAG	A TGGTTGACA	T TTTTCCCTA	T TTTCCTCACT	8880
. AIAIIAAIA	A GTTAAAACA	T TTCTAAACC	A GAGGACAAA	A GGGGTTAAT	G TTACTTTGAA	8940
ITATGGAAG	Z OTTAGE					





TCTGGGTGCA AAGATGTTCA TTCTTTTAAA AAATGTTTAA AAAAAA

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9046

# (2) INFORMATION FOR SEQ ID NO:13:

#### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7326 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

	ATGGCCGAGA	ACTTGCTGGA	CGGACCGCCC	AACCCCAAAC	GAGCCAAACT	CAGCTCGCCC	60
	GGCTTCTCCG	CGAATGACAA	CACAGATTTT	GGATCATTGT	TTGACTTGGA	AAATGACCTT	120
	CCTGATGAGC	TGATCCCCAA	TGGAGAATTA	AGCCTTTTAA	ACAGTGGGAA	CCTTGTTCCA	180
	GATGCTGCGT	CCAAACATAA	ACAACTGTCA	GAGCTTCTTA	GAGGAGGCAG	CGGCTCTAGC	240
	ATCAACCCAG	GGATAGGCAA	TGTGAGTGCC	AGCAGCCCTG	TGCAACAGGG	CCTTGGTGGC	300
	CAGGCTCAGG	GGCAGCCGAA	CAGTACAAAC	ATGGCCAGCT	TAGGTGCCAT	GGGCAAGAGC	360
	CCTCTGAACC	AAGGAGACTC	ATCAACACCC	AACCTGCCCA	AACAGGCAGC	CAGCACCTCT	420
	GGGCCCACTC	CCCCTGCCTC	CCAAGCACTG	AATCCACAAG	CACAAAAGCA	AGTAGGGCTG	480
	GTGACCAGTA	GTCCTGCCAC	ATCACAGACT	GGACCTGGGA	TCTGCATGAA	TGCTAACTTC	540
	AACCAGACCC	ACCCAGGCCT	TCTCAATAGT	AACTCTGGCC	ATAGCTTAAT	GAATCAGGCT	600
	CAACAAGGGC	AAGCTCAAGT	CATGAATGGA	TCTCTTGGGG	CTGCTGGAAG	AGGAAGGGGA	660
	GCTGGAATGC	CCTACCCTGC	TCCAGCCATG	CAGGGGGCCA	CAAGCAGTGT	GCTGGCGGAG	720
	ACCTTGACAC	AGGTTTCCCC	ACAAATGGCT	GGCCATGCTG	GACTAAATAC	AGCACAGGCA	780
	GGAGGCATGA	CCAAGATGGG	AATGACTGGT	ACCACAAGTC	CATTTGGACA	ACCCTTTAGT	840
	CAAACTGGAG	GGCAGCAGAT	GGGAGCCACT	GGAGTGAACC	CCCAGTTAGC	CAGCAAACAG	900
	AGCATGGTCA	ATAGTTTACC	TGCTTTTCCT	ACAGATATCA	AGAATACTTC	AGTCACCACT	960
	GTGCCAAATA	TGTCCCAGTT	GCAAACATCA	GTGGGAATTG	TACCCACACA	AGCAATTGCA	1020
	ACAGGCCCCA	CAGCAGACCC	TGAAAAACGC	AAACTGATAC	AGCAGCAGCT	GGTTCTACTG	1080
	CTTCATGCCC	ACAAATGTCA	GAGACGAGAG	CAAGCAAATG	GAGAGGTTCG	NGCCTGTTCT	1140
	CTCCCACACT	GTCGAACCAT	GAAAAACGTT	TTGAATCACA	TGACACATTG	TCAGGCTCCC	1200
	AAAGCCTGCC	AAGTTGCCCA	TTGTGCATCT	TCACGACAAA	TCATCTCTCA	TTGGAAGAAC	1260
	TGCACACGAC	ATGACTGTCC	TGTTTGCCTC	CCTTTGAAAA	ATGCCAGTGA	CAAGCGAAAC	1320
	CAACAAACCA	TCCTGGGATC	TCCAGCTAGT	GGAATTCAAA	ACACAATTGG	TTCTGTTGGT	1380
	GCAGGGCAAC	AGAATGCCAC	TTCCTTAAGT	AACCCAAATC	CCATAGACCC	CAGTTCCATG	1440
	CAGCGGGCCT	ATGCTGCTCT	AGGACTCCCC	TACATGAACC	AGCCTCAGAC	GCAGCTGCAG	1500
	CCTCAGGTTC	CTGGCCAGCA	ACCAGCACAG	CCTCCAGCCC	ACCAGCAGAT	GAGGACTCTC	1560
	AATGCCCTAG	GAAACAACCC	CATGAGTGTC	CCAGCAGGAG	GAATAACAAC	AGATCAACAG	1620
	CCACCAAACT	TGATTTCAGA	ATCAGCTCTT	CCAACTTCCT	TGGGGGCTAC	CAATCCACTG	1680
	ATGAATGATG	GTTCAAACTC	TGGTAACATT	GGAAGCCTCA	GCACGATACC	TACAGCAGCG	1740
	CCTCCTTCCA	GCACTGGTGT	TCGAAAAGGC	TGGCATGAAC	ATGTGACTCA	GGACCTACGG	1800
	AGTCATCTAG	TCCATAAACT	CGTTCAAGCC	ATCTTCCCAA	CTCCAGACCC	TGCAGCTCTG	1860
	AAAGATCGCC	GCATGGAGAA	CCTGGTTGCC	TATGCTAAGA	AAGTGGAGGG	AGACATGTAT	1920
	GAGTCTGCTA	ATAGCAGGGA	TGAATACTAT	CATTTATTAG	CAGAGAAAAT	CTATAAAATA	1980
	CAAAAAGAAC	TAGAAGAAAA	GCGGAGGACA	CGTTTACATA	AGCAAGGCAT	CCTGGGTAAC	2040
•	CAGCCAGCTT	TACCAGCTTC	TGGGGCTCAG	CCCCCTGTGA	TTCCACCAGC	CCAGTCTGTA	2100
	AGACCTCCAA	ATGGGCCCCT	GCCTTTGCCA	GTGAATCGCA	TGCAGGTTTC	TCAAGGGATG	2160
	AATTCATTTA	ACCCAATGTC	CCTGGGAAAC	GTCCAGTTGC	CACAGGCACC	CATGGGACCT	2220
	CGTGCAGCCT	CCCCTATGAA	CCACTCTGTG	CAGATGAACA	GCATGGCCTC	AGTTCCGGGT	2280
	ATGGCCATTT	CTCCTTCACG	GATGCCTCAG	CCTCCAAATA	TGATGGGCAC	TCATGCCAAC	2340
	AACATTATGG	CCCAGGCACC	TACTCAGAAC	CAGTTTCTGC	CACAGAACCA	GTTTCCATCA	2400
	TCCAGTGGGG	CAATGAGTGT	GAACAGTGTG	GGCATGGGGC	AACCAGCAGC	CCAGGCAGGT	2460
	GTTTCACAGG	GTCAGGAACC	TGGAGCTGCT	CTCCCTAACC	CTCTGAACAT	GCTGGCACCC	2520
	CAGGCCAGCC	AGCTGCCTTG	CCCACCAGTG	ACACAGTCAC	CATTGCACCC	GACTCCACCT	. 2580
			CATGCCCTCT				2640
	CCTCCTCAGC	CAGCAGCTCC	CACTCAGCCA	TCTACTCCTG	TGTCATCTGG	GCAGACTCCT	2700
	ACCCCAACTC	CTGGCTCAGT	GCCCAGCGCT	GCCCAAACAC	AGAGTACCCC	TACAGTCCAG	2760
	GCAGCAGCAC	AGGCTCAGGT	GACTCCACAG	CCTCAGACCC	CAGTGCAGCC	ACCATCTGTG	2820
			GCAGCAACCA				2880
	CCGCTTTCTC	AGGCAGCAGC	CAGCATTGAT	AATAGAGTCC	CTACTCCCTC	CACTGTGACC	2940





AGTGCTGAAA CCAGTTCCCA GCAGCCAGGA CCCGATGTGC CCATGCTGGA AATGAAGACA 3000 GAGGTGCAGA CAGATGATGC TGAGCCTGAA CCTACTGAAT CCAAGGGGGA ACCTCGGTCT 3060 GAGATGATGG AAGAGGATTT ACAAGGTTCT TCCCAAGTAA AAGAAGAGAC AGATACGACA 3120 GAGCAGAAGT CAGAGCCAAT GGAAGTAGAA GAAAAGAAAC CTGAAGTAAA AGTGGAAGCT 3180 AAAGAGGAAG AAGAGAACAG TTCGAACGAC ACAGCCTCAC AATCAACATC TCCTTCCCAG 3240 CCACGCAAAA AAATCTTTAA ACCCGAGGAG CTACGCCAGG CACTTATGCC AACTCTAGAA 3300 GCACTCTATC GACAGGACCC AGAGTCTTTG CCTTTTCGTC AGCCTGTAGA TCCTCAGCTC 3360 CTAGGAATCC CAGATTATTT TGATATAGTG AAGAATCCTA TGGACCTTTC TACCATCAAA 3420 CGAAAGCTGG ACACAGGGCA ATATCAAGAA CCCTGGCAGT ATGTGGATGA TGTCAGGCTT ATGTTCAACA ATGCGTGGCT ATATAATCGT AAAACGTCCC GTGTATATAA ATTTTGCAGT 3540 AAACTTGCAG AGGTCTTTGA ACAAGAAATT GACCCTGTCA TGCAGTCTCT TGGATATTGC 3600 TGTGGACGAA AGTATGAGTT CTCCCCACAG ACTTTGTGCT GTTACGGAAA GCAGCTGTGT 3660 ACAATTCCTC GTGATGCAGC CTACTACAGC TATCAGAATA GGTATCATTT CTGTGGGAAG 3720 TGTTTCACAG AGATCCAGGG CGAGAATGTG ACCCTGGGTG ACGACCCTTC CCAACCTCAG 378C ACGACAATTT CCAAGGATCA ATTTGAAAAG AAGAAAAATG ATACCTTAGA TCCTGAACCT 3840 TTTGTTGACT GCAAAGAGTG TGGCCGGAAG ATGCATCAGA TTTGTGTTCT ACACTATGAC 3900 ATCATTTGGC CTTCAGGTTT TGTGTGTGAC AACTGTTTGA AGAAAACTGG CAGACCTCGG 3960 AAAGAAAACA AATTCAGTGC TAAGAGGCTG CAGACCACAC GATTGGGAAA CCACTTAGAA 4020 GACAGAGTGA ATAAGTTTTT GCGGCGCCAG AATCACCCTG AAGCTGGGGA GGTTTTTGTC 4080 AGAGTGGTGG CCAGCTCAGA CAAGACTGTG GAGGTCAAGC CGGGAATGAA GTCAAGGTTT 4140 GTGGATTCTG GAGAGATGTC GGAATCTTTC CCATATCGTA CCAAAGCACT CTTTGCTTTT 4200 GAGGAGATCG ATGGAGTCGA TGTGTGCTTT TTTGGGATGC ATGTGCAAGA TACGGCTCTG 4260 ATTGCCCCC ACCAATACA AGGCTGTGTA TACATATCTT ATCTGGACAG TATTCATTTC 4320 TTCCGGCCCC GCTGCCTCCG GACAGCTGTT TACCATGAGA TCCTCATCGG ATATCTCGAG 4380 TATGTGAAGA AATTGGTGTA TGTGACAGCA CATATTTGGG CCTGTCCCCC AAGTGAAGGA 4440 GATGACTATA TCTTTCATTG CCACCCCCT GACCAGAAAA TCCCCAAACC AAAACGACTA 4500 CAGGAGTGGT ACAAGAAGAT GCTGGACAAG GCGTTTGCAG AGAGGATCAT TAACGACTAT 4560 AAGGACATCT TCAAACAAGC GAACGAAGAC AGGCTCACGA GTGCCCAAGGA GTTGCCCTAT 4620 TTTGAAGGAG ATTTCTGGCC TAATGTGTTG GAAGAAAGCA TTAAGGAACT AGAACAAGAA 4680 GAAGAAGAAA GGAAAAAAGA AGAGAGTACT GCAGCGAGTG AGACTCCTGA GGGCAGTCAG 4740 GGTGACAGCA AAAATGCGAA GAAAAAGAAC AACAAGAAGA CCAACAAAAA CAAAAGCAGC 4800 ATTAGCCGCG CCAACAAGAA GAAGCCCAGC ATGCCCAATG TTTCCAACGA CCTGTCGCAG 4860 AAGCTGTATG CCACCATGGA GAAGCACAAG GAGGTATTCT TTGTGATTCA TCTGCATGCT 4920 GGGCCTGTTA TCAGCACTCA GCCCCCCATC GTGGACCCTG ATCCTCTGCT TAGCTGTGAC 4980 CTCATGGATG GGCGAGATGC CTTCCTCACC CTGGCCAGAG ACAAGCACTG GGAATTCTCT 5040 TCCTTACGCC GCTCCAAATG GTCCACTCTG TGCATGCTGG TGGAGCTGCA CACACAGGGC 5100 CAGGASCGCT TTGTTTATAC CTGCAATGAG TGCAAACACC ATGTGGAAAC ACGCTGGCAC TGCACTGTGT GTGAGGACTA TGACCTTTGT ATCAATTGCT ACAACACAAA GAGCCACACC 5220 CATAAGATGG TGAAGTGGGG GCTAGGCCTA GATGATGAGG GCAGCAGTCA GGGTGAGCCA 5280 CAGTCCAAGA GCCCCCAGGA ATCCCGGCGT CTCAGCATCC AGCGCTGCAT CCAGTCCCTG 5340 GTGCATGCCT GCCAGTGTCG CAATGCCAAC TGCTCACTGC CGTCTTGCCA GAAGATGAAG 5400 CGAGTCGTGC AGCACCCAA GGGCTGCAAG CGCAAGACTA ATGGAGGATG CCCAGTGTGC 5460 AAGCAGCTCA TTGCTCTTTG CTGCTACCAC GCCAAACACT GCCAAGAAAA TAAATGCCCT 5520 GTGCCCTTCT GCCTCAACAT CAAACATAAC GTCCGCCAGC AGCAGATCCA GCACTGCCTG 5580 CAGCAGGCTC AGCTCATGCG CCGGCGAATG GCAACCATGA ACACCCGCAA TGTGCCTCAG 5640 CAGAGTTTGC CTTCTCCTAC CTCAGCACCA CCCGGGACTC CTACACAGCA GCCCAGCACA CCCCAAACAC CACAGCCCC AGCCCAGCCT CAGCCTTCAC CTGTTAACAT GTCACCAGCA 5760 GGCTTCCCTA ATGTAGCCCG GACTCAGCCC CCAACAATAG TGTCTGCTGG GAAGCCTACC 5820 AACCAGGTGC CAGCTCCCCC ACCCCCTGCC CAGCCCCCAC CTGCAGCAGT AGAAGCAGCC 5880 CGGCAAATTG AACGTGAGGC CCAGCAGCAG CAGCACCTAT ACCGAGCAAA CATCAACAAT 5940 GGCATGCCCC CAGGACGTGA CGGTATGGGG ACCCCAGGAA GCCAAATGAC TCCTGTGGGC 6000 CTGAATGTGC CCCGTCCCAA CCAAGTCAGT GGGCCTGTCA TGTCTAGTAT GCCACCTGGG 6060 CAGTGGCAGC AGGCACCCAT CCCTCAGCAG CAGCCGATGC CAGGCATGCC CAGGCCTGTA 6120 ATGTCCATGC AGGCCCAGGC AGCAGTGGCT GGGCCACGGA TGCCCAATGT GCAGCCAAAC 6180 AGGAGCATCT CGCCAAGTGC CCTGCAAGAC CTGCTACGGA CCCTAAAGTC ACCCAGCTCT 6240 CCTCAGCAGC AGCAGCAGGT GCTGAACATC CTTAAATCAA ACCCACAGCT AATGGCAGCT 6300 TTCATCAAAC AGCGCACAGC CAAGTATGTG GCCAATCAGC CTGGCATGCA GCCCCAGCCC 6360 GGACTTCAAT CCCAGCCTGG TATGCAGCCC CAGCCTGGCA TGCACCAGGA GCCTAGTTTG 6420 CAAAACCTGA ACGCAATGCA AGCTGGTGTG CCACGGCCTG GTGTGCCTCC ACCACAACCA 6480 GCAATGGGAG GCCTGAATCC CCAGGGACAA GCTCTGAACA TCATGAACCC AGGACACAAC 6540 CCCAACATGA CAAACATGAA TCCACAGTAC CGAGAAATGG TGAGGAGACA GCTGCTACAG 6600 CACCAGCAGC AGCAGCAGCA ACAGCAGCAG CAGCAGCAGC AACAACAAAA TAGTGCCAGC 6660 TTGGCCGGGG GCATGGCGGG ACACAGCCAG TTCCAGCAGC CACAAGGACC TGGAGGTTAT

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GCCCCAGCCA	TGCAGCAGCA	ACGCATGCAA	CAGCACCTCC	CCATCCAGGG	CAGCTCCATG	6780
GGCCAGATGG	CTGCTCCAAT	GGGACAACTT	GGCCAGATGG	GGCAGCCTGG	GCTAGGGGCA	6840
GACAGCACCC	CTAATATCCA	GCAGGCCCTG	CAGCAACGGA	TTCTGCAGCA	GCAGCAGATG	6900
AAGCAACAAA	TTGGGTCACC	AGGCCAGCCG	AACCCCATGA	GCCCCCAGCA	GCACATGCTC	6960
TCAGGACAGC	CACAGGCCTC	ACATCTCCCT	GGCCAGCAGA	TCGCCACATC	CCTTAGTAAC	7020
CAGGTGCGAT	CTCCAGCCCC	TGTGCAGTCT	CCACGGCCCC	AATCCCAACC	TCCACATTCC	7080
AGCCCGTCAC	CACGGATACA	ACCCCAGCCT	TCACCACACC	ATGTTTCACC	CCAGACTGGA	7140
ACCCCTCACC	CTGGACTCGC	AGTCACCATG	GCCAGCTCCA	TGGATCAGGG	ACACCTGGGG	7200
AACCCTGAAC	AGAGTGCAAT	GCTCCCCCAG	CTGAATACCC	CCAACAGGAG	CGCACTGTCC	7260
AGTGAACTGT	CCCTGGTTGG	TGATACCACG	GGAGACACAC	TAGAAAAGTT	TGTGGAGGGT	7320
TTGTAG						7326

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#### (2) INFORMATION FOR SEQ ID NO:14:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 2499 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

TCACTTGTCA ATTAATCCAG CTTCCTTAAT TTTACTGAAG AAGAATTTCT CCAGGATATT GGCACATTTG TAGTATTCAC TCTCAGGGGC GTTGTACTCT TTGCAATTGG TAAAGACTCG 120 CTGTAAGTCT GCCATGAATA ATTTCTTAGA CACGTAGTAC CTATTCTTGA GGCGTTCACT 180 CATGGTTTTG AGATCCATGG GGGACCTTAT AACTTCATAA TATCCTGGAG CTTCTGTTCT 240 CTTCACAGGT TCCATGAAGG GCCAAGCGCT TTGATGGCTC TTCACCTGCT GGAGGATGCT CTTGAGCGTG CTGTAAAGCT GGTCAGGGTC TCTGGGCTCT TTACTTTTCT CTTTTCCACT CGGTTTCCAG CCTGTCTCTC TAATTCCAGG AATGCTTTCT ATAGGAATCT GTCGAACTCC ATCTTTAAAA CATGAAAGTC CAGGGTAAAC TTTTCGAATT TGTGCCTGTT TTCTTTCAAT 480 540 CAGTTTTTTA ATTATCTCCT TCTGCTTTTT AATGATGACA GAAAATTCTG TGTACGGGAT CCGTGGATTT AGCTCACATC CCATTAAAGT GGCTCCTTCA TAATCCTTGA TATAGCCAAC ATATTTGGTT TTAGGTATTT TAATTTCTTT GGAGAAACCC TGTTTCTTAA AGTATCCAAT 660 TGCATATTCA TCTGCATATG TGAGGAAGTT CAGGATGTCA TGCTTTATGT GATATTCTTT 720 CARATGATTC ATCAGGTGTG TTCCATAGCC CTTGACTTGC TCATTTGAGG TTACAGCACA 780 GAAGACAATC TCTGTGAATC CTTGAGATGG GAACATACGG AAACAGATAC CACCAATAAC ACGGCCATCT TTAATTAAAG CAAGGGTTTT GTGTTTCGGG TCAAAGACGA GCCGTGTGAT 900 GTATTCTTTT GGCATTCGGG GCAGCTGGTG GGAGAAAACG TTCTGTAGGC CAACCAGCCA 960 CATCAGGATC TTCTTGTTTG GTTTCTGGTT GAGGGAATTG CCAACCACGT GAAATTCAAT 1020 TACACCCCTG CGCTCTTCCA ACCTTGCCGC CTCATCCCTG GCCGAGTGTG CTGACAGAAA 1080 ATTGGTCTCT GGTCCAAGCA TTGCTGCAGG GTCCGTGATG GTAGACATAA CCTCGTTGAT 1140 TAATTCCATC GGAATATCCC CCATAACTCG GGGTTTCTTG GCCTCCTCCA GAACATGAGA 1200 ATCAGTCATT TTCCTCTTTT CTCCTGGGTT TGCCTCAAGT CCAGAAGAGG CTTTGCAGGC 1260 AGGACTGCTG CTCCCTGCGT TTGGCTGCTC AAGGGAAGAT GAGGTTGAAT TGTATGAAAT 1320 TGTCCCAGCC ACAGGAGGTG GATTGATAAC TGTTTGGATG CCTAGCTGGC TGGTTCTGGA 1380 AGAGGCTGAG AGAAAATCCT GATCCCAGAT GGGAGAGTTT TGACTATATA CTTCTTCTTC TAGCATGGAC AGAAATTTTG GGAAATGAGT GAGGATTAGA GTTCGTTTTT CAAGAGGCAG 1500 TTTATCTTTT TCCTGTCTTG CTTGTTCCAG GAGTTGTCGC CTCATAACAG TGAAGACCGA 1560 GCGAAGCAAT GTTCTCCCAA ACACCTGTGT GGTTTCGTAC CGAGGTAGAC TGTCGCAGAA 1620 CTGTGGCACG TTGCAGTAAC ACAGCCACCT TGTGTAGTTC TCTTTGTATC CAGAAATATC ATCATTGGGA GATCGCAGTC TTCGTTGAGA TGGTGCCTCC AGATGCCAAT AGTTGATGCG 1740 GTTTAGGAAC ATTTTTGCCA ACTCAACTAT TGTTTGCCTT TCTTTTGCTG GCAGGTGACT 1800 AAATTTGTAC TGCACAAAGT TATTCACACC CTGTTCAATG CTAGGTTTTT CAAATGGGGG 1860 TTTCTTTTCC AAAGAGCCTT CAACCACAGG TTTTCCTCTT TGTAAAATAG ACTTTCTCAA 1920 GAGCTTAAAT AGATAGAAAT AAACTTGTTT GGTATCTGCA TCTTCTTCCT TGTGGACACA 1980 GGTAAAGAGA TATTCCACAT CCAATACTAT TCCCAGGAGT CTGTTCATTT CTTCCTCTGA 2040 CACATTCTCC AGGTGGGAAA CATGAGCAGC TAGGGCATGG CTACAACTCC GACAGGATTC 2100 TGTTAGACTG ACAATTATTT GCTGCAGGTC GGCTCTGGGG GGAGTGGGTG AGGGGTTAGG 2160 GTTTTTCCAG CCATTACATT TACAAGACTC CTCGGCCTTG CAGGCGGAGT ACACTCCGAG 2220 TTTCTCCAGT TTCTTGGCCC GCGGAGCGGA GCGTAGTTGC GCTTTCTTCA CGGCGATTCG 2280 GGCCGAGCCA CCGCCTCCCG GTCCTTCGGC CGTGCCCGCT GCAGCCACTG CCGTCGCCGG 2340 ACCGCAGGCG CCCGAGCCCC CGGCGGCAGC GGCGCAGGGG GAGCCCTGCG GGGGCGCGGG 2400





CGGAAGCGCC GCAGGCTGCG GGGGCAGCGC CCCGGGCCCG GCCCCTGCCC CGGCTCCTGC 2460 2499 CCCGCAGCCG CCCGGCCAGC CTCGGACAT

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(2) INFORMATION FOR SEQ ID NO:15:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2442 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

TCACTTGTCA	ATCAACCCTG	CTTCCTTAAT	TTTACTGAAG	AAGAACTTCT	CCAGGATGCT	60
GGCGCATTTG	TAGTACTCGC	TCTCGGGAGG	GTTGTACTCC	TTGCAGTTGG	TGAACACTCG	120
TTGCAAGTCC	GCCATGAATA	ACTTCTTAGA	0, 10, 12, 10 10	CTGTTCCTGA		180
CATGGTTTTC	AGATCCATGG	GGAACCTTAT			CTTCTGTTCT	240
CTTCACTGGT	TCCATGAAAG	GCCAAGCATT	TGGATGGTTC	TTCACCTGCT	GCAGGATGTT	300
CTTGAGGGTG	CTGTAAACGT	GCTCAGGGTC	TTTGGGCTCT	TTACTTTTCT	CTTTTCCACT	360
TGGTTTCCAG	CCTGTCTCTC	TGATTCCAGG	AATGCTTTCT	ATAGGAATCT	GCCGAACTCC	420
ATCTTTGAAA	CACGAAAGTC	CAGGGTAGAC	TTTTCGAATC	TGGGCTTGTT	TTCTTTCTAT	480
CAGCTTTTTA	ATGATCTCCT	TCTGCTTTTT	AATGATGACA	GAGAACTCTG	TGTATGGGAT	540
CTGAGGGTTC	AGCTCACATC	CCATCAAAGT	GGCCCCTTCA	TAATCCTTGA	TGTAGCCAAC	600
ATATTTGGTT	TTAGGTATTT	TGATTTCTTT	GGAGAAACCC	TGCTTCTTGA		660
GGCATACTCA	TCTGCATATG	TGAGGAAGTT	GAGGATCTCG	TGCTTTATGT	GGTATTCTTT	720
GAGATGGTTC	ATCAGGTGGG	TTCCATAGCC	CTTGACTTGT	TCATTTGAGG	TTACTGCACA	780
GAAAACAATC	TCTGTGAATC	CCTGGGATGG	AAACATCCGG		CACCAATGAC	840
ACGGCCATCT	TTAATTAAAG	CAAGGGTTTT	GTGTTTCGGG	TCAAAGACGA	GCCGTGTGAT	900
GTACTCTTTG	GGCATTCTGG	GCAGCTGGTG	GGAAAACACA	TTCTGGAGGC	CCACGAGCCA	960
CATCAGGATC	TTCTTGTTTG	GTTTCTGGTT	CAGGGAGTTG	CCCACCACGT	GGAATTCAAT	1020
GACACCCCTG	CGTTCTTCCA	GCCGTGCCGC	CTCATCTCTG	GCCGAATGGG	CTGACAGAAA	1080
ATTGGTCTCT	GGTCCAAGCA			GTAGACATGA	CCTCATTGAT	1140
CAATTCCACG	GGAATATCCC	CCATCACTCG	AGATCTCTTG	GCCTCCTCGG	GAGCATGAGA	1200
GTTGTTCATT	TTCCTCTTTT	CTCCCGGGTT	TGCTTCAAGC	CCAGAAGAGC	CTCTGCATCC	1260
AGGACTTGTT	CTCCCTCCAT		ATGGGAAGTT	GAATTTGAAC	TGAACAATGC	1320
	ACAGGAGGAC		TTGGATTCCT	AGCGGGCTGG	TTCTGGAAGA	1380
GGCTGAGAGA	AAATCCTGAT	CCCAGATAGG	AGAATTTTGA		CTTCTTCCAA	1440 1500
CATGGACAGA	. AACTTTGGGA	AATGTGTGAG		CGTTTCTCAA		1560
GTCTTTTTC				ATGATGGTGA		1620
AAGCAATGTT		CCTTTGTGGT	TTCGTACCGA		CACAGAACTG	1680
CGGTACATTC		ACCACCTTGT		TTGTATCCAG	AGATGTCATC	1740
ATTGGGAGAC			AGCCTCCAGA		TGATGCGGTT	1800
CAGAAACATO	TTGGCCAGCT					1860
CTTGTACTGC					ATGGCGGCTT	1920
	GAGCCTTCA				TTCTCAAGAG GGACGCAGGT	1980
CTTGAATAGO						2040
GAAGAGGTA					• • • • • • • • • • • • • • • • • • • •	2100
ATTCTCCAA			GGCATGGCTA	CAGCTTCGAC	GGTTAGGGTT	2160
CAAACTGAC				GTAGGAGAGG		2220
CTTCCAGCC						2280
CTCCAGCTT						2340
CGAGCCGCC'				GCCACCGGCG		2400
	A GAGCTCCCG	G CAGCGGTGG		GGGGAACCGT	. eceeeecec	2442
GGGAGGCAG'	T GCTGGGGAC	c ceeccceco	AGCCTCGGCC	. AT		2442

## (2) INFORMATION FOR SEQ ID NO:16:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear





(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CCCGCCAGCC TCGGACATGC

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#### (2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 20 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

## (x1) SEQUENCE DESCRIPTION: SEQ ID NO:17:

#### CCCGCCAGCC TCGGCCATGC

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## (2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2442 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

ATGGCCGAGG	CTGGCGGGGC	CGGGTCCCCA	GCACTGCCTC	CCGCGCCCCC	GCACGGTTCC	60
CCCCGGACCC	TGGCCACCGC	TGCCGGGAGC	TCTGCTTCCT	GCGGGCCAGC	GACGCCGGTG	120
GCCGCGGCGG	GCACCGCCGA	GGGACCGGGA	GGAGGCGGCT	CGGCCCGGAT	CGCCGTGAAG	180
AAGGCGCAGT	TGCGCTCTGC	TCCGCGGGCG	AAGAAGCTGG	AGAAACTCGG	CGTGTACTCC	240
GCCTGCAAGG	CAGAGGAGTC	CTGTAAATGC	AATGGCTGGA	AGAACCCTAA	CCCCTCTCCT	300
ACTCCACCAA	GAGGAGACCT	CCAGCAGATA	ATTGTCAGTT	TGACAGAATC	CTGTCGAAGC	360
TGTAGCCATG	CCCTTGCTGC	TCACGTTTCT	CACTTGGAGA	ATGTGTCAGA	GGAAGAGATG	420
GACAGACTCC	TGGGAATTGT	GTTGGATGTG	GAGTACCTCT	TCACCTGCGT	CCACAAAGAA	480
GAAGATGCAG	ATACCAAACA	AGTGTACTTC	TACCTATTCA	AGCTCTTGAG	AAAGTCAATT	540
TTACAAAGAG	GAAAACCTGT	GGTTGAAGGC	TCCTTGGAGA	AGAAGCCGCC	ATTTGAGAAG	600
CCCAGTATTG	AACAGGGTGT	GAACAACTTC	GTGCAGTACA	AGTTTAGTCA	CTTGCCATCG	660
AAAGAGAGGC	AGACAACGAT	CGAGCTGGCC	AAGATGTTTC	TGAACCGCAT	CAACTACTGG	720
CATCTGGAGG	CTCCATCTCA	GCGGAGACTA	CGGTCTCCCA	ATGATGACAT	CTCTGGATAC	780
AAGGAAAACT	ACACAAGGTG	GTTGTGCTAC	TGCAATGTAC	CGCAGTTCTG	TGACAGCTTA	840
	AAACCACAAA			TTCGCTCGGT	CTTCACCATC	900
ATGAGACGAC	AGCTCTTGGA				TCTTGAGAAA	960
CGCACGCTTA	TCCTCACACA	TTTCCCAAAG	TTTCTGTCCA			1020
AGTCAAAATT	CTCCTATCTG	GGATCAGGAT		CCTCTTCCAG	AACCAGCCCG	1080
CTAGGAATCC	AAACAGTAAT	CAGTCCTCCT	GTTACTGGGA		CAGTTCAAAT	1140
	ATGAGCAGAT			CTGGATGCAG	AGGCTCTTCT	1200
GGGCTTGAAG	CAAACCCGGG	AGAAAAGAGG	AAAATGAACA	ACTCTCATGC	TCCCGAGGAG	1260
GCCAAGAGAT	CTCGAGTGAT		CCCGTGGAAT	TGATCAATGA		1320
ACCATCACAG	ACCCTGCAGG				AGCCCATTCG	1380
GCCAGAGATG	AGGCGGCACG		CGCAGGGGTG	TCATTGAATT	CCACGTGGTG	1440
GGCAACTCCC	TGAACCAGAA		AAGATCCTGA		GGGCCTCCAG	1500
AATGTGTTTT	CCCACCAGCT	GCCCAGAATG	CCCAAAGAGT	ACATCACACG	GCTCGTCTTT	1560
GACCCGAAAC	ACAAAACCCT		AAAGATGGCC	GTGTCATTGG	TGGTATCTGT	1620
TTCCGGATGT	TTCCATCCCA	GGGATTCACA		TCTGTGCAGT	AACCTCAAAT	1680
GAACAAGTCA	AGGGCTATGG	AACCCACCTG	ATGAACCATC	TCAAAGAATA		1740
CACGAGATCC	TCAACTTCCT	CACATATGCA		CCATCGGCTA	TTTCAAGAAG	1800
CAGGGTTTCT	CCAAAGAAAT		AAAACCAAAT	ATGTTGGCTA		1860
TATGAAGGGG		GGGATGTGAG		AGATCCCATA		1920
	TTAAAAAGCA			TGATAGAAAG		1980
CAGATTCGAA	AAGTCTACCC	TGGACTTTCG	TGTTTCAAAG	ATGGAGTTCG	GCAGATTCCT	2040



ATAGAAAGCA TTCCTGGAAT	CAGAGAGACA	GGCTGGAAAC	CAAGTGGAAA	AGAGAAAAGT	2100
AAAGAGCCCA AAGACCCTGA	GCACGTTTAC	AGCACCCTCA	AGAACATCCT	GCAGCAGGTG	2160
AAGAACCATC CAAATGCTTG	GCCTTTCATG	GAACCAGTGA	AGAGAACAGA	AGCTCCGGGA	2220
TATTATGAAG TTATAAGGTT	CCCCATGGAT	CTGAAAACCA	TGAGTGAACG	CCTCAGGAAC	2280
AGGTACTATG TGTCTAAGAA	GTTATTCATG	GCGGACTTGC	AACGAGTGTT	CACCAACTGC	2340
AAGGAGTACA ACCCTCCCGA	GAGCGAGTAC	TACAAATGCG	CCAGCATCCT	GGAGAAGTTC	2400
TTCTTCAGTA AAATTAAGGA	DECAGGETTE	ATTGACAAGT	GA .		2442

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### What is claimed is:

- 1. A purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones.
- 2. The protein of claim 1 consisting of the amino acid sequence of SEQ ID NO:1.
- 3. The protein of claim 1 comprising the amino acid sequence of SEQ ID NO:2.
- 4. The protein of claim 1, which also binds to the amino acid sequence of SEQ ID NO:3 on a p300 cellular protein and to amino acid residues 1805-1854 of a CBP cellular protein (SEQ ID NO:9).
- 5. A fragment of the protein of claim 1 having histone acetyltransferase activity.
- 6. A polypeptide consisting of the amino acid sequence of SEQ ID NO: 2.
- 7. A fragment of the protein of claim 1 which binds to the amino acid sequence of SEQ ID NO: 3 on the p300 cellular protein and the amino acid sequence of SEQ ID NO:9 on the CBP cellular protein.
- 8. A polypeptide consisting of the amino acid sequence of SEQ ID NO:4.
- 9. A nucleic acid consisting of the nucleotide sequence of SEQ ID NO:10.
- 10. A nucleic acid having a nucleotide sequence which encodes the protein of claim

- A nucleic acid having a nucleotide sequence which encodes the protein of claim
- 2.
- 12. A nucleic acid having a nucleotide sequence which encodes the protein of claim

3.

- A nucleic acid consisting of the nucleotide sequence which encodes the protein of claim 4.
- 14 A nucleic acid complementary to and which selectively hybridizes with the nucleic acid of claim 11 under stringent hybridization conditions.
- 15. A fragment of the nucleic acid of claim 9, which encodes a polypeptide that acetylates histones.
- A fragment of the nucleic acid of claim 9, which encodes a polypeptide which binds to the amino acid sequence of SEQ ID NO:3 on the p300 cellular protein and the amino acid sequence of SEQ ID NO:9 on the CBP cellular protein.
- 17. A purified antibody which specifically binds the protein of claim 1.
- 18. A purified antibody which specifically binds the protein of claim 2.
- 19. A purified antibody which specifically binds the protein of claim 3.
- A purified antibody which specifically binds the protein of claim 4.
- An assay for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of P/CAF comprising:
- a) contacting the substance with a system in which histone acetylation by P/CAF can be determined,





- b) determining the amount of histone acetylation by P/CAF in the presence of the substance, and
- c) comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased or increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of P/CAF.
- 22. An assay for screening substances for the ability to inhibit binding of P/CAF to p300/CBP comprising:
- a) contacting the substance with a system in which the P/CAF binding of P300/CBP can be determined,
- b) determining the amount of P/CAF binding of p300/CBP in the presence of the substance; and
- c) comparing the amount of binding of P/CAF to p300/CBP in the presence of the substance with the amount of binding of P/CAF to p300/CBP in the absence of the substance, a decreased amount of binding of P/CAF to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of P/CAF to p300/CBP.
- The method of claim 22, wherein the system consists of a cell free reaction mixture comprising a fragment of the p300 protein comprising amino acid residues 1767-1816 (SEQ ID NO:3) and the protein of claim 4.
- The method of claim 22, wherein the system consists of a cell free reaction mixture comprising a fragment of the CBP protein comprising amino acid residues 1805-1854 (SEQ ID NO:9) and the protein of claim 4.
- The method of claim 22, wherein the system consists of a cell extract produced from cells producing both p300 and P/CAF.



- An assay for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of p300/CBP comprising:
- a) contacting the substance with a system in which histone acetylation by p300/CBP can be determined;
- b) determining the amount of histone acetylation by p300/CBP in the presence of the substance; and
- c) comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, a decreased or increased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of p300/CBP.
- An assay for screening substances for the ability to inhibit binding of a DNAbinding transcription factor to p300/CBP comprising:
- a) contacting the substance with a system in which the DNA-binding transcription factor binding of P300/CBP can be determined,
- b) determining the amount of DNA-binding transcription factor binding of p300/CBP in the presence of the substance; and
- c) comparing the amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance with the amount of binding of DNA-binding transcription factor to p300/CBP in the absence of the substance, a decreased amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of DNA-binding transcription factor to p300/CBP.
- 28. The method of claim 27, wherein the system consists of a cell free reaction mixture comprising a DNA-binding transcription factor and p300/CBP.
- The method of claim 27, wherein the system consists of a cell free reaction mixture comprising a fragment of the CBP protein comprising a DNA-binding transcription factor and p300/CBP.



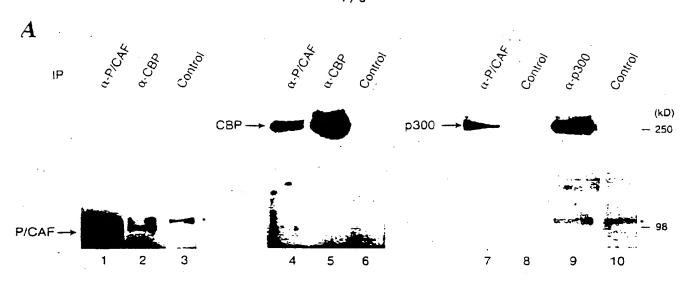


- The method of claim 27, wherein the system consists of a cell extract produced from cells producing both a DNA-binding transcription factor and p300/CBP.
- The method of claim 27, wherein the DNA-binding transcription factor is selected from the group consisting of a nuclear hormone receptor, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YY1, Sap-1a, c-Fos, MyoD and SRC-1
- 32. A method for inhibiting the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.
- 33. The method of claim 32, wherein the substance can inhibit the transcription modulating activity of P/CAF by preventing the binding of P/CAF to p300/CBP.
- 34. A method for stimulating the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity stimulating amount of a substance in a pharmaceutically acceptable carrier
- -35 The method of claim 34, wherein the substance can stimulate the transcription modulating activity of P/CAF by promoting the binding of P/CAF to p300/CBP
  - 36. The method of claim 34, wherein the substance can stimulate the transcription modulating activity of P/CAF by stimulating the histone acetlytransferase activity of P/CAF.
  - 37. A method for inhibiting the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity inhibiting amount of a substance in a pharmaceutically acceptable carrier





- 104
- The method of claim 37, wherein the substance can inhibit the transcription modulating activity of p300/CBP by preventing the binding of a DNA-binding transcription factor to p300/CBP
- The method of claim 38, wherein the DNA-binding transcription factor is selected from the group consisting of a nuclear hormone receptor, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YY1, Sap-1a, c-Fos, MyoD and SRC-1.
- The method of claim 37, wherein the substance is an antibody which binds p300/CBP.
- A method for stimulating the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity stimulating amount of a substance in a pharmaceutically acceptable carrier.
- The method of claim 41, wherein the substance can stimulate the histone acetyltransferase activity of p300/CBP by promoting the binding of a DNA-binding transcription factor to p300/CBP.



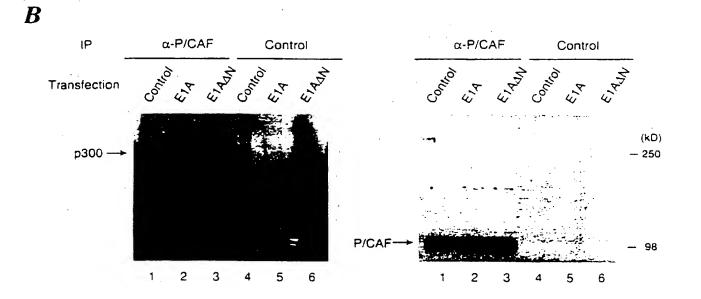


Fig. 1

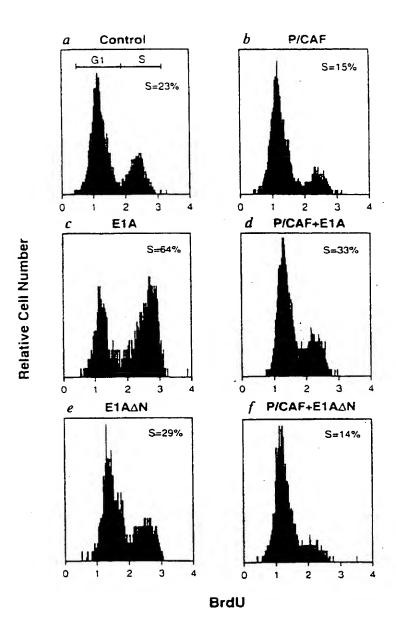


Fig. 2

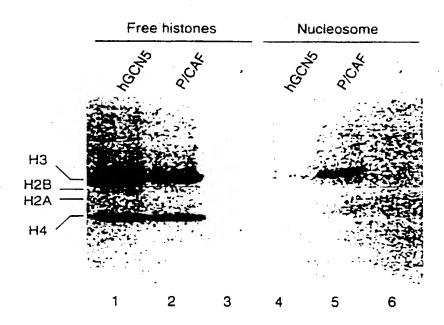


Fig. 3

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# INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(71) Applicant (for all designated States except US): THE GOV-ERNMENT OF THE UNITED STATES OF AMERICA, represented by THE SECRETARY, DEPARTMENT OF HEALTH AND HUMAN SERVICES [US/US]; National Institutes of Health, Office of Technology Transfer Suite 325, 6011 Executive Boulevard, Rockville, MD 20852 (US).

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(74) Agents: MILLER, Mary, L. et al., Needle & Rosenberg, Suite 1200, 127 Peachtree Street, N.E., Atlanta, GA 30303 (US).

(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD. SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN, ARIPO patent (GH, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD,

#### Published

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Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

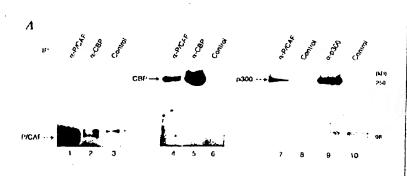
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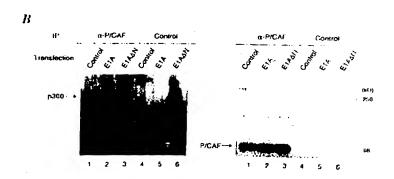
26 February 1998 (26.02.98)

(54) Title: P300/CBP-ASSOCIATED TRANSCRIPTIONAL CO-FACTOR P/CAF AND USES THEREOF

#### (57) Abstract

The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones and which also binds to the p300/CBP cellular protein. The present invention further provides a nucleic acid encoding the P/CAF protein as well as a vector containing the nucleic acid and a host for the vector. A purified antibody which specifically binds the P/CAF protein is also provided. Also provided are methods of screening for compounds that inhibit or stimulate the transcription modulating and histone acetyltransferase activity of P/CAF and p300/CBP.





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nal Application No PCT/US 97/12877

A. CLASS	C12N15/12 C07K14/47 G01N33	/50 A61K38/17	
According t	to International Patent Classification (IPC) or to both national classif	fication and IPC	
B. FIELDS	SEARCHED		
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Documento	ation searched other than minimum documentation to the extent tha	t such documents are included in the fields a	earched
Electronia d	data base consulted during the international search (name of data i	Dase and, where practical, search terms used	d)
C. DOCUM	ENTS CONSIDERED TO BE RELEVANT		
Category °	Citation of document, with indication, where appropriate, of the re	elevant passages	Relevant to claim No.
A	EMBL EST, Accession numberN3952 Sequence no yv27b08.sl Homo sap		1
	clone 243927 3' 25 January 1996 XP002050402 see the whole document		
<b>A</b>	GEORGAKOPOULOS, T. & THIREOS, G EMBO JOURNAL., vol. 11, 1992, EYNSHAM, OXFORD pages 4145-4152, XP002050399 see the whole document	.: GB,	1
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X Furth	ner documents are listed in the continuation of box C.	Patent family members are listed in	п аппех.
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Date of the a	ctual completion of the international search	Date of mailing of the international sear	ch report
. 17	December 1997	1 4. 01. 98	
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C.(Continu	ation) DOCUMENTS CONSIDERED TO BE RELEVANT		· · · · · · · · · · · · · · · · · · ·
Category °	Citation of document, with indication, where appropriate, of the relevant passages		Relevant to claim No.
Р,Х	YANG, X.Y. ET AL.: "A p300-CBP-associated factor that competes with the adenoviral oncoprotein E1A" NATURE., vol. 382, no. 8589, 25 July 1996, LONDON GB, pages 319-324, XP002050400 see the whole document		1
P,X	OGRYZKO, V.V. ET AL.: "The transcriptional coactivators p300 and CBP are histone acetyltransferases" CELL, vol. 87, no. 5, November 1996, NA US, pages 953-959, XP002050401 see the whole document		1
P,X	EMBL EST, Accession number U57316, Sequence reference human GCN5 (hGCN5) complete cds. 26 august 1996 XP002050403 see the whole document		
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## INTERNATIONAL SEARCH REPORT

mis-national application No. PCT/US 97/12877

Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This Int	ernational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. χ	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
	see FURTHER INFORMATION sheet PCT/ISA/210
2.	Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
, []	Claims Nos.
السا	because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Inte	ernational Searching Authority found multiple inventions in this international application, as follows:
•	
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4.	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark	on Protest  The additional search fees were accompanied by the applicant's protest.
	No protest accompanied the payment of additional search fees.

International Application No PCT/US 97 /12877

# FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Remark: Although claims 32 to 42 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the composition.







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## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(71) Applicant (for all designated States except US): THE GOVERNMENT OF THE UNITED STATES OF AMERICA, represented by THE SECRETARY, DEPARTMENT OF HEALTH AND HUMAN SERVICES [US/US]; National Institutes of Health, Office of Technology Transfer Suite 325, 6011 Executive Boulevard, Rockville, MD 20852 (US).

(72) Inventors; and

(75) Inventors/Applicants (for US only): NAKATANI, Yoshihiro [JP/US]; 4624 Edgefield Road, Bethesda, MD 20814 (US). HOWARD, Bruce, H. [US/US]; 8715 Fallen Oak Drive, Bethesda, MD 20817 (US).

(74) Agents: MILLER, Mary, L. et al.; Needle & Rosenberg, Suite 1200, 127 Peachtree Street, N.E., Atlanta, GA 30303 (US).

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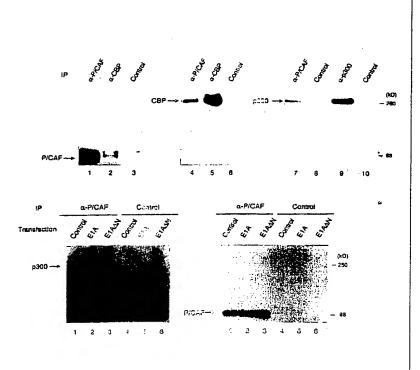
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The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones and which also binds to the p300/CBP cellular protein. The present invention further provides a nucleic acid encoding the P/CAF protein as well as a vector containing the nucleic acid and a host for the vector. A purified antibody which specifically binds the P/CAF protein is also provided. Also provided are methods of screening for compounds that inhibit or stimulate the transcription modulating and histone acetyltransferase activity of P/CAF and p300/CBP.



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#### P300/CBP-ASSOCIATED TRANSCRIPTIONAL CO-FACTOR P/CAF AND USES THEREOF

### **BACKGROUND OF THE INVENTION**

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#### Field of the Invention

The present invention provides a transcriptional co-factor, p300/CBP-associated factor (P/CAF), which modulates transcription through binding to the cellular transcription co-factors p300 and CBP and through acetylation of histones. Also provided are methods for screening for the presence of P/CAF and for substances which alter the transcription modulating effect and growth regulatory activity of P/CAF.

### Background Art

Cellular proteins p300 and CBP are global transcriptional coactivators that are involved in the regulation of various DNA-binding transcriptional factors (Janknecht and Hunter, 1996). Recently, p300 was found to be very closely related to CBP, a factor that binds selectively to the protein kinase A-phosphorylated form of CREB (3-5). Cellular factors p300 and CBP exhibit strong amino acid sequence similarity and share the capacity to bind both CREB and E1A (6-8). Although neither p300 nor CBP by itself binds to DNA, each can be recruited to promoter elements via interaction with sequence-specific activators and functions to be a transcriptional adaptor. For simplicity, p300 and CBP will be termed p300/CBP in the context of discussing their shared functional properties.

p300/CBP is a large protein consisting of over 2,400 amino acids, known to interact with a variety of DNA-binding transcriptional factors including nuclear hormone receptors (13,57), CREB (3,4, 7), c-Jun/v-Jun (9,11), YY1 (10), c-Myb/v-Myb (12,58), Sap-1a (59), c-Fos (11) and MyoD (60). DNA-binding factors recruit p300/CBP not only by direct but also indirect interactions through cofactors, for example, nuclear hormone receptors recruit p300/CBP directly as well as through indirect interactions, via SRC-1, which stimulates transcription by binding to various nuclear hormone receptors (13,61).



PCT/US97/12877

The transforming proteins encoded by adenovirus and several other small DNA tumor viruses disturb host cell growth control by interacting with cellular factors that normally function to repress cell proliferation. One of the most intensively studied of these viral proteins, the product of the adenovirus E1A gene, is itself sufficient for transformation (1). E1A transforming activity resides in two distinct domains, the targets of which include p300/CBP and products of the retinoblastoma (RB) susceptibility gene family (1,2). Interactions of E1A with p300/CBP and RB are thought to influence functionally distinct growth regulatory pathways, allowing the two domains to contribute additively to transformation (1).

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The paradigm for how E1A and functionally related viral proteins perturb cell growth regulation derives in large part from studies on their interactions with RB (1,2). The molecular function of E1A is based on its capacity to interfere with cellular protein-protein interactions. Since both E1A and various cellular targets bind to a site in RB termed the pocket domain (2), E1A can competitively disrupt the complex formation between RB and its cellular targets.

The second cellular factor implicated in E1A-dependent transformation, p300, is believed to inhibit G0/G1 exit, to activate certain enhancers, and to stimulate differentiation (1,2). E1A inhibits the p300/CBP-mediated transcriptional activation of many promoters (14). In one case that has been examined, the complex of p300 and YY1, E1A inhibits transcription without disrupting the complex (10).

The present invention provides a cellular protein designated P/CAF which binds to p300/CBP and plays an important role in both transcription and cell cycle regulation associated with a histone acetyltransferase activity. The present invention also provides a histone acetyltransferase activity in the p300/CBP cellular protein, thus providing targets for modulating transcription and cell cycle regulation in cells.

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The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones and which also binds to the p300/CBP cellular protein.

The present invention further provides a nucleic acid encoding the P/CAF protein as well as a vector containing the nucleic acid and a host for the vector. A purified antibody which specifically binds the P/CAF protein is also provided.

In addition, also provided is a bioassay for screening substances for the ability to inhibit the transcription modulating activity of P/CAF and/or histone acetyltransferase activity, comprising contacting the substance with a system in which histone acetylation by P/CAF can be determined, determining the amount of histone acetylation by P/CAF in the presence of the substance, and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit the transcription modulating activity and/or histone acetyltransferase activity of P/CAF.

Furthermore, the present invention provides a bioassay for screening substances for the ability to inhibit the transcription modulating activity and/or histone acetyltransferase activity of P/CAF comprising contacting the substance with a system in which the p300 binding of P/CAF can be determined; determining the amount of p300 binding of P/CAF in the presence of the substance, and comparing the amount of p300 binding of P/CAF in the presence of the substance with the amount of p300 binding of P/CAF in the absence of the substance, a decreased amount of p300 binding of P/CAF in the presence of the substance indicating a substance that can inhibit the transcription modulating activity and/or histone acetyltransferase activity of P/CAF.

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Also provided is a method for determining the amount of P/CAF in a biological sample comprising contacting the biological sample with a polypeptide comprising the amino acid sequence of SEQ ID NO:3 under conditions whereby a P/CAF/p300 complex can be formed, and determining the amount of the P/CAF/p300 complex, the amount of the complex indicating the amount of P/CAF in the sample.

The present invention additionally provides a method for determining the amount of P/CAF in a biological sample comprising contacting the biological sample with an antibody which specifically binds P/CAF under conditions whereby a P/CAF/antibody complex can be formed, and determining the amount of the P/CAF/antibody complex, the amount of the complex indicating the amount of P/CAF in the sample.

Also provided herein is an assay for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of P/CAF, comprising: contacting the substance with a system in which histone acetylation by P/CAF can be determined; determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased or increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of P/CAF.

The present invention further provides an assay for screening substances for the ability to inhibit binding of P/CAF to p300/CBP comprising: contacting the substance with a system in which the P/CAF binding of P300/CBP can be determined; determining the amount of P/CAF binding of p300/CBP in the presence of the substance; and comparing the amount of binding of P/CAF to p300/CBP in the presence of the substance with the amount of binding of P/CAF to p300/CBP in the absence of the substance, a decreased amount of binding of P/CAF to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of P/CAF to p300/CBP.

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In addition, an assay is provided for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of p300/CBP, comprising: contacting the substance with a system in which histone acetylation by p300/CBP can be determined; determining the amount of histone acetylation by p300/CBP in the presence of the substance; and comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, a decreased or increased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of p300/CBP.

Furthermore, the present invention provides an assay for screening substances for the ability to inhibit binding of a DNA-binding transcription factor to p300/CBP comprising: contacting the substance with a system in which the DNA-binding transcription factor binding of P300/CBP can be determined, determining the amount of DNA-binding transcription factor binding of p300/CBP in the presence of the substance; and comparing the amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance with the amount of binding of DNA-binding transcription factor to p300/CBP in the absence of the substance, a decreased amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of DNA-binding transcription factor to p300/CBP.

A method is also provided for inhibiting the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.

Also provided in the present invention is a method for stimulating the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity stimulating amount of a substance in a pharmaceutically acceptable carrier.

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Furthermore, the present invention provides a method for inhibiting the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.

Finally, the present invention additionally provides a method for stimulating the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity stimulating amount of a substance in a pharmaceutically acceptable carrier.

#### BRIEF DESCRIPTION OF THE FIGURES

Figs. 1A-B. Fig 1A: P/CAF-p300/CBP interaction *in vivo*. Cell extract was immunoprecipitated with rabbit anti-P/CAF (lanes 1, 4, and 7), rabbit anti-CBP (lanes 2 and 5), and mouse anti-p300 (lane 9) antibodies. For controls, cell extract was precipitated with rabbit control IgG (lanes 3, 6, and 8) or mouse anti-HA monoclonal antibody (lane 10). The precipitates were analyzed by immunoblotting with anti-P/CAF (lanes 1-3), anti-CBP (lanes 4-6), and anti-p300 (lanes 7-10) antibodies. The positions of non-specific bands are indicated by asterisks. Fig. 1B. E1A inhibits the P/CAF-p300 interaction *in vivo*. Osteosarcoma cells were transfected with either control vector (lanes 1 and 4) or E1A- (lanes 2 and 5) or E1AΔN- (lanes 3 and 6) expression vectors. Extract from the transfected subpopulation was immunoprecipitated with anti-P/CAF (lanes 1-3) or control (lanes 4-6) IgG. The precipitates were analyzed by immunoblotting with anti-p300 and anti-P/CAF.

Figs. 2A-F. P/CAF and E1A mediate antagonistic effects on cell cycle progression. HeLa cells (ATCC accession number CCL 2) were transfected by electroporation with 7  $\mu$ g of P/CAF-expression plasmid and/or 3  $\mu$ g of the full-length or the N-terminally deleted ( $\Delta 2$ -36) E1A 12S-expression plasmid as indicated in the figure. These plasmids were constructed by subcloning FLAG-P/CAF and E1A cDNAs into

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pCX (34) and pcDNAI (Invitrogen), respectively. All samples, in addition, contained 1  $\mu$ g of sorting plasmid (pCMV-IL2R) (31) and carrier plasmid (pCX) to normalize the total amount of DNA to 11  $\mu$ g. After transfection, cells were incubated in Dulbecco's modified Eagle's medium with 10% fetal bovine calf serum for 12 hours and subsequently labeled in medium containing 10  $\mu$ M bromo-deoxyuridine (BrdU) for 30 min. Subsequently, the transfected subpopulation was purified by magnetic affinity cell sorting and nuclei were analyzed by dual parameter flow cytometry as described (32). Histograms show percentages of cells in G1 and S phases. Abscissa values represent fluorescence intensity of bound anti-BrdU antibodies in log scale.

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Fig. 3. Histone acetyltransferase activity of P/CAF. Activity of hGCN5 (lanes 1 and 4) and P/CAF (lanes 2 and 5) that acetylates free histones (lanes 1-3) or histones in the nucleosome core particle (35) (lanes 4-6) was measured as described (36). Each reaction contains 0.3 pmol of affinity purified FLAG-hGCN5 or FLAG-P/CAF, 4 pmol of the histone octamer or the nucleosome core particle and 10 pmol of [1-14C]acetyl-CoA. Note that the histone octamer dissociates into dimers or tetramers under assay conditions. Acetylated histones were detected by autoradiography after separation by SDS-PAGE. The bands corresponding to acetylated histones H3 and H4 are indicated by arrows.

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## DETAILED DESCRIPTION OF THE INVENTION

As used in the specification and in the claims, "a" can mean one or more, depending upon the context in which it is used.

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## P/CAF protein and fragments

The present invention provides a purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones. The P/CAF protein can also bind to the amino acid region of SEQ ID NO 3

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(amino acid (aa) residues 1753 - 1966) of the cellular transcriptional factor, p300 (which has the complete amino acid sequence of SEQ ID NO:6 and the nucleotide sequence of SEQ ID NO:12), and the amino acid region of SEQ ID NO:6 (amino acid residues 1805 - 1854) of the cellular transcriptional factor, CBP (which has the complete amino acid sequence of SEQ ID NO:7 and the nucleotide sequence of SEQ ID NO:13). The P/CAF protein can be defined by any one or more of the typically used parameters. Examples of these parameters include, but are not limited to molecular weight (calculated or empirically determined), isoelectric focusing point, specific epitope(s), complete amino acid sequence, sequence of a specific region (e.g., N-terminus) of the amino acid sequence and the like

For example, The P/CAF protein can consist of the amino acid sequence of SEQ ID NO:1 or the P/CAF protein can comprise the amino acid sequence of SEQ ID NO:2 which represents the carboxy terminal end of the P/CAF protein and contains the histone acetyltransferase activity, or the amino acid sequence of SEQ ID NO:4, which represents the amino terminal end of the P/CAF protein, containing the binding site for p300/CBP. Because the amino-terminal region is specific for P/CAF it can be used to define and identify P/CAF.

As used herein, "purified" refers to a protein (polypeptide, peptide, etc.) that is sufficiently free of contaminants or cell components with which it normally occurs to distinguish it from the contaminants or other components of its natural environment. The purified protein need not be homogeneous, but must be sufficiently free of contaminants to be useful in a clinical or research setting, for example, in an assay for detecting antibodies to the protein. Greater levels of purity can be obtained using methods derived from well known protocols. Specific methods for purifying P/CAF proteins are known in the art.

As will be appreciated by those skilled in the art, the invention also includes those P/CAF polypeptides having slight variations in amino acid sequence which yield polypeptides equivalent to the P/CAF protein defined herein. Such variations may arise

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naturally as allelic variations (e.g., due to genetic polymorphism) or may be produced by human intervention (e.g., by mutagenesis of cloned DNA sequences), such as induced point, deletion, insertion and substitution mutants. Minor changes in amino acid sequence are generally preferred, such as conservative amino acid replacements, small internal deletions or insertions, and additions or deletions at the ends of the molecules. Substitutions may be designed based on, for example, the model of Dayhoff, et al. (37). These modifications can result in changes in the amino acid sequence, provide silent mutations, modify a restriction site, or provide other specific mutations.

Modifications to any of the P/CAF proteins or fragments can be made, while preserving the specificity and activity (function) of the native protein or fragment thereof. As used herein, "native" describes a protein that occurs in nature. The modifications contemplated herein can be conservative amino acid substitutions, for example, the substitution of a basic amino acid for a different basic amino acid. Modifications can also include creation of fusion proteins with epitope tags or known recombinant proteins or genes encoding them created by subcloning into commercial or non-commercial vectors (e.g., polyhistidine tags, flag tags, myc tag, glutathione-S-transferase [GST] fusion protein, *xylE* fusion reporter construct). Furthermore, the modifications can be such as do not affect the function of the protein or the way the protein accomplishes that function (e.g., its secondary structure or the ultimate result of the protein's activity). These products are equivalent to the P/CAF protein. The means for determining the function, way and result parameters are well known.

Having provided an example of a purified P/CAF protein, the invention also enables the purification of P/CAF homologs from other species and allelic variants from individuals within a species. For example, an antibody raised against the exemplary human P/CAF protein can be used routinely to screen preparations from different humans for allelic variants of the P/CAF protein that react with the P/CAF protein-specific antibody. Similarly, an antibody raised against an epitope, for example, from a conserved amino acid region of the human P/CAF protein can be used to routinely screen for homologs of the P/CAF protein in other species. A P/CAF protein can be

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routinely identified in and obtained from other species and from individuals within a species using the methods taught herein and others known in the art. For example, given the present sequence, the DNA encoding a conserved amino acid sequence can be used to probe genomic DNA or DNA libraries of an organism to predictably obtain the P/CAF gene for that organism. The gene can then be cloned and expressed as the P/CAF protein and purified according to any of a number of routine, predictable methods. An example of the routine protein purification methods available in the art can be found in Pei et al. (38).

A purified polypeptide fragment of the P/CAF protein is also provided. The term "fragment" as used herein regarding a P/CAF protein, means a molecule of at least five contiguous amino acids of P/CAF protein that has at least one function shared by P/CAF protein or a region thereof. These functions can include antigenicity, binding capacity, acetyltransferase activity and structural roles, among others. The P/CAF fragment can be specific for a recited source. As used herein to describe an amino acid sequence (protein, polypeptide, peptide, etc.), "specific" means that the amino acid sequence is not found identically in any other source. The determination of specificity is made routine by the availability of computerized amino acid sequence databases and sequence comparison programs, wherein an amino acid sequence of almost any length can be quickly and reliably checked for the existence of identical sequences. If an identical sequence is not found, the protein is "specific" for the recited source. For example, a P/CAF fragment can be species-specific (e.g., found in the P/CAF protein of humans, but not of other species).

A fragment of the P/CAF protein having histone acetyltransferase activity can consist of the amino acid sequence of SEQ ID NO:2. A fragment of the P/CAF protein which binds to the amino acid sequence of SEQ ID NO:3 on p300 and the amino acid sequence of SEQ ID NO:9 on CBP can consist of the amino acid sequence of SEQ ID NO:4. To the extent that these fragments are specific for P/CAF, they can be used to identify and define P/CAF.

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An antigenic fragment of P/CAF protein is provided. An antigenic fragment has an amino acid sequence of at least about five consecutive amino acids of a P/CAF protein amino acid sequence and binds an antibody or elicits an immune response in an animal. An antigenic fragment can be selected by applying the routine technique of epitope mapping to P/CAF protein to determine the regions of the proteins that contain epitopes reactive with antibodies or are capable of eliciting an immune response in an animal. Once the epitope is selected, an antigenic polypeptide containing the epitope can be synthesized directly, or produced recombinantly by cloning nucleic acids encoding the antigenic polypeptide in an expression system, according to standard

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Alternatively, an antigenic fragment of the antigen can be isolated from the whole P/CAF protein or a larger fragment of the P/CAF protein by chemical or mechanical disruption. Fragments can also be randomly chosen from a known P/CAF protein sequence and synthesized. The purified fragments thus obtained can be tested to determine their antigenicity and specificity by routine methods.

## Nucleic Acids Encoding P/CAF Protein

An isolated nucleic acid that encodes a P/CAF protein is also provided. As used herein, the term "isolated" means a nucleic acid separated or substantially free from at least some of the other components of the naturally occurring organism, for example, the cell structural components commonly found associated with nucleic acids in a cellular environment and/or other nucleic acids. The isolation of nucleic acids can therefore be accomplished by techniques such as cell lysis followed by phenol plus chloroform extraction, followed by ethanol precipitation of the nucleic acids (39). It is not contemplated that the isolated nucleic acids are necessarily totally free of all nonnucleic acid components or all other nucleic acids, but that the isolated nucleic acids are isolated to a degree of purification to be useful in clinical, diagnostic, experimental, or other procedures such as, for example, gel electrophoresis, Southern, Northern or dot blot hybridization, or polymerase chain reaction (PCR).

A skilled artisan in the field will readily appreciate that there are a multitude of procedures which may be used to isolate the nucleic acids prior to their use in other procedures. These include, but are not limited to, lysis of the cell followed by gel filtration or anion exchange chromatography, binding DNA to silica in the form of glass beads, filters or diatoms in the presence of high concentrations of chaotropic salts, or ethanol precipitation of the nucleic acids.

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The nucleic acids of the present invention can include positive and negative strand RNA as well as DNA and can include genomic and subgenomic nucleic acids found in the naturally occurring organism. The nucleic acids contemplated by the present invention include double stranded and single stranded DNA of the genome, complementary positive stranded cRNA and mRNA, and complementary cDNA produced therefrom and any nucleic acid which can selectively or specifically hybridize to the isolated nucleic acids provided herein. Stringent conditions (further described below) are used to distinguish selectively or specifically hybridizing nucleic acids from non-selectively and non-specifically hybridizing nucleic acids.

An isolated nucleic acid that encodes a P/CAF protein can be species-specific (i.e., does not encode the P/CAF protein of other species and does not occur in other species). Examples of the nucleic acids contemplated herein include the nucleic acid of SEQ ID NO:10 as well as the nucleic acids that encode each of the P/CAF proteins or fragments thereof described herein. P/CAF proteins and protein fragments can be routinely obtained as described herein and their structure (sequence) determined by routine means including the methods as used herein.

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P/CAF protein-encoding nucleic acids can be isolated from an organism in which they are normally found (e.g., humans), using any of the routine techniques. For example, a genomic DNA or cDNA library can be constructed and screened for the presence of the nucleic acid of interest using one of the present P/CAF protein-encoding nucleic acids as a probe. Methods of constructing and screening such libraries are well known in the art and kits for performing the construction and screening steps are

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commercially available (for example, Stratagene Cloning Systems, La Jolla, CA). Once isolated, the nucleic acid can be directly cloned into an appropriate vector, or if necessary, be modified to facilitate the subsequent cloning steps. Such modification steps are routine, an example of which is the addition of oligonucleotide linkers, which contain restriction sites, to the termini of the nucleic acid (See, for example, ref. 39).

P/CAF protein-encoding nucleic acids can also be synthesized. For example, a method of obtaining a DNA molecule encoding a specific P/CAF protein is to synthesize a recombinant DNA molecule which encodes the P/CAF protein. For example, nucleic acid synthesis procedures are routine in the art and oligonucleotides coding for a particular protein region are readily obtainable through automated DNA synthesis. A nucleic acid for one strand of a double-stranded molecule can be synthesized and hybridized to its complementary strand. One can design these oligonucleotides such that the resulting double-stranded molecule has either internal restriction sites or appropriate 5' or 3' overhangs at the termini for cloning into an appropriate vector.

Oligonucleotides complementary to or identical with the P/CAF proteinencoding nucleic acid sequence can be synthesized as primers for amplification reactions, such as PCR, or as probes to detect P/CAF protein encoding nucleic acids by various hybridization protocols (e.g., Northern blot, Southern blot, dot blot, colony screening, etc.)

Double-stranded molecules coding for relatively large proteins can readily be synthesized by first constructing several different double-stranded molecules that code for particular regions of the protein, followed by ligating these DNA molecules together. For example, Cunningham, et al. (40), have constructed a synthetic gene encoding the human growth hormone by first constructing overlapping and complementary synthetic oligonucleotides and ligating these fragments together. See also, Ferretti, et al. (41), wherein synthesis of a 1057 base pair synthetic bovine rhodopsin gene from synthetic oligonucleotides is disclosed.

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By constructing a P/CAF protein-encoding nucleic acid in this manner, one skilled in the art can readily obtain any particular P/CAF protein with modifications at any particular position or positions. See also, U.S. Patent No. 5,503,995 which describes an enzyme template reaction method of making synthetic genes. Techniques such as this are routine in the art and are well documented. DNA encoding the P/CAF protein or P/CAF protein fragments can then be expressed *in vivo* or *in vitro*.

The nucleic acid encoding the P/CAF protein can be any nucleic acid that functionally encodes the P/CAF protein. To functionally encode the protein (i.e., allow the nucleic acid to be expressed), the nucleic acid can include, but is not limited to, expression control sequences, such as an origin of replication, a promoter, regions upstream or downstream of the promoter, such as enhancers that may regulate the transcriptional activity of the promoter, appropriate restriction sites to facilitate cloning of inserts adjacent to the promoter, antibiotic resistance genes or other markers which can serve to select for cells containing the vector or the vector containing the insert, and necessary information processing sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites and transcription termination sequences as well as any other sequence which may facilitate the expression of the inserted nucleic acid.

Preferred expression control sequences are promoters derived from metallothionine genes, actin genes, immunoglobulin genes, CMV, SV40, adenovirus, bovine papilloma virus, etc. A nucleic acid encoding a P/CAF protein can readily be determined based upon the genetic code for the amino acid sequence of the P/CAF protein and many nucleic acid sequences will encode a P/CAF protein. Modifications in the nucleic acid sequence encoding the P/CAF protein are also contemplated. Modifications that can be useful are modifications to the sequences controlling expression of the P/CAF protein to make production of P/CAF protein inducible or repressible as controlled by the appropriate inducer or repressor. Such means are standard in the art (see, e.g., ref. 39). The nucleic acids can be generated by means standard in the art, such as by recombinant nucleic acid techniques, as exemplified in the examples herein, and by synthetic nucleic acid synthesis or in vitro enzymatic synthesis.

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After a nucleic acid encoding a particular P/CAF protein of interest, or a region of that nucleic acid, is constructed, modified, or isolated, that nucleic acid can then be cloned into an appropriate vector, which can direct the *in vivo* or *in vitro* synthesis of that wild-type and/or modified P/CAF protein. The vector is contemplated to have the necessary functional elements that direct and regulate transcription of the inserted nucleic acid, as described above. The vector containing the P/CAF nucleic acid or nucleic acid fragment can be in a host (e.g., cell or transgenic animal) for expressing the nucleic acid. The P/CAF protein or fragment thereof can thus be produced in a host system containing the expression vector and its functional activity as described herein can be demonstrated according to methods well known in the art.

There are numerous *E. coli* (*Escherichia coli*) expression vectors known to one of ordinary skill in the art useful for the expression of proteins. Other microbial hosts suitable for use include bacilli, such as *Bacillus subtilis*, and other enterobacteria, such as *Salmonella*, *Serratia*, as well as various *Pseudomonas* species. These prokaryotic hosts can support expression vectors which will typically contain expression control sequences compatible with the host cell (e.g., an origin of replication). In addition, any number of a variety of well-known promoters will be present, such as the lactose promoter system, a tryptophan (Trp) promoter system, a beta-lactamase promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence and have ribosome binding site sequences, for example, for initiating and completing transcription and translation. If necessary, an amino terminal methionine can be provided by insertion of a Met codon 5' and in-frame with the gene sequence. Also, the carboxy-terminal extension of the protein can be removed using standard oligonucleotide mutagenesis procedures.

Additionally, yeast expression can be used. There are several advantages to yeast expression systems. First, evidence exists that proteins produced in yeast secretion systems exhibit correct disulfide pairing. Second, post-translational glycosylation is efficiently carried out by yeast secretory systems. The *Saccharomyces cerevisiae* prepro-alpha-factor leader region (encoded by the  $MF\alpha$ -1 gene) is routinely used to direct

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protein secretion from yeast (42). The leader region of pre-pro-alpha-factor contains a signal peptide and a pro-segment which includes a recognition sequence for a yeast protease encoded by the *KEX2* gene. This enzyme cleaves the precursor protein on the carboxyl side of a Lys-Arg dipeptide cleavage-signal sequence. The polypeptide coding sequence can be fused in-frame to the pre-pro-alpha-factor leader region. This construct is then put under the control of a strong transcription promoter, such as the alcohol dehydrogenase I promoter or a glycolytic promoter. The protein coding sequence is followed by a translation termination codon which is followed by transcription termination signals. Alternatively, the polypeptide encoding sequence of interest can be fused to a second protein coding sequence, such as Sj26 or β-galactosidase, used to facilitate purification of the resultant fusion protein by affinity chromatography. The insertion of protease cleavage sites to separate the components of the fusion protein is applicable to constructs used for expression in yeast.

Efficient post-translational glycosylation and expression of recombinant proteins can also be achieved in *Baculovirus* expression systems in insect cells.

Mammalian cells permit the expression of proteins in an environment that favors important post-translational modifications such as folding and cysteine pairing, addition of complex carbohydrate structures and secretion of active protein. Vectors useful for the expression of proteins in mammalian cells are characterized by insertion of the protein encoding sequence between a strong viral promoter and a polyadenylation signal. The vectors can contain genes conferring either gentamicin or methotrexate resistance for use as selectable markers. For example, the antigen and immunoreactive fragment coding sequence can be introduced into a Chinese hamster ovary (CHO) cell line using a methotrexate resistance-encoding vector. Presence of the vector RNA in transformed cells can be confirmed by Northern blot analysis and production of a cDNA or opposite strand RNA corresponding to the protein encoding sequence can be confirmed by Southern and Northern blot analysis, respectively. A number of other suitable host cell lines capable of secreting intact proteins have been developed in the art and include the CHO cell lines, HeLa cells, myeloma cell lines, Jurkat cells, and the like

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Expression vectors for these cells can include expression control sequences, as described above. The vectors containing the nucleic acid sequences of interest can be transferred into the host cell by well-known methods, which vary depending on the type of cell host. For example, calcium chloride transfection is commonly utilized for prokaryotic cells, whereas calcium phosphate treatment or electroporation may be used for other cell hosts.

Alternative vectors for the expression of protein in mammalian cells, similar to those developed for the expression of human gamma-interferon, tissue plasminogen activator, clotting Factor VIII, hepatitis B virus surface antigen, protease Nexin I, and eosinophil major basic protein, can be employed. Further, the vector can include CMV promoter sequences and a polyadenylation signal available for expression of inserted nucleic acid in mammalian cells (such as COS7).

The nucleic acid sequences can be expressed in hosts after the sequences have been positioned to ensure the functioning of an expression control sequence. These expression vectors are typically replicable in the host organisms either as episomes or as an integral part of the host chromosomal DNA. Commonly, expression vectors can contain selection markers, e.g., tetracycline resistance or hygromycin resistance, to permit detection and/or selection of those cells transformed with the desired nucleic acid sequences (see, e.g., U.S. Patent 4,704,362).

The nucleic acids produced as described above can also be expressed in a host which is a non-human animal to create a transgenic animal, containing, in a germ or somatic cell, a nucleic acid comprising the coding sequence for all or a portion of the P/CAF protein, as well as all of the other regulatory elements required for expression of the P/CAF protein-encoding sequence. The animal will express the P/CAF gene or portion thereof to produce the P/CAF protein or protein fragment and such expression can be detected by determination of a particular phenotype unique to the transgenic animal expressing the transferred nucleic acid.

The nucleic acid can be the nucleic acid of SEQ ID NO:10, a nucleic acid having a nucleotide sequence which encodes the P/CAF protein, a nucleic acid having a nucleotide sequence which encodes the protein of SEQ ID NO:1, as well as the nucleic acids that encode the proteins comprising the fragments of SEQ ID NOS:2 and 4.

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The nucleic acids of the invention can contain substitutions or deletions which provide a particular phenotype of interest. For example, various deletions or base substitutions can be introduced into the nucleic acid encoding the P/CAF protein for the purpose of studying the effects of these particular deletions or substitutions on the transcription modulation activity of the P/CAF protein. These effects can be monitored by observation of such characteristics as growth and development of the animal, the ability to develop tumors, survival rates and the like. The gene construct introduced into the animal cells to produce the transgenic animal can contain any of the regulatory elements described above to modulate expression of the foreign genes. As used herein, the term "phenotype" includes morphology, biochemical profiles, changes in tumor formation and other parameters that are affected by the presence of the P/CAF protein.

The transgenic animals of the invention can also be used in a method for determining the effectiveness of administering a nucleic acid encoding a functional P/CAF protein to a subject in need of a functional P/CAF protein. First, a nucleic acid encoding a nonfunctional P/CAF protein can be introduced into the animal's cells and expressed to yield a characteristic phenotype. Then, using standard gene therapy techniques, a nucleic acid encoding a functional P/CAF protein can be introduced into the animal's cells and the effects on the animal's phenotypic characteristics can be determined.

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Having provided and taught how to obtain a nucleic acid that encodes a P/CAF protein, an isolated nucleic acid that encodes a fragment of P/CAF protein is also provided. The nucleic acid encoding the fragment can be obtained using any of the methods applicable to the nucleic acid encoding the entire P/CAF protein. The nucleic acid fragment can encode a species-specific P/CAF protein fragment (e.g., found in the

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P/CAF protein of humans, but not in the P/CAF proteins of other species). Nucleic acids encoding species-specific fragments of P/CAF protein are themselves species-specific or allele-specific fragments of the P/CAF gene.

Examples of fragments of a nucleic acid encoding a fragment of the P/CAF protein can include the nucleic acid sequences which encode the amino acid sequences of the fragments of SEQ ID NOS:2 or 4. The same routine computer analyses used to select these examples of fragments can be routinely used to obtain others. Fragments of P/CAF-encoding nucleic acids can be primers for PCR or probes, which can be species-specific, gene-specific or allele-specific. P/CAF-encoding nucleic acid fragments can encode antigenic or immunogenic fragments of P/CAF protein that can be used in therapeutic assays or screening protocols. P/CAF gene fragments can encode fragments of P/CAF protein having histone acetylase activity and/or p300/CBP binding activity as described above, as well as other uses that may become apparent.

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An isolated nucleic acid of at least ten nucleotides that selectively hybridizes with the nucleic acid of SEQ ID NO:10 under selected conditions is provided. For example, the conditions can be PCR amplification conditions and the hybridizing nucleic acid can be a primer consisting of a specific fragment of the reference sequence or a nearly identical nucleic acid that hybridizes only to the exemplified P/CAF-encoding nucleic acid or allelic variants thereof.

The invention provides an isolated nucleic acid that selectively hybridizes with the P/CAF-encoding nucleic acid sequence of SEQ ID NO:10 under stringent conditions. The hybridizing nucleic acid can be a probe that hybridizes only to the exemplified P/CAF-encoding nucleic acid sequence. Thus, the hybridizing nucleic acid can be a naturally occurring species-specific allelic variant of the exemplified P/CAF gene. The hybridizing nucleic acid can also include insubstantial base substitutions that do not prevent hybridization under the stated stringent conditions or affect either the function of the encoded protein, the way the protein accomplishes that function (e.g., its

secondary structure) or the ultimate result of the protein's activity. The means for determining these parameters are well known.

As used herein to describe nucleic acids, the term "selectively hybridizes" excludes the occasional randomly hybridizing nucleic acids as well as nucleic acids that encode other known homologs of the P/CAF protein. The selectively hybridizing nucleic acids of the invention can have at least 70%, 73%, 78%; 80%, 85%, 88%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or 100% complementarity with the segment and strand of the sequence to which it hybridizes. This list is not intended to exclude percent complementarity values between these values. The nucleic acids can be at least 10, 15, 16, 17, 18, 20, 21, 23, 24, 25, 30, 35; 40, 50, 100, 150, 200, 300, 500, 550, 750, 900, 950, or 1000 nucleotides in length or any intervening length, depending on whether the nucleic acid is to be used as a primer, probe or for protein expression. The hybridizing nucleic acid can comprise a region of at least ten nucleotides (up to full length) that is completely complementary to a unique region of the nucleic acid to which it hybridizes.

The nucleic acid can be an alternative coding sequence for the P/CAF protein, or can be used as a probe or primer for detecting the presence of or obtaining the P/CAF protein. If used as primers, the invention provides compositions including at least two nucleic acids which selectively hybridize with different regions of the nucleic acid so as to amplify a desired region. Depending on the length of the probe or primer, it can range between 70% complementary bases and full complementarity and still hybridize under stringent conditions.

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For example, for the purpose of obtaining or determining the presence of a nucleic acid encoding the P/CAF protein, the degree of complementarity between the hybridizing nucleic acid (probe or primer) and the sequence to which it hybridizes (P/CAF DNA in a sample) should be at least enough to exclude hybridization with a nucleic acid from another species. The invention provides examples of these nucleic acids of P/CAF, so that the degree of complementarity required to distinguish selectively



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hybridizing from nonselectively hybridizing nucleic acids under stringent conditions can be clearly determined for each nucleic acid. It should also be clear that the hybridizing nucleic acids of the invention will not hybridize with nucleic acids encoding unrelated proteins (hybridization is selective) under stringent conditions.

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"Stringent conditions" refers to the washing conditions used in a hybridization protocol. In general, the washing conditions should be a combination of temperature and salt concentration chosen so that the denaturation temperature is approximately 5-20°C below the calculated  $T_m$  of the nucleic acid hybrid under study. The temperature and salt conditions are readily determined empirically in preliminary experiments in which samples of reference DNA immobilized on filters are hybridized to the probe or protein encoding nucleic acid of interest and then washed under conditions of different stringencies. For example, the nucleic acid sequence of SEQ ID NO:10 was used as a specific radiolabeled probe for the detection of messenger RNA transcribed from the P/CAF gene by performing hybridizations under stringent conditions. The  $T_m$  of such an oligonucleotide can be estimated by allowing 2°C for each A or T nucleotide, and 4°C for each G or C. For example, an 18 nucleotide probe of 50% G+C would, therefore, have an approximate  $T_m$  of 54°C.

The invention provides an isolated nucleic acid that selectively hybridizes with the P/CAF gene shown in the sequence set forth as SEQ ID NO:10 under stringent conditions. The invention further provides an isolated nucleic acid complementary to the nucleotide sequence set forth in SEQ ID NO:10.

### 25 Antibodies to the P/CAF protein

A purified antibody and an antiserum containing polyclonal antibodies that specifically bind the P/CAF protein or antigenic fragment are also provided. The term "bind" means the well understood antigen/antibody binding as well as other nonrandom association with an antigen. "Specifically bind" as used herein describes an antibody or other ligand that does not cross react substantially with any antigen other than the one specified, in this case, an antigen of the P/CAF protein. Antibodies can be made as

described in Harlow and Lane (33). Briefly, purified P/CAF protein or an antigenic fragment thereof can be injected into an animal in an amount and in intervals sufficient to elicit a humoral immune response. Serum polyclonal antibodies can be purified directly, or spleen cells from the animal can be fused with an immortal cell line and screened for monoclonal antibody secretion, according to procedures well known in the art. Purified monospecific polyclonal antibodies that specifically bind the P/CAF antigen are also within the scope of the present invention. The antibodies of the present invention can bind the protein of claim 1, the protein of claim 2, the protein of claim 3 and/or the protein of claim 4, as well as any other proteins of the present invention.

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A ligand that specifically binds the antigen is also contemplated. The ligand can be a fragment of an antibody, such as, for example, an Fab fragment which retains P/CAF binding activity, or a smaller molecule designed to bind an epitope of the P/CAF antigen. The antibody or ligand can be bound to a substrate or labeled with a detectable moiety or both bound and labeled. The detectable moieties contemplated within the compositions of the present invention include those listed above in the description of the diagnostic methods, including fluorescent, enzymatic and radioactive markers.

The antibody can be bound to a solid support substrate or conjugated with a

detectable moiety or therapeutic compound or both bound and conjugated. Such
conjugation techniques are well known in the art. For example, conjugation of
fluorescent, radioactive or enzymatic moieties can be performed as described in the art
(33,43). The detectable moieties contemplated in the present invention can include
fluorescent, radioactive and enzymatic markers and the like. Therapeutic drugs
contemplated with the present invention can include cytotoxic moieties such as ricin A
chain, diphtheria toxin, pseudomonas exotoxin and other chemotherapeutic compounds.

It is well understood by one of skill in the art that all of the above discussion regarding antibodies to P/CAF can also be applied with regard to production, characterization and use of antibodies which bind the p300/CBP protein or any of the DNA-binding transcription factors of this invention.

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## Measuring the P/CAF protein in a sample

The present invention also provides a method for determining the presence and thus the amount of P/CAF protein in a biological sample. As used herein, a biological sample includes any tissue or cell which would contain the P/CAF protein. Examples of cells include tissues taken from surgical biopsies, isolated from a body fluid or prepared in an *in vitro* tissue culture environment.

One example of determining the amount of P/CAF in a biological sample can comprise contacting the biological sample with a polypeptide comprising the amino acid sequence of SEQ ID NO:3 under conditions whereby a P/CAF/p300 complex can be formed; and determining the amount of the P/CAF/p300 complex, the amount of the complex indicating the amount of P/CAF in the sample. Determination of the amount of P/CAF/p300 complex can be accomplished through techniques standard in the art. For example, the complex may be precipitated out of a solution and detected by the addition of a detectable moiety conjugated to the p300 protein or by the detection of an antibody which binds p300 or the P/CAF protein, as taught in the Examples herein. Antibodies which bind p300 or the P/CAF protein can be either monoclonal or polyclonal antibodies and can be obtained as described herein. Detection of P/CAF/p300 complexes by the detection of the binding of antibodies reactive with p300 or the P/CAF protein can be accomplished using various immunoassays as are available in the art, as described below.

Alternatively, determination of the amount of P/CAF in a biological sample can comprise contacting the biological sample with a polypeptide comprising the amino acid sequence of SEQ ID NO:9 under conditions whereby a P/CAF/CBP complex can be formed, and determining the amount of the P/CAF/CBP complex, the amount of the complex indicating the amount of P/CAF in the sample. Determination of the amount of P/CAF/CBP complex can be accomplished through techniques standard in the art. For example, the complex may be precipitated out of a solution and detected by the addition of a detectable moiety conjugated to the CBP protein or by the detection of an antibody which binds either CBP or the P/CAF protein, as taught in the Examples

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described below.

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herein. Antibodies which bind CBP or the P/CAF protein can be either monoclonal or polyclonal antibodies and can be obtained as described herein. Detection of P/CAF/CBP complexes by the detection of the binding of antibodies reactive with CBP or the P/CAF protein can be accomplished using various immunoassays as are available in the art, as

Another example of determining the amount of P/CAF in a biological sample comprises contacting the biological sample with an antibody which specifically binds P/CAF under conditions whereby a P/CAF/ antibody complex can be formed and determining the amount of the P/CAF/antibody complex, the amount of the complex indicating the amount of P/CAF in the sample. Antibodies which bind P/CAF can be either monoclonal or polyclonal antibodies and can be obtained as described herein. Determination of P/CAF/antibody complexes can be accomplished using various immunoassays as are available in the art, as described below.

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Immunoassays such as immunofluorescence assays, radioimmunoassays (RIA), immunoblotting and enzyme linked immunosorbent assays (ELISA) can be readily adapted for detection and measurement of P/CAF in a biological sample. Both polyclonal and monoclonal antibodies can be used in the assays. Available immunoassays are well known in the art and are extensively described in the patent scientific literature. See, for example, U.S. Patent Nos. 3,791,932; 3,839,153; 3,850,752; 3,850,578; 3,853,987; 3,867,517; 3,879,262; 3,901,654; 3,935,074; 3,984,533; 3,996,345; 4,034,074; and 4,098,876.

# 25 Screening assays for P/CAF

The present invention also provides a bioassay for screening substances for the ability to inhibit the histone acetyltransferase activity of P/CAF comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance under conditions whereby histone acetylation by P/CAF can occur, determining the amount of histone acetylation by P/CAF in the presence of the substance, and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the

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amount of histone acetylation by P/CAF in the absence of the substance, a decreased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit the histone acetyltransferase activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to inhibit the histone acetyltransferase activity of P/CAF can be added to this system and assayed for inhibiting ability.

The present invention also provides a bioassay for screening substances for the ability to inhibit the transcription modulating activity of P/CAF, comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance under conditions whereby histone acetylation by P/CAF can occur, determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit the transcription modulating activity and cell cycle progression suppressing activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to inhibit the transcription modulating activity of P/CAF by interfering with the histone acetyltransferase activity of P/CAF can be added to this system and assayed for inhibiting ability.



Also provided in the present invention is a bioassay for screening substances for the ability to inhibit the binding of p300 to P/CAF, comprising contacting a system in which the binding of p300 to P/CAF can be determined, with the substance under conditions whereby the binding of p300 and P/CAF can occur, determining the amount of p300 binding to P/CAF in the presence of the substance, and comparing the amount of p300 binding to P/CAF in the presence of the substance with the amount of p300 binding to P/CAF in the absence of the substance, a decreased amount of p300 binding to P/CAF in the presence of the substance, a decreased amount of p300 binding to P/CAF in the presence of the substance indicating a substance that can inhibit the binding of p300 to P/CAF. The binding of p300 to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the p300 protein comprising the amino acid sequence of SEQ ID NO:3 and P/CAF. Alternatively, the system can comprise a cell extract produced from cells producing both p300 and P/CAF. Determination of the binding of p300 to P/CAF can be carried out as taught herein.

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Additionally provided in the present invention is a bioassay for screening substances for the ability to inhibit the binding of CBP to P/CAF, comprising contacting a system in which the binding of CBP to P/CAF can be determined, with the substance under conditions whereby the binding of CBP to P/CAF can occur; determining the amount of CBP binding to P/CAF in the presence of the substance; and comparing the amount of CBP binding to P/CAF in the presence of the substance with the amount of CBP binding to P/CAF in the absence of the substance, a decreased amount of CBP binding to P/CAF in the presence of the substance indicating a substance that can inhibit the binding of CBP to P/CAF. The binding of CBP to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the CBP protein comprising the amino acid sequence of SEQ ID NO:9 and P/CAF. Alternatively, the system can comprise a cell extract produced from cells producing both CBP and P/CAF. Determination of the binding of CBP to P/CAF can be carried out as taught herein.

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The present invention further contemplates a bioassay for screening substances for the ability to stimulate the histone acetyltransferase activity of P/CAF comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance; determining the amount of histone acetylation by P/CAF in the presence of the substance; and comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, an increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can stimulate the histone acetyltransferase activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to stimulate the histone acetyltransferase activity of P/CAF can be added to this system and assayed for stimulating ability.

The present invention further contemplates a bioassay for screening substances for the ability to stimulate the transcription modulating activity of P/CAF comprising contacting a system, in which histone acetylation by P/CAF can be determined, with the substance; determining the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, an increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can stimulate the transcription modulating activity of P/CAF. The acetylation of histones by P/CAF can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P/CAF protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples.

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Thus, the compound to be tested for the ability to stimulate the transcription modulating activity of P/CAF by increasing the histone acetyltransferase activity of P/CAF can be added to this system and assayed for stimulating ability.

The present invention further provides a bioassay for screening substances for the ability to stimulate binding of p300 to P/CAF, comprising contacting a system in which the binding of p300 to P/CAF can be determined, with the substance under conditions whereby the binding of p300 to P/CAF can occur; determining the amount of p300 binding to P/CAF in the presence of the substance; and comparing the amount of p300 binding to P/CAF in the presence of the substance with the amount of p300 binding to P/CAF in the absence of the substance, an increased amount of p300 binding to P/CAF in the presence of the substance indicating a substance that can stimulate the binding of p300 to P/CAF. The binding of p300 to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the p300 protein comprising the amino acid sequence of SEQ ID NO:3 and P/CAF. Alternatively, the system can comprise a cell extract produced from cells producing both p300 and P/CAF. Determination of the binding of p300 to P/CAF can be carried out as taught herein.

Additionally provided in the present invention is a bioassay for screening substances for the ability to stimulate the binding of CBP to P/CAF, comprising contacting a system in which the binding of CBP to P/CAF can be determined, with the substance under conditions whereby the binding of CBP to P/CAF can occur; determining the amount of CBP binding to P/CAF in the presence of the substance; and 25 comparing the amount of CBP binding to P/CAF in the presence of the substance with the amount of CBP binding to P/CAF in the absence of the substance, an increased amount of CBP binding to P/CAF in the presence of the substance indicating a substance that can stimulate the binding of CBP to P/CAF. The binding of CBP to P/CAF can be determined in a system, for example, which can include a cell free reaction mixture comprising a fragment of the CBP protein comprising the amino acid sequence of SEO ID NO 9 and P/CAF. Alternatively, the system can comprise a cell

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extract produced from cells producing both CBP and P/CAF. Determination of the binding of CBP to P/CAF can be carried out as taught herein.

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# Transcription modulating activity of P/CAF

The present invention contemplates a method for inhibiting the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity inhibiting amount of a substance in a pharmaceutically acceptable carrier. For example, the substance can be identified according to the protocols provided herein as one that can inhibit the transcription modulating activity of P/CAF by preventing the binding of P/CAF to p300/CBP or by inhibiting the histone acetyltransferase activity of P/CAF as well as by any other inhibitory mechanism as identified by the protocols provided herein. Inhibition of the transcription modulating activity of P/CAF in a subject is desirable, for example, to inhibit HIV TAT-mediated transcription and therefore, the method of the present invention can be used to treat HIV-infected subjects.

The substance can be in a pharmaceutically acceptable carrier. By "pharmaceutically acceptable" is meant a material that is not biologically or otherwise undesirable, i.e., the material may be administered to a subject, along with the substance, without causing any undesirable biological effects or interacting in a deleterious manner with any of the other components of the pharmaceutical composition in which it is contained. The carrier would naturally be selected to minimize any degradation of the active ingredient and to minimize any adverse side effects in the subject.

The transcription modulating activity and/or histone acetyltransferase activity of P/CAF can be inhibited in a subject by administering to the subject a substance which binds p300/CBP at the P/CAF binding site or a substance which binds the P/CAF protein at the p300/CBP binding site, the ultimate result being that P/CAF and p300/CBP do not bind with one another and P/CAF cannot exert its transcription modulating and/or histone acetyltransferase effect. The substance can be a protein, such as an antibody which binds the P/CAF protein binding site at or near the p300/CBP

binding site, thereby preventing its binding or an antibody which binds the p300/CBP protein at or near the P/CAF binding site, thereby preventing its binding. The substance can also bind the histone acetyltransferase site on P/CAF or at the acetylation site on the histone, thereby preventing acetylation by P/CAF.

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The substance which binds p300/CBP, the P/CAF protein or the histone and has the net effect of inhibiting the transcription modulating effect and or histone acetyltransferase activity of P/CAF in the cell can be delivered to a cell in the subject by mechanisms well known in the art.

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Alternatively, a nucleic acid encoding a protein which binds either to p300/CBP or the P/CAF protein and has the net effect of inhibiting the transcription modulating effect and/or histone acetyltransferase activity of P/CAF in the cell can be delivered to a cell in the subject by gene transduction mechanisms well known in the art. For example, nucleic acid can be introduced by liposomes as well as via retroviral or adeno-associated viral vectors; as described below.

The substance which inhibits the transcription modulating effect and/or histone acetyltransferase activity of P/CAF can be an antisense RNA or an antisense DNA which binds the RNA or DNA of P/CAF, thereby preventing translation or transcription of the RNA or DNA encoding P/CAF and having the net effect of inhibiting the transcription modulating effect and/or histone acetyltransferase activity of P/CAF by inhibiting P/CAF production. The antisense RNA of the present invention can be generated from the nucleic acid of SEQ ID NO:14 (human) or SEQ ID NO:15 (mouse). Furthermore, the antisense DNA can be a phosphorothioate oligodeoxyribonucleotide having the nucleotide sequence of SEQ ID NO:16 (human) or of SEQ ID NO:17 (mouse). The mouse antisense RNA can be used to inhibit the activity of mouse P/CAF, having the nucleotide sequence of SEQ ID NO:18 and the amino acid sequence of SEQ ID NO:8. The present invention also contemplates an antisense nucleic acid sequence which can bind the DNA or RNA of any of the transcription factors or other proteins now known or later identified to bind P/CAF, thereby inhibiting expression of the gene products of



these proteins and having the net effect of inhibiting the transcription modulating effect and/or histone acetyltransferase activity of P/CAF.

The antisense nucleic acid can comprise a typical nucleic acid, but the antisense nucleic acid can also be a modified nucleic acid or a derivative of a nucleic acid such as a phosphorothioate analogue of a nucleic acid. The composition can comprise, for example, an antisense RNA that specifically binds an RNA encoded by the gene encoding the serum protein. Antisense RNAs can be synthesized and used by standard methods (62).

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Antisense RNA can inhibit gene expression by forming an RNA/RNA duplex between the antisense RNA and the RNA transcribed from the target gene. The precise mechanism by which this duplex formation decreases the production of the protein encoded by the endogenous gene probably involves binding of complementary regions of the normal sense mRNA and the antisense RNA strand with duplex formation in a manner that blocks RNA processing and translation. Alternative mechanisms include the formation of a triplex between the antisense RNA and duplex DNA or the formation of an DNA-RNA duplex with subsequent degradation of DNA-RNA hybrids by RNAse H. Furthermore, an antigene effect can result from certain DNA-based oligonucleotides via triple-helix formation between the oligomer and double-stranded DNA which results in the repression of gene transcription. Regardless of the specific molecular mechanism, the present invention results in inhibition of expression of the P/CAF gene by the introduced and replicated DNA resulting in inhibition of the transcription modulating and/or histone acetyltransferase activity of P/CAF, by a reduction in the expression of the nucleic acid to which the antisense nucleic acid is hybridized, and therefore a reduction of the gene product from the targeted gene.

The antisense nucleic acid may be obtained by any number of techniques known to one skilled in the art. One method of constructing an antisense nucleic acid is to synthesize a recombinant antisense DNA molecule. For example, oligonucleotide synthesis procedures are routine in the art and oligonucleotides coding for a particular



protein or regulatory region are readily obtainable through automated DNA synthesis. A nucleic acid for one strand of a double-stranded molecule can be synthesized and hybridized to its complementary strand. One can design these oligonucleotides such that the resulting double-stranded molecule has either internal restriction sites or appropriate 5' or 3' overhangs at the termini for cloning into an appropriate vector. Double-stranded molecules coding for relatively large proteins or regulatory regions can be synthesized by first constructing several different double-stranded molecules that code for particular regions of the protein or regulatory region, followed by ligating these DNA molecules together. Once the appropriate DNA molecule is synthesized, this DNA can be cloned downstream of a promoter in an antisense orientation. Techniques such as this are routine in the art and are well documented.

An example of another method of obtaining an antisense nucleic acid is to isolate that nucleic acid from the organism in which it is found and clone it in an antisense orientation. For example, a DNA or cDNA library can be constructed and screened for the presence of the nucleic acid of interest. Methods of constructing and screening such libraries are well known in the art and kits for performing the construction and screening steps are commercially available (for example, Stratagene Cloning Systems, La Jolla, CA). Once isolated, the nucleic acid can be directly cloned into an appropriate vector in an antisense orientation, or if necessary, be modified to facilitate the subsequent cloning steps. Such modification steps are routine, an example of which is the addition of oligonucleotide linkers which contain restriction sites to the termini of the nucleic acid. General methods are set forth in Sambrook *et al.* (39).

The DNA that is introduced into the cell is in an expression orientation that is antisense to a corresponding endogenous DNA or RNA of the cells. For example, where an endogenous DNA comprises a gene which encodes for a particular protein, the introduced DNA is in an expression orientation opposite the expression of the endogenous DNA, that is the DNA operatively linked to a promoter is in an antisense expression orientation relative to the corresponding endogenous gene. The introduced DNA may be homologous to the entire transcribed gene or homologous to only part of

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the transcribed gene. Alternatively, the sequence of the introduced DNA may be divergent to that of the endogenous DNA but only divergent to the extent that hybridization of the nucleic acids occurs, thereby preventing transcription. One skilled in the art can determine the maximum extent of this divergence by routine screening of antisense DNAs corresponding to an endogenous DNA of the cell. In this manner, one skilled in the art can readily determine which fragments, or alternatively the extent of homology of the fragments or the entire gene that is necessary to inhibit gene expression.

The antisense nucleic acids of the present invention can be made according to protocols standard in the art, as well as described in the Examples provided herein. The antisense nucleic acids can be administered to a subject according to the gene transduction protocols standard in the art, as described below.

The present invention also contemplates a method for stimulating the transcription modulating activity and/or histone acetyltransferase activity of P/CAF in a subject comprising administering to the subject a substance, in a pharmaceutically acceptable carrier, determined according to the methods taught herein, to have a stimulatory affect on the transcription modulating and/or histone acetyltransferase activity of P/CAF. The substance can be one which has been identified, according to the protocols provided herein, to stimulate histone acetyltransferase activity in P/CAF or promote binding of P/CAF to p300/CBP. The stimulation of the transcription modulation activity and/or histone acetyltransferase activity of P/CAF in a subject is desirable, for example, to activate tumor suppressor p53 (which promotes apoptosis) or to activate the muscle differentiation factor, MyoD. Thus, the method of the present invention can be employed to treat cancer and to promote muscle differentiation in conditions where muscle differentiation is desired. The substance can be delivered to a cell in the subject by mechanisms well known in the art.

Further contemplated in the present invention is a method for promoting binding of P/CAF to p300/CBP in a subject, comprising administering to the subject a substance

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identified by the methods provided herein to promote binding of P/CAF to either p300 or CBP.

Additionally, a nucleic acid encoding a protein which stimulates the transcription modulating activity and/or histone acetyltransferase activity of P/CAF can be delivered to a cell in the subject by gene transduction mechanisms, as described below.

Also provided in the present invention is a method of inhibiting the cell cycle progression inducing effect of an oncoprotein which binds p300/CBP in a subject 10 comprising transducing the cells of the subject with a vector comprising a nucleic acid encoding the P/CAF protein; inducing expression of the nucleic acid in the cell to produce the P/CAF in an amount which will allow the P/CAF gene product to replace the oncoprotein bound to p300/CBP, whereby the replacement of the oncoprotein bound to p300/CBP by the P/CAF gene product inhibits the cell cycle progression inducing effect of the oncoprotein. The oncoprotein which binds p300/CBP in the cell can be the adenovirus E1A oncoprotein.

A method for providing a functional P/CAF protein to a subject in need of the functional P/CAF protein is also provided, comprising transducing the cells of the subject with a vector comprising a nucleic acid encoding the P/CAF protein and inducing expression of the nucleic acid to produce the functional P/CAF protein in the cell, thereby providing the functional P/CAF protein to the subject. The transduction of the vector nucleic acid into the subject's cells can be carried out according to standard gene therapy protocols well known in the art (see, for example, U.S. Patent No. 5,339,346).

#### Screening assays for p300/CBP

The present invention also provides a bioassay for screening substances for the ability to inhibit the histone acetyltransferase activity of p300/CBP comprising contacting a system, in which histone acetylation by p300/CBP can be determined, with the substance under conditions whereby histone acetylation by p300/CBP can occur;



determining the amount of histone acetylation by p300/CBP in the presence of the substance, and comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, a decreased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can inhibit the histone acetyltransferase activity of p300/CBP. The acetylation of histones by p300/CBP can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the P300/CBP protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to inhibit the histone acetyltransferase activity of p300/CBP can be added to this system and assayed for acetyltransferase inhibiting ability.

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Also provided in the present invention is a bioassay for screening substances for the ability to inhibit the binding of a transcriptional factor to p300/CBP, comprising contacting a system in which the binding of a transcriptional factor to p300/CBP can be determined, with the substance under conditions whereby the binding of the transcriptional factor and p300/CBP can occur; determining the amount of transcriptional factor binding to p300/CBP in the presence of the substance, and comparing the amount of transcriptional factor binding to p300/CBP in the presence of the substance with the amount of transcriptional factor binding to p300/CBP in the absence of the substance, a decreased amount of transcriptional factor binding to p300/CBP in the presence of the substance indicating a substance that can inhibit the binding of a transcriptional factor to p300/CBP. The binding of a transcriptional factor to p300/CBP can be determined in a system, for example, which can include a cell free reaction mixture comprising a transcriptional factor which binds p300/CBP and p300/CBP. Alternatively, the system can comprise a cell extract produced from cells producing both a transcriptional factor which binds p300/CBP and p300/CBP. The transcriptional factor which binds p300/CBP can be selected from, but is not limited to



the group consisting of nuclear hormone receptors, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YYI, Sap-1a, c-Fos, MyoD and SRC-1, as well as any other transcriptional factor now known or later identified to bind p300/CBP. The screening assay of the present invention can also be used to identify substances which inhibit the binding of p300/CBP to other components to which it is known to bind, for example, P/CAF, pp90<sub>RSK</sub>, TFIIB, E1A, SV40 large T antigen, as well as any other substances now known or later identified to bind p300/CBP. Determination of the binding of a transcriptional factor or other substance to p300/CBP can be carried out as taught in the Examples herein as well as by protocols described in the literature.

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The present invention further contemplates a bioassay for screening substances for the ability to stimulate the histone acetyltransferase activity of p300/CBP comprising contacting a system, in which histone acetylation by p300/CBP can be determined, with the substance, determining the amount of histone acetylation by p300/CBP in the presence of the substance; and comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, an increased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can stimulate the histone acetyltransferase activity of p300/CBP. The acetylation of histones by p300/CBP can be determined in a system including, for example, either core histones (histones H2A, H2B, H3 and H4) or the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) as substrates, the p300/CBP protein and radiolabeled acetyl-CoA (e.g., [1-14C]acetyl CoA). The presence of acetylated histones can be detected by autoradiography after separation by SDS-PAGE as described herein in the Examples. Thus, the compound to be tested for the ability to stimulate the histone acetyltransferase activity of p300/CBP can be added to this system and assayed for stimulating ability.

The present invention further provides a bioassay for screening substances for the ability to stimulate binding of a component, which binds p300/CBP, to p300/CBP, comprising contacting a system in which the binding of the component to p300/CBP can

be determined, with the substance under conditions whereby the binding of the component to p300/CBP can occur; determining the amount of component binding to p300/CBP in the presence of the substance; and comparing the amount of component binding to p300/CBP in the presence of the substance with the amount of component binding to p300/CBP in the absence of the substance, an increased amount of component binding to p300/CBP in the presence of the substance indicating a substance that can stimulate the binding of the component to p300/CBP. The binding of the component to p300/CBP can be determined in a system, for example, which can include a cell free reaction mixture comprising the component and p300/CBP. Alternatively, the system can comprise a cell extract produced from cells producing both the component and p300/CBP. The component which binds p300/CBP can be any of the transcriptional factors or other proteins which are known or are identified in the future to bind p300/CBP, as set forth above. Determination of the binding of the component to p300/CBP can be carried out as taught in the Examples provided herein and according

#### Histone acetyltransferase activity of p300/CBP

to protocols available in the literature.

A method for inhibiting the histone acetyltransferase activity of p300/CBP in a subject is provided in the present invention, comprising administering to the subject a histone acetyltransferase activity inhibiting amount of a substance in a pharmaceutically acceptable carrier. The mechanism of the inhibitory action of the substance can be the inhibition of the binding of a DNA-binding transcription factor, such as, for example, a nuclear hormone receptor, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YY1, Sap-1a, c-Fos, MyoD or SRC-1, to p300/CBP.

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The histone acetyltransferase activity of p300/CBP can be inhibited in a subject by administering to the subject a substance which binds p300/CBP at the transcription factor binding site or a substance which binds the transcription factor protein at the p300/CBP binding site, the ultimate result being that the transcription factor and p300/CBP do not bind with one another and p300/CBP cannot acetylate histones.

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The substance which binds either to the transcription factor or the p300/CBP protein and has the net effect of inhibiting the histone acetyltransferase activity of p300/CBP in the cell can be identified according to the screening methods provided herein and delivered to a cell in the subject by mechanisms well known in the art. The substance can be a protein, such as an antibody which binds the p300/CBP protein binding site at or near the DNA-binding transcription factor binding site, thereby preventing its binding or an antibody which binds the DNA-binding transcription factor at or near the p300/CBP binding site, thereby preventing its binding. The substance can also bind the histone acetyltransferase site on p300/CBP (aa 1195-1673 on p300 or aa 1174-1850 on CBP) or at the acetylation site on the histone, thereby preventing acetylation by p300/CBP.

Additionally, the substance can be a nucleic acid which can be expressed in the cell to produce a protein which inhibits the histone acetyltransferase activity of p300/CBP. For example, a nucleic acid encoding a protein which binds either to a transcription factor or the p300/CBP protein and has the net effect of inhibiting the histone acetyltransferase activity of p300/CBP in the cell can be delivered to a cell in the subject by gene transduction mechanisms well known in the art. For example, nucleic acid can be introduced by liposomes as well as via retroviral or adeno-associated viral vectors, as described below.

The substance which inhibits the histone acetyltransferase activity of p300/CBP can be an antisense RNA or an antisense DNA which binds the RNA or DNA of p300/CBP thereby preventing translation or transcription of the RNA or DNA encoding p300/CBP and having the net effect of inhibiting the histone acetyltransferase activity of P/CAF by inhibiting p300/CBP production. The antisense RNA or DNA of the present invention can be produced and introduced into cells according to the same methods as set forth above for P/CAF antisense nucleic acids.

The present invention also contemplates a method for stimulating the histone acetyltransferase activity of p300/CBP in a subject comprising administering to the

subject a histone acetyltransferase activity stimulating amount of a substance, in a pharmaceutically acceptable carrier, determined according to the methods taught herein, to have a stimulatory affect on the histone acetyltransferase activity of p300/CBP. The substance can exert a stimulatory effect by promoting the binding of a DNA-binding transcription factor of the present invention to p300/CBP. The substance can be delivered to a cell in the subject by mechanisms well known in the art. A nucleic acid encoding a protein which stimulates the transcription modulating activity of p300/CBP can be delivered to a cell in the subject by gene transduction mechanisms, as described below.

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#### Gene transduction

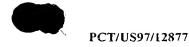
In the methods described above which include gene transduction into cells (i.e., addition of exogenous DNA into cells), the nucleic acids of the present invention can be in a vector for delivering the nucleic acids to the site for expression of the P/CAF protein. The vector can be one of the commercially available preparations, such as the pGM plasmid (Promega). Vector delivery can be by liposome, using commercially available liposome preparations or newly developed liposomes having the features of the present liposomes. Additionally, vector delivery can be via a viral system, including, but not limited to, retroviral, adenoviral and adeno-associated viral systems. Other delivery methods can be adopted and routinely tested according to the methods taught herein.

The modes of administration of the liposome will vary predictably according to the disease being treated and the tissue being targeted. For example, for treating cancer in either the lung or the liver, which are both sinks for liposomes, intravenous delivery is reasonable. For other localized cancers, as well as precancerous conditions, catheterization of an artery upstream from the target organ is a preferred mode of delivery, because it avoids significant clearance of the liposome by the lung and liver. For cancerous lesions at a number of other sites (e.g., skin cancer, localized dysplasias), topical delivery is expected to be effective and may be preferred, because of its convenience.

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Leukemias and other disorders involving dysregulated proliferation of certain isolatable cell populations may be more readily treated by ex vivo administration of the nucleic acid.

The liposomes may be administered topically, parenterally (e.g., intravenously), by intramuscular injection, by intraperitoneal injection, transdermally, extracorporeally or the like, although intravenous or topical administration is typically preferred. The exact amount of the liposomes required will vary from subject to subject, depending on the species, age, weight and general condition of the subject, the severity of the disease being treated, the particular compound used, its mode of administration and the like. Thus, it is not possible to specify an exact amount. However, an appropriate amount may be determined by one of ordinary skill in the art using only routine experimentation given the teachings herein.

Parenteral administration, if used, is generally characterized by injection. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution of suspension in liquid prior to injection, or as emulsions. A more recently revised approach for parenteral administration involves use of a slow release or sustained release system such that a constant level of dosage is maintained. See, e.g., U.S. Patent No. 3,610,795, which is incorporated by reference herein.

Topical administration can be by creams, gels, suppositories and the like. Ex vivo (extracorporeal) delivery can be as typically used in other contexts.

The present invention is more particularly described in the following examples which are intended as illustrative only since numerous modifications and variations therein will be apparent to those skilled in the art.

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#### **EXAMPLES**

#### I. P/CAF studies.

# 5 Cloning and characterization of P/CAF protein.

In human cells, CBP binds to c-Jun in a phosphorylation-dependent manner in association with stimulation of transcription (9). In yeast, GCN4 is believed to be a c-Jun counterpart on the basis of similarities in DNA recognition (15) as well as the participation of both proteins in UV signaling pathways (16). Yeast genetic screening has led to the isolation of various cofactors for GCN4, including GCN5 (yGCN5), ADA2 (yADA2) and ADA3 (yADA3) (17-19). These factors are considered to function as a complex (or in a common pathway) based on genetic and protein-protein interaction studies (18-22). Finally, p300/CBP and yADA2 exhibit significant sequence similarity within a 50 amino acid region including a Zn<sup>2+</sup> finger motif (3). Human counterparts to yGCN5, yADA2, or yADA3 that interact with p300/CBP to mediate transcriptional activation by c-Jun were searched for in various nucleotide sequence databases.

Comparison of the yGCN5 protein sequence with various databases (23) revealed significant similarities with the two randomly sequenced human cDNAs, ETS05039 (24) (P=4.0x10<sup>-15</sup>) and NIB2000-5R (P=6.5x10<sup>-9</sup>). Given that these cDNAs were truncated, human fetal liver and fetal brain cDNA libraries (Clontech) were screened with ETS05039 and NIB2000-5R, respectively and complete clones were isolated from the human fetal liver cDNA library. The complete sequences revealed that the ETS05039- and NIB2000-5R-derived clones are encoded by distinct genes but are highly related within the protein coding regions (68% identity at the DNA level, 75% identity and 86% similarity at the protein level). The former encodes an N-terminal region with no sequence similarity to any proteins in the databases besides the yGCN5-related C-terminal region, whereas the latter encodes only the yGCN5-related region. Given that p300/CBP-binding activity was observed in the former polypeptide as shown below, it was designated p300/CBP-associated factor (P/CAF), having the amino acid

sequence of SEQ ID NO:1 and the nucleotide sequence of SEQ ID NO:10 and the latter was named human GCN5 (hGCN5), having the amino acid sequence of SEQ ID NO:5 and the nucleotide sequence of SEQ ID NO:11.

Additionally, an RNA blot (Clontech) was hybridized with a random-primed probe made from the cDNA encoding P/CAF. RNA blotting indicated that transcripts detected by the P/CAF and hGCN5 cDNAs are ubiquitously expressed, but the former is most abundant in heart and skeletal muscle, whereas the latter is most abundant in pancreas and skeletal muscle.

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# P/CAF-p300/CBP interaction in vitro

The P/CAF binding site was presumed to reside in the C terminal one third of CBP (residues 1,678-2,442) because it was observed that this region, when fused to a DNA binding domain, activates transcription (4) in a manner repressed by coexpression of 12S E1A. This region was divided into 6 overlapping fragments and each was expressed in *E. coli* as a glutathione-S-transferase (GST) fusion protein. GST-CBP fusions were incubated with recombinant P/CAF protein and, subsequently, purified using glutathione-Sepharose. Co-purified P/CAF was detected by immunoblotting analysis.

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To construct GST-fusions, various regions of CBP and p300 were amplified by PCR. A series of deletions of the CBP segment B was created by site-directed *in vitro* mutagenesis (30). These fragments were subcloned into pGEX-2T (Pharmacia). GST-fusions were expressed in *E. coli* and extracted with buffer B [20 mM Tris-HCl (pH 8.0), 5 mM MgCl<sub>2</sub>, 10% glycerol, 1 mM AEBSF, 0.1% NP40, 10 μg/ml of aprotinin, 10 μg/ml of leupeptin, 1 μg/ml of pepstatin A, 1 mM DTT] containing 0.1 M KCl for these experiments. GST-CBP-segment B was purified by glutathione-Sepharose and phenyl-Sepharose chromatographic steps, P/CAF, hGCN5, and E1A were expressed as FLAG-fusions in Sf9 cells via baculovirus vectors and affinity-purified with M2-agarose (ref. 30; Kodak-IBI). For interaction, a crude *E. coli* extract containing 20 pmol of GST-fusion was incubated with 40-60 pmol of P/CAF or E1A in a total volume of 50 μl of

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buffer B with 0.1 M KCl on ice for 10 min. Samples were further incubated with 10 µl (packed volume) of glutathione-Sepharose at 4°C for 30 min, washed four times with 200 µl of buffer B containing 0.1 M KCl, and eluted with 20 µl of buffer E [50 mM Tris-HCl (pH 8.0), 0.2 M KCl, 20 mM glutathione] for 60 min. Interacting proteins were detected by anti-FLAG immunoblotting or silver staining.

For p300 interactions, the segment spanning residues 1763-1966 (segment B') of p300, which is analogous to the CBP segment-B, was used. Twenty percent of the P/CAF and hGCN5 inputs and 100% of the E1A input were also analyzed. In the GST precipitation assays, almost identical amounts of the GST fusions were recovered in all samples. Interaction between P/CAF and CBP (segment B) was determined in the absence and in the presence of E1A. Control reactions with GST-CBP alone and without GST-CBP were also performed. Input proteins were analyzed.

Two CBP segments, A and B, interacted specifically with P/CAF. The stronger interaction was observed in the latter segment, which does not include the yADA2-like Zn<sup>2+</sup> finger. Given that the CBP segment-B is well conserved in p300 (66% identity, 75% similarity), the binding of P/CAF to p300 *in vitro* was also analyzed. For this experiment, the p300 segment spanning residues 1763-1966, termed segment B', which is analogous to the CBP segment-B, was used. Like CBP, p300 interacted specifically with P/CAF. These studies demonstrated that P/CAF binds specifically to both p300 and CBP *in vitro*. In contrast to P/CAF, hGCN5 did not bind to CBP or p300.

These studies also demonstrated that the Zn<sup>2+</sup> finger region of p300/CBP, which shares sequence similarity with yADA2, is not essential for the interaction with P/CAF. Cloning of a human structural homolog of yADA2, termed hADA2 (25) has revealed that, unlike the sequence similarity between p300/CBP and yADA2, which is restricted to a 50 amino acid region, hADA2 shares extensive similarity (30% identity, 52% similarity) to yADA2 over the entire protein sequence. Moreover, a computer search of the complete genomic sequence of *Saccharomyces cerevisiae* revealed that yeast does



not have counterparts of p300/CBP or P/CAF. Thus, the p300/CBP-P/CAF pathway may have been acquired during metazoan evolution.

#### Action of E1A in vitro 5

Previous reports indicated that E1A binds to both the p300 segment spanning residues 1767-1816 and the CBP segment spanning residues 1805-1854 (7). These interactions were reconfirmed in the present system; thus, both p300 and CBP segments covering the previously identified regions interacted with E1A.

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For further mapping, a series of deletions was introduced within the CBP segment-B and tested for interactions with P/CAF and E1A. Deletions of residues 1801-1825 or 1824-1851 markedly reduced interactions with both P/CAF and E1A, whereas deletion of residues 1850-1878 did not affect these interactions. Furthermore, deletion of residues 1801-1851 completely abolished interactions with both P/CAF and E1A. These data indicate that residues 1801-1851 of CBP are critical for interaction with both P/CAF and E1A. Taken together with the evidence that CBP segment A (aa residues 1,678-1,880) also binds to these factors, the above findings demonstrate that P/CAF and E1A bind to the same or very closely spaced sites on CBP.

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Evidence that both P/CAF and E1A recognize the same p300/CBP segments raises the possibility of direct competition between P/CAF and E1A for binding to p300/CBP. To test this possibility, a competition experiment was performed with the use of affinity purified recombinant proteins. The interaction of P/CAF with the CBPsegment B was progressively inhibited by the addition of increasing amounts of E1A. In contrast, no inhibition was caused by an E1A mutant which does not bind to p300/CBP (E1AΔN). Similar results were obtained with the p300-segment B', leading to the conclusion that P/CAF and E1A compete for the same binding sites in p300/CBP.

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#### P/CAF-p300/CBP interaction in vivo

The *in vivo* interaction between P/CAF and p300/CBP was established by co-immunoprecipitation from a human osteosarcoma cell extract. Proteins in this extract were immunoprecipitated with rabbit anti-P/CAF, rabbit anti-CBP and anti-p300 antibodies. For controls, cell extract was precipitated with rabbit control IgG or mouse anti-HA monoclonal antibody. The precipitates were analyzed by immunoblotting with anti-P/CAF, anti-CBP and anti-p300 antibodies.

Osteosarcoma cells were transfected with either control vector or E1A- or

E1AΔN-expression vectors. Extract from the transfected subpopulation was immunoprecipitated with anti-P/CAF or control IgG. The precipitates were analyzed by immunoblotting with anti-p300 and anti-P/CAF antibodies.

Rabbit anti-P/CAF antibody was raised to the P/CAF segment spanning residues 125-397 and purified by immunoaffinity chromatography (33). A mixture of 15 monoclonal antibodies raised to the human p300 segment spanning residues 1572-2371 (5) and rabbit polyclonal antibodies raised to the mouse CBP segment spanning residues 2-23 (for immunoprecipitation) and 1736-2179 (immunoblotting) were purchased from Upstate Biotechnology. Approximately 2 x 10<sup>7</sup> human osteosarcoma U-2 OS cells (ATCC accession number HTB 96) were extracted with 10 ml of lysis buffer [25 mM 20 HEPES-KOH (pH 7.2), 150 mM potassium acetate, 2 mM EDTA, 1 mM DTT, 1 mM AEBSF, 10 μg/ml of aprotinin, 10 μg/ml of leupeptin, 1 μg/ml of pepstatin A, 20 mM sodium fluoride, 0.1% NP40]. Two to 10 ml of extract were incubated with 2 µg of the respective antibody for four hours at 4°C. Fifty µl (packed volume) of protein-A Trisacryl (Pierce) were added and incubation was continued for two hours. The matrix was washed four times with 1 ml of the lysis buffer, then boiled in 2x SDS sample buffer. Human osteosarcoma U-2 OS cells were transfected with 20 µg of the indicated plasmid and 1 µg of sorting plasmid (pCMV-IL2R)(3.1). The transfected subpopulation was purified by magnetic affinity cell sorting (32). Extract from approximately 2 x 10<sup>5</sup> sorted cells was immunoprecipitated as described. 30

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Anti-P/CAF antibody specifically detected a 95 kDa protein, which is very close to the calculated value for the full-length P/CAF, in the immunoprecipitates. Anti-P/CAF antibody co-immunoprecipitated both CBP and p300. Similarly, anti-CBP antibody also co-immunoprecipitated P/CAF. However, anti-p300 antibody did not co-immunoprecipitate P/CAF. This is most likely due to steric interference since the anti-p300 antibody was raised to the p300 segment spanning residues 1572-2371 which includes the P/CAF binding region. These data demonstrate that P/CAF forms complexes with both p300 and CBP *in vivo*.

#### 10 Action of E1A in vivo

The *in vitro* experiments described herein indicate that P/CAF and E1A compete for the binding sites in p300/CBP. Thus, a study was conducted to determine whether E1A targets the endogenous interaction between P/CAF and p300. An E1A-expression vector was transiently transfected into human osteosarcoma cells and the transfected subpopulation was purified by cell sorting. Then, the interaction between P/CAF and p300 in transfected cells was examined by co-immunoprecipitation with anti-P/CAF antibody. The endogenous interaction of P/CAF with p300 was drastically inhibited by expression of E1A. On the other hand, no inhibition was observed by the E1A mutant lacking the p300 binding domain (E1AΔN), indicating that E1A disrupts the P/CAF-p300 complex *in vivo* through an interaction with p300.

# 'Cell cycle regulation by P/CAF

Given that binding of P/CAF to p300/CBP is inhibited by E1A, experiments were performed to evaluate whether P/CAF, by binding to and forming a functional complex with p300, is involved in the regulation of entry into S phase. This possibility was addressed by examining whether transient expression of P/CAF would affect the rate of G1/S transit in HeLa cells. P/CAF negatively affected the distribution of cells between G1 and S phases in this assay.

HeLa cells were transfected by electroporation with 7  $\mu g$  of P/CAF-expression plasmid and/or 3  $\mu g$  of the full-length or the N-terminally deleted ( $\Delta 2$ -36) E1A 12S-



expression plasmid as indicated. These plasmids were constructed by subcloning FLAG-P/CAF and E1A cDNAs into pCX (34) and pcDNAI (Invitrogen), respectively. All samples, in addition, contained 1 µg of sorting plasmid (pCMV-IL2R) (31) and carrier plasmid (pCX) to normalize the total amount of DNA to 11 µg. After transfection, cells were incubated in Dulbecco's modified Eagle's medium with 10% fetal bovine calf serum for 12 h, and subsequently labeled in medium containing 10 µM bromo-deoxyuridine (BrdU) for 30 min. Subsequently, the transfected subpopulation was purified by magnetic affinity cell sorting and nuclei were analyzed by dual parameter flow cytometry as described (32).

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The fraction of cells accumulating in S phase in control cultures was 23%, compared to 15% in P/CAF-transfected cells. This effect was reproducible in multiple independent experiments. In parallel experiments to verify the utility of this experimental protocol, plasmids encoding E2F-1, simian virus 40 small t, cyclin A or cyclin E increased the accumulation of cells in S phase, whereas plasmids encoding the cyclin-dependent kinase inhibitors p21 or p27 reduced the number of S phase cells.

On the basis of evidence that E1A and P/CAF compete for binding sites on p300, it seemed possible that cotransfection of P/CAF with E1A would oppose the mitogenic effect caused by E1A. As shown by the data herein, this is indeed the case. E1A alone has mitogenic activity in this experimental setting, while the E1A mutant lacking the p300 binding domain (E1AAN) has very weak activity. Comparable expression levels between wild type and mutant E1A in the transfected cells were revealed by immunoblotting analysis with anti-E1A. Intriguingly, when P/CAF was cotransfected with E1A, the mitogenic activity of E1A was significantly counteracted by P/CAF. These results show that P/CAF and E1A mediate antagonistic effects on cell cycle progression.

In the course of assessing P/CAF activity, it was also revealed that p300 is able to inhibit cell cycle progression under the same assay conditions. These findings suggest

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that P/CAF and p300, perhaps by forming a complex, act in concert to suppress cell cycle progression.

# Histone acetyltransferase activity in P/CAF

Acetylation of the N-terminal histone tails has been considered to play a crucial role in accessibility of transcription factors to nucleosomal templates (26-27). Recently, yGCN5 has been identified as a histone acetyltransferase (28). On the basis of this information, intrinsic histone acetyltransferase activity in P/CAF and hGCN5 was examined. As substrates, the core histones (histones H2A, H2B, H3 and H4) and the nucleosome core particles (146 base pairs of DNA wrapped around the octamer of core histones) were used.

Activity of hGCN5 and P/CAF that acetylates free histones or histones in the nucleosome core particle (35) was measured as described (36). Each reaction contained 0.3 pmol of affinity purified FLAG-hGCN5 or FLAG-P/CAF, 4 pmol of the histone octamer or the nucleosome core particle and 10 pmol of [1-14C]acetyl-CoA. The histone octamer dissociated into dimers or tetramers under assay conditions. Acetylated histones were detected by autoradiography after separation by SDS-PAGE.

P/CAF and hGCN5 acetylated the core histones with almost the same efficiency. Both factors acetylated histones H3 and H4, but preferentially H3. In contrast, very weak or no acetylation by hGCN5 was detected in the nucleosome core particles. Remarkably, significant acetylation by P/CAF was observed in a nucleosomal context. Although all core histones are acetylated in the nucleus, P/CAF and hGCN5 did not acetylate histones H2A and H2B *in vitro*.

Direct function of P/CAF is likely to involve its intrinsic histone acetyltransferase activity. Although exact molecular mechanisms by which acetylation of core histones contribute to transcription remains undefined, acetylation of the histones is considered to play an important role in transcriptional regulation (26-27). The positively charged N-terminal tails of core histones are believed to affect nucleosome structure by interacting



with DNA at or near the nucleosome-spacer junction. Acetylation of the histone tails presumably destabilizes the nucleosome and facilitates access by regulatory factors. Likewise, there is a general correlation between the level of acetylation and transcriptional activity of nucleosomal domains. The findings of the present invention provide insights into the mechanisms of targeted histone acetylation.

Cellular factor p300/CBP binds to various sequence-specific factors that are involved in cell growth and/or differentiation, including CREB (3,4), c-Jun (9), Fos (11), c-Myb (12) and nuclear receptors (13). P/CAF could stimulate the activation function of these factors via promoter-specific histone acetylation. The present invention demonstrates that E1A appears to perturb normal cellular regulation by disrupting the connection between p300/CBP and its associated histone acetyltransferase.

#### II. p300/CBP studies.

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# Purification of E1A associated histone acetyltransferase.

FLAG-epitope tagged E1A (or ΔE1A) was expressed in Sf9 cells (ATCC accession number CRL 1711) by infecting recombinant baculovirus (43). All purification steps were carried out at 4°C. Extract was prepared from infected cells by one cycle of freeze and thaw in buffer B (20 mM Tris-HCl, pH 8.0; 5 mM MgCl<sub>2</sub>, 10% glycerol; 1 mM PMSF; 10 mMβ-mercaptoethanol; 0.1% Tween 20) containing 0.1 M KCl and the complete protease inhibitor cocktail (Boehringer Mannheim). To prepare E1A-immobilized beads, the extract was incubated with M2 anti-FLAG antibody agarose (Kodak-IBI) for four hours with rotating and subsequently washed with the same buffer three times. The resulting beads were incubated with HeLa (ATCC accession number CCL 2) nuclear extract for four to eight hours and thereafter washed with the same buffer six times. Finally, FLAG-E1A was eluted from the beads along with associated polypeptides by incubating with the same buffer containing 0.1 mg/ml FLAG peptide.

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For further purification, eluted polypeptides were dialyzed in 0.05 M KCl-buffer B and subsequently loaded onto a SMART Mono Q column (Pharmacia) equilibrated with the same 0.05 M KCl-buffer B. After washing, the column was developed with a linear gradient of 0.05-1.0 M KCl in buffer B. Mono Q fractions were concentrated with a MICROCON spin-filter (Amicon) and consequently loaded onto a SMART Superdex 200 column (Pharmacia) equilibrated with 0.1 M KCl-buffer B.

#### Histone acetyltransferase assays

Filter binding assays were performed as described (80) with minor modifications. Samples were incubated at 30°C for 10-60 minutes in 30 ml of assay buffer containing 50 mM Tris-HCl, pH 8.0, 10% glycerol, 1 mM DTT, 1 mM PMSF, 10 mM sodium butyrate; 6 pmol of [³H]acetyl CoA (4.3 mCi/mmole, Amersham Life Science Inc.); and 33 mg/ml of calf thymus histones (Sigma Chemical Co.). In experiments where synthetic peptides were substituted for core histones, 50 pmol of each peptide were used. After incubation, the reaction mixture was spotted onto Whatman P-81 phosphocellulose filter paper and washed for 30 minutes with 0.2 M sodium carbonate buffer pH 9.2 at room temperature with 2-3 changes of the buffer. The dried filters were counted in a liquid scintillation counter.

PAGE analysis was done as above except that 90 pmol of [14C] acetyl CoA (55 mCi/mmole, Amersham Life Science Inc.) and 9 pmol of core histones or mononucleosomes were used. Core histones and mononucleosomes were prepared as described (35). For trypsin digestion, reaction mixtures were further incubated with various amounts of trypsin on ice for 30 minutes. The samples were analyzed on one dimensional SDS-PAGE gels or two dimensional gels, where the first dimension was an acid-urea-PAGE gel (44) and the second dimension was an SDS-PAGE gel.

#### Protein expression

For baculovirus expression, cDNAs corresponding to p300 portions of aa 1-670, aa 671-1194 and aa 1135-2414 were amplified by PCR (EXPAND High Fidelity PCR System, Boehringer Mannheim) as KpnI-NotI fragments. The resulting fragments were

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subcloned into a baculovirus transfer vector having the FLAG-tag sequence (43). The recombinant viruses were isolated using the BACULOGOLD system (Pharmingen), according to the manufacturer's protocol and were infected into Sf9 cells (ATCC accession number CRL 1711) to express FLAG-p300. Recombinant proteins were affinity purified with M2 anti-FLAG antibody-immobilized agarose (Kodak-IBI) according to the manufacturer's protocol.

For bacterial expression, cDNAs encoding the p300 portions and the CBP portion (aa 1174-1850) were first subcloned into the baculovirus transfer vector having the FLAG-tag as described above. Thereafter, the XhoI and NotI fragments encoding FLAG-p300 or FLAG-CBP fusions were resubcloned into the *E. coli* expression vector pET-28c (Novagene) digested with SalI and NotI. Recombinant proteins were expressed in *E. coli* BL21(DE3) and affinity purified with M2-antibody agarose.

#### 15 Histone acetyltransferases that associate with E1A

Although the adenovirus E1A 12S protein (E1A) inhibits transcription in a variety of genes via direct binding to p300/CBP (45), E1A also stimulates transcription in some contexts (46). Thus, p300/CBP-bound E1A was tested to determine whether it might recruit histone acetyltransferases or deacetylases to regulate transcription. In addition, experiments were conducted as described below to determine if p300/CBP per se is a histone acetyltransferase.

Initially, recombinant FLAG-epitope tagged E1A was immobilized on anti-FLAG antibody beads. Immobilized E1A was incubated with a HeLa nuclear extract for affinity purification of E1A-associated polypeptides. FLAG-E1A was then eluted from the beads, along with E1A-associated polypeptides, by incubating with FLAG-peptide. Although E1A per se has no histone acetyltransferase activity, E1A recruited significant amounts of histone acetyltransferase activity from the nuclear extract. It is very unlikely that this activity is derived from P/CAF given that E1A and P/CAF cannot bind to p300/CBP simultaneously (43). Consistent with this, no P/CAF was detected in these fractions by immunoblotting.

The E1A N-terminus, a region that is not highly conserved among the various adenovirus serotypes, is involved in p300/CBP binding *in vivo*. Mutations in the N-terminal region lead to loss of the ability for p300/CBP binding without affecting RB binding (1,47). Thus, the requirement of the E1A N-terminal region for the recruitment of histone acetyltransferase activity was tested. In contrast to the wild type, the N-terminal deleted form of E1A ( $\Delta$ N-E1A) recruited only a background level of acetyltransferase activity. In agreement with previous reports (47), the  $\Delta$ N-E1A showed no ability to interact with p300/CBP, although it still retained the ability to interact with a variety of other polypeptides, including RB

To define the relationship between p300/CBP and histone acetyltransferase activity, affinity purified E1A-binding polypeptides were separated by Mono Q ion-exchange column. Both p300/CBP and the acetyltransferase activity were coeluted at 140 mM KCl, while most of polypeptides were eluted at 260 mM KCl. The active fraction of Mono Q column (~140 mM KCl) was further separated by Superdex-200 gel filtration column. Both p300/CBP and the acetyltransferase activity coeluted after the void volume, indicating that p300/CBP is involved in the histone acetyltransferase activity.

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#### p300 is a histone acetyltransferase

The data provided herein indicate that p300 per se, or a polypeptide(s) associated with p300, possesses histone acetyltransferase activity. To test the former possibility, the acetyltransferase activity of recombinant p300 was measured. p300 was divided into three fragments, each of which was expressed in and purified from Sf9 cells via a baculovirus expression vector. Histone acetyltransferase activity was readily detected in the C-terminal fragment containing amino acids 1135-2414, whereas no activity was found in the other fragments, demonstrating conclusively that p300 per se is a histone acetyltransferase:

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# p300/CBP-histone acetyltransferase domain

To map the histone acetyltransferase domain of p300, a series of deletions was prepared. Given the poor conservation of the glutamine-rich region (aa 1815-2414) in the C. elegans p300/CBP homolog (6), the p300 fragment encoding aa 1135-1810 was expressed in and purified from E. coli. Importantly, this candidate region of p300 (aa 1135-1810) showed significant histone acetyltransferase activity. For further mapping within this region, a series of N-terminal deletions was constructed. Deletion of 60 residues, resulting in a fragment containing as 1195-1810, had no effect on the acetyltransferase activity, whereas the deletion of 185 residues, yielding a fragment comprising as residues 1320-1810, completely eliminated the acetyltransferase activity.

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Next, a series of C-terminal deletions was analyzed to determine the requirement of the P/CAF (or E1A) -binding domain. The p300 fragments lacking the E1A binding domain (aa 1195-1760, 1195-1706 and 1195-1673) still retained the acetyltransferase activity, whereas the further truncated mutant (aa 1195-1652) completely lost the acetyltransferase activity. Consistent with these results, the internal deletion of residues 1418-1720 showed no acetyltransferase activity. These data demonstrate that the histone acetyltransferase domain is located between the bromodomain and the E1A-binding domain. Given that the histone acetyltransferase domain is highly conserved between p300 and CBP (91% similarity), the corresponding region of CBP, aa residues 1174-1850, was expressed to confirm the acetyltransferase activity. As expected, comparable activity was detected, indicating that both p300 and CBP are histone acetyltransferases.

Among various acetyltransferases including histone acetyltransferases GCN5 and P/CAF, putative acetyl-CoA binding sites are conserved (48). However, multiple alignment analysis (49) showed that the p300/CBP histone acetyltransferase domain does not belong to this group. Moreover, comparison of the p300/CBP histone acetyltransferase domain with peptide sequence databases (23) showed no sequence similarity to any other proteins. Accordingly, this invention shows that p300/CBP represents a novel class of acetyltransferases in that it does not have the conserved motif found among previously described acetyltransferases (48).





# p300 acetylates all core histones in mononucleosomes

Substrate specificity for acetylation by p300 was also examined. As substrates, histone octamers and mononucleosomes (146 base pairs of DNA wrapped around the octamer of core histones) were used. Given that the histone octamer dissociates into dimers or tetramers under physiological conditions, the histone octamer is referred to here as core histones. When core histones were used, p300 acetylated all four proteins, but preferentially H3 and H4. More importantly, in a nucleosomal context, p300 acetylated all four core histones nearly stoichiometrically. In contrast, p300 acetylated neither BSA nor lysozyme.

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Hyperacetylated histones are believed to be linked with transcriptionally active chromatin (26,27,50,51). Hyperacetylated forms are found in histones H4, H3 and H2B, which have multiple acetylation sites *in vivo*. Thus, the level of acetylation by p300 was also tested.

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Mononucleosomes treated with p300 were analyzed by two-dimensional gel electrophoresis. A Coomassie blue-stained gel and the corresponding autoradiogram showed that a significant amount of histones, especially H4, were hyperacetylated. Importantly, acetylation levels by p300 were very close to those of hyperacetylated histones prepared from HeLa nuclei treated with sodium butyrate, a histone deacetylase inhibitor. In contrast, no acetylated forms were detected in the reaction without p300. These results indicate that p300 acetylates histones in mononucleosomes to the hyperacetylated state by targeting multiple lysine residues.

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# p300 acetylates the four lysines in the histone H4 N-terminal tail in vitro which are acetylated in vivo

Lysines at positions 5, 8, 12 and 16 of histone H4 are acetylated *in vivo* (51). Recent studies with yeast histone acetyltransferases demonstrate the position-specific acetylation by distinct acetyltransferases, i.e., while cytoplasmic acetyltransferases for histone deposition and chromatin assembly modify

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positions 5 and 12, GCN5 modifies positions 8 and 16 (52). Accordingly, the positions of acetylation by p300 were also determined. A series of synthetic peptides containing acetylated lysines at various positions was used to determine the acetylation site-specificity of p300. Consistent with the two-dimensional gel electrophoresis analysis, the experiments with peptide substrates showed that p300 acetylates all four lysines in the histone H4 that are acetylated *in vivo*. These results are consistent with the view that deposition-related diacetylated histones are deacetylated during maturation of chromatin (53).

#### p300 preferentially acetylates the N-terminal histone tail

Histone acetyltransferases modify specific lysine residues in the N-terminal tail of core histones but not the C-terminal globular domain in vivo (26,27,50,51). Structural models of nucleosomes (54,55,56) suggest that most of the lysine residues in the C-terminal globular domain are buried. Therefore, experiments were conducted to examine whether restricted acetylation of the N-terminal tail resulted from the substrate 15 specificity of the enzyme or inaccessibility of the enzyme to the core domain in nucleosomes. The globular domains of all core histones contain a long helix flanked on either side by a loop segment and short helix, termed the "histone fold" (54,55,56). The histone fold is involved in formation of the stable H2A-H2B and H3-H4 hetero-dimers, consisting of extensive hydrophobic contacts between the paired 20 molecules. Therefore, it is likely that a histone monomer cannot fold properly, thereby increasing access of the histone acetyltransferase to the core domain. Based on these considerations, experiments were conducted to determine whether p300 acetylates free histone H4 in a N-terminal-specific manner.

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Histone H4 was acetylated with p300 and subsequently the histone tail was removed by partial digestion with trypsin. The distributions of radioactivity between intact and core histones were compared. While the globular core histone domain was predominant at the higher trypsin concentrations, radioactivity was detected mostly in the intact histone. These data demonstrate that p300 preferentially acetylates the N-terminal tail of histone H4.

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#### III. P/CAF interaction with MyoD

### Tissue culture and transfection experiments

 $C_2C_{12}$  mouse cells (ATCC accession number CRL 1772) were grown in Dulbecco's modified Eagle medium (DMEM) supplemented with 20% fetal bovine serum (FBS) until they reached confluence. Differentiation was induced by switching medium to differentiation medium (DM), consisting of DMEM containing 2% horse serum.  $C_3H/10T1/2$  fibroblasts (ATCC accession number CCL 226) were grown in DMEM supplemented with 10% FBS. Cells were transfected by the calcium phosphate precipitation method. Total amounts of transfected DNA were equalized by empty vector DNA. After 12 h incubation in medium containing the precipitated DNA, the cells were washed and incubated in fresh DMEM containing 10% FBS for an additional 24 h. Afterwards, differentiation was induced by incubating in DM for 36 to 72 h. Chloramphenicol acetyltransferase (CAT) assays were performed as previously described (64,69). The quantities of cell extracts used for CAT assays were normalized to  $\beta$ -galactosidase activity by cotransfection of 1 mg of the  $\beta$ -galactosidase expression vector, pON260.

Expression vectors used for transfection experiments are as follows:

pCX-P/CAF for P/CAF (43), pCMV-bp300 for p300 (65), pCMV-p300 (1869-2414)

(64) and pCMV-p300 (1514-1922) (60) for p300 wild type and mutants, pE1A12S, pE1A12S R2G, pE1A12S D2-36 and pE1A12S D121-130 for E1A wild type and mutants (66,67,68), and pEMSV-MyoD for MyoD (64).

The antisense P/CAF RNA expression vector, pcDNA3 P/CAF-AS, was created as follows. The 2.5 Kb EcoRI-KpnI fragment containing the entire P/CAF open reading frame was isolated from pCX-P/CAF (43). This fragment was subcloned into the

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EcoRI-KpnI sites of plasmid pcDNA3 (Invitrogen) so that the antisense P/CAF RNA is driven under the CMV promoter. Reporter genes employed were 4RE-CAT and MCK-CAT (69). 4RE-CAT is driven by a synthetic promoter containing 4 copies of the E-box, whereas MCK-CAT is driven by the native MCK promoter (nucleotides -1256 to +7).

# Microinjection and immunofluorescence

Cells were grown on small glass slides, subdivided into numbered squares of 2 mm x 2 mm and microinjected with purified and concentrated antibodies, as previously described (70). For immunofluorescence, cells were fixed in either 2% paraformaldehyde or 1:2 methanol/acetone solution, preincubated with 5% BSA/PBS and incubated with the primary antibodies for 30 min at 37° C. Subsequently, antibody was visualized by incubating with either rhodamine- or fluorescein-conjugated secondary antibody for 30 min at 37° C. Injected antibodies were stained with a rhodamine-conjugated secondary antibody and nuclei were counter-stained by DAPI as previously described (69).

Antibodies employed are as follows; rabbit polyclonal affinity purified affti-P/CAF antibody (43), rabbit polyclonal anti-p300/CBP antiserum (71), mouse monoclonal anti-MyoD antibody (clone 5.8A, kindly provided by P. Houghton), goat polyclonal anti-c-Jun affinity purified antibody (Santa Cruz) and rabbit pre-immune serum.

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# Immunoprecipitation and DNA affinity purification

Cells were resuspended in lysis buffer (20 mM NaPO<sub>4</sub>, 150 mM NaCl, 5mM MgCl<sub>2</sub>, 0.1% NP40, 1 mM DTT, 10 mM sodium fluoride, 0.1 mM sodium vanadate, 1 mM phenylmethylsulfonyl-fluoride and 10 mg/ml each of leupeptin, aprotinin and pepstatin). After 30 min incubation on ice, samples were centrifuged at 12,000 x g for 30 min and supernatants were used as cell extracts. Extracts were pre-cleared by

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for 2 h at 4 C. For immunoprecipitation, the supernatants were incubated with the respective antibodies for 3 h at 4 C. Protein A/G Plus-Agarose was added, and incubation continued for 3 h. The matrix was washed with lysis buffer, then boiled in 2 X SDS sample buffer. Immunoblotting was performed by using the ECL chemiluminescent detection kit (Amersham) according to the manufacturer's protocol.

Affinity purification of E-box-bound complexes was done as previously described (69). Briefly, 100 ng of the biotinylated double stranded DNA containing the E-box were immobilized on streptavidin-conjugated magnetic beads and incubated with 500 mg of cell extracts in the presence of poly dI-dC. After extensive washing, bound proteins were eluted with SDS sample buffer and analyzed by immunoblotting.

#### In vitro protein-protein interaction assays

The CBP-B fragment and its deletion derivatives were expressed as 15 GST-fusions described previously (43). MyoD and E1A (43) were expressed as FLAG-fusion proteins in Sf9 cells via a baculovirus expression system and affinity-purified on M2 anti-FLAG antibody-agarose (Kodak-IBI). Crude E. coli extracts containing GST-fusions were incubated with various amounts of MyoD and/or E1A in 50 ml of buffer B (20 mM Tris-HCl, pH 8.0, 0.1 M KCl, 5 mM MgCl<sub>2</sub>, 10% 20 glycerol, and 0.1% Nonidet P-40) on ice for 10 min. GST-precipitation was performed as described (43). MyoD and E1A were detected by immunoblotting with anti-FLAG M2 antibody. For the interaction between P/CAF and MyoD, 1.5 pmol of FLAG-P/CAF and 15 pmol of FLAG-MyoD were incubated in 50 ml of buffer B on ice for 10 min. The mixture was further incubated with 2 mg of anti-P/CAF (43) or 25 anti-hADA2 antibody for 60 min. The immunocomplexes were precipitated by incubation with the nil of protein A-Trisacryl (Pierce) and rotated for 1-4 hr at 4oC. The matrix was washest 4 times with 200 ml of buffer B and boiled in 10 ml of 2 X SDS sample buffer. The proteins were resolved on a 4%-20% gradient SDS-PAGE and subjected to immunoblotting with the anti-FLAG M2 antibody. The blot was developed 30 with the SUPERSIGNAL chemiluminescent substrates (Pierce).

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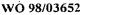


# P/CAF coactivates muscle-specific transcription

P/CAF and MyoD were co-transfected into mouse C3H10T1/2 fibroblasts, and MyoD-mediated transcription was determined from reporter activity driven by the artificial (4RE) and the naturally-occurring muscle creatine kinase (MCK) promoters. Overexpression of P/CAF stimulated MyoD-dependent transcription several folds in both promoters. Similar results were obtained for the myoD activated myogenin promoter Transcriptional activation was further stimulated by co-transfecting with MyoD, P/CAF and p300 expression vectors, suggesting that P/CAF may function by forming a complex with p300/CBP. Consistent with the lack of DNA binding capacity in P/CAF, overexpression of P/CAF alone did not increase the basal transcriptional activity of either enhancer. To test whether P/CAF and p300/CBP function in the same pathway, two dominant negative forms of p300 were employed which specifically inhibit p300/CBP-mediated transcription (60,64). The p300 segment spanning residues 1514-1922 inhibits the MyoD-dependent activation via direct interaction with MyoD (60), whereas the p300 segment spanning residues 1869-2414 inhibit it without direct interaction (64). Both dominant negative mutants inhibited MyoD-coactivation by P/CAF), suggesting that P/CAF and p300/CBP function in the same pathway.

20 -For further elucidation of the activation mechanism by P/CAF, the effect of E1A, which inhibits MyoD-dependent transcription and differentiation (66,72,73) via direct interaction with p300/CBP (65,78), was tested. Expression of E1A in C3H10T1/2 fibroblasts inhibited stimulation of MyoD-directed transcription by P/CAF overexpression. E1A mutants lacking p300/CBP-binding activity, E1A D2-36 and E1A R2G (67,79), had almost no effect. On the other hand, an E1A mutant retaining p300/CBP-binding activity, E1A D121-130, behaved like the wild type. Since E1A associates with p300/CBP, but not with P/CAF, these results suggest that P/CAF functions in MyoD-directed transcription via interaction with p300/CBP.

To address the role of P/CAF as a myogenic coactivator in a more relevant environment, P/CAF was overexpressed in proliferating C2C12 myoblasts which express







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endogenous myogenic bHLH factors. As observed in fibroblasts, overexpression of P/CAF stimulated muscle specific transcription. Concomitant expression of exogenous p300 increased P/CAF-mediated coactivation. The repression exerted by wild type E1A, but not mutant E1A D2-36, on P/CAF coactivation of MyoD was also observed in muscle cells.

Similar experiments were performed with myogenic cell lines that were stably transformed with wild type or mutant E1A-expressing vectors (66). Coactivation by P/CAF was inhibited by wild type E1A or the E1A mutant that retains

10 p300/CBP-binding activity (E1AΔ121-130). In contrast, E1A mutants that lack p300/CBP-binding (E1A Δ2-36 and E1A R2G) allowed transcriptional coactivation by P/CAF. Taken together, these experiments show that P/CAF coactivates MyoD-directed transcription via interaction with p300/CBP.

# 15 P/CAF stimulates myogenic differentiation

Given that P/CAF potentiates MyoD-directed transcription, the ability of P/CAF to assist MyoD in promoting myogenic differentiation was investigated. To this aim, C3H10T1/2 fibroblasts were transiently transfected with P/CAF and MyoD expression vectors. An expression vector for the green fluorescent protein (GFP) was

20 co-transfected to identify transfected cells. After incubation in differentiation medium, the myogenic conversion of transfected cells was determined by simultaneous expression of the GFP and the differentiation-specific marker myosin heavy chain (MHC). Forced expression of MyoD in fibroblasts caused muscle differentiation in 12% of the transfected fibroblasts. This myogenic conversion was 20% by co-expressing MyoD and P/CAF. As observed in transcription experiments, stimulation of differentiation by P/CAF was counteracted by co-transfection with the p300 dominant negative mutant, p300 (1869-2414). Consistent with a general role for coactivators, overexpression of P/CAF alone was unable to differentiate fibroblasts.

Similar experiments were done using proliferating C2C12 myoblasts in which the differentiation program is already committed. Most of the myoblasts differentiated into

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myotubes by overexpressing P/CAF, whereas only a modest effect was observed by overexpressing p300. In contrast, differentiation was inhibited slightly by overexpressing c-Jun. This inhibitory effect presumably was caused by titration of p300/CBP, which associates directly with c-Jun (74). A similar inhibition was observed in the p300 dominant negative mutant. Consistent with the transcriptional effect, E1A almost completely inhibited differentiation. The E1A mutant RG2, lacking p300/CBP-binding capability but retaining the retinoblastoma protein (Rb)-binding capability, only partially inhibited differentiation, although this same mutant inhibited transcription as severely as the wild type. Taken together, these data show that P/CAF stimulates muscle differentiation by coactivating MyoD function via p300/CBP association.

# P/CAF is essential for myogenic transcription and differentiation

To test the necessity of P/CAF for myogenic transcription, experiments were conducted whereby P/CAF synthesis was inhibited by expressing antisense P/CAF RNA. A vector from which the P/CAF mRNA is transcribed in the antisense orientation (P/CAF-AS) was transfected with P/CAF and MyoD expression vectors into fibroblasts and MyoD-dependent transcription was examined. Cotransfection of the antisense expression vector strongly inhibited MyoD-dependent transcription below the level of induction elucidated by MyoD alone, demonstrating that expression of P/CAF antisense RNA inhibits not only the coactivation exerted by exogenous P/CAF but also that of endogenous P/CAF. These results indicate that P/CAF is essential for MyoD-dependent transcription.

Studies were also carried out to determine whether expression of P/CAF antisense RNA inhibits myogenic differentiation. C3H10T1/2 fibroblasts were transiently transfected with various expression vectors with or without the P/CAF antisense RNA expression vector. Expression of P/CAF antisense RNA reduced MyoD-mediated myogenic conversion of fibroblasts. Expression of P/CAF antisense RNA also counteracted the stimulatory effect of both P/CAF and p300 on myogenic differentiation. These data support the view that P/CAF and p300/CBP coactivate



MyoD-dependent transcription in the same pathway. More drastic inhibition was observed in C2C12 myoblasts in similar experiments. Therefore, it can be concluded that P/CAF is essential for transcription of muscle specific genes and hence differentiation into myotubes.

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To further confirm the essential role of P/CAF for myogenic differentiation, we blockage experiments by antibody microinjection were performed. Antibodies were injected into the cytoplasm of proliferating C2C12 myoblasts to prevent the nuclear transport of newly synthesized target proteins. After incubating in the differentiation medium, the degree of differentiation was determined. Microinjection of an anti-P/CAF antibody almost completely inhibited differentiation. Similar results were obtained by microinjecting anti-p300/CBP antibodies. Although microinjection of either anti-p300/CBP or P/CAF antibody was sufficient to inhibit differentiation, an even greater inhibition was observed by coinjecting both of them. Microinjection of anti-P/CAF or anti-p300/CBP antibody did not interfere with induction of p53 by DNA damaging agents, showing specificity of the inhibition by the antibodies. In contrast to anti-P/CAF or anti-p300/CBP antibodies, the injection of anti-MyoD antibody only partially inhibited differentiation, supporting the view of functional redundancy between MyoD and Myf-5 (75,76). Injection of anti-c-Jun antibody or control antibody did not interfere with muscle differentiation.

Similar experiments were performed with C3H10T1/2 fibroblasts stably expressing MyoD. In these cells, either anti-p300/CBP or anti-P/CAF antibody completely inhibited muscle differentiation. In contrast to myoblasts, anti-MyoD antibody completely blocked differentiation in the fibroblasts expressing MyoD. Anti-c-Jun and control antibodies did not interfere with differentiation. Taken together, these results demonstrate that P/CAF and p300/CBP are indispensable for activation of the myogenic program.



# p300/CBP, P/CAF and MyoD form a multimeric complex in vivo

The data described above indicate that P/CAF stimulates MyoD-directed transcription via association with p300/CBP. Thus, experiments were conducted to investigate whether P/CAF, p300/CBP and MyoD could associate in a complex.

First, cellular extracts derived from C2C12 myotubes were subjected to immunoprecipitation. Both anti-MyoD and anti-p300/CBP antibodies co-precipitated P/CAF. In a complementary experiment, both anti-p300/CBP and anti-P/CAF antibodies also co-precipitated MyoD, suggesting that these factors form a multimeric protein complex in myotubes.

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Next, attempts were made to detect this complex on the E-box, the DNA binding site for MyoD. Immobilized DNA containing an E-box sequence was incubated with myotube extracts. After extensive washing, P/CAF, p300/CBP and MyoD were analyzed by immunoblotting. P/CAF, p300/CBP and MyoD were all affinity purified on the immobilized DNA, whereas they were not purified on the control DNA lacking the E-box. Given that P/CAF and p300/CBP per se cannot bind to DNA, these observations indicate that P/CAF and p300/CBP are recruited through MyoD at the E-box sites to form a multi-protein complex.

# 20 Complex formation is inhibited by viral transforming factors

Since the oncoviral proteins E1A and large T antigen inhibit myogenic transcription and differentiation, the effect of these factors on the formation of complexes on the E-box was tested. Importantly, very small amouts of P/CAF and p300/CBP were co-purified on the E-box from myocyte extracts which stably express E1A or large T antigen, although MyoD was detected under these conditions. The lower recovery of MyoD from E1A-expressing muscle cells could reflect the low level of MyoD in the extracts (66). These results indicate that E1A and large T antigen dissociate P/CAF and p300/CBP from MyoD without altering MyoD binding to DNA.

Consistent with the previous observations that transiently expressed E1A prevents interaction between P/CAF and p300/CBP in vivo (43), the association



between p300/CBP and P/CAF was abolished in myoblasts stably transformed by wild type E1A but not in those clones transformed with the E1A mutant R2G unable to bind p300/CBP. Similarly, the interaction between p300/CBP and P/CAF was abolished by large T antigen but not by the mutant protein that localizes into the cytoplasm (77).

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# Interaction between MyoD, P/CAF and CBP in vitro

Previous interaction experiments *in vitro* indicate that the CBP region spanning residues 1801 to 1850 is crucial for interaction with both P/CAF and E1A (43). While most sequence-specific factors bind to CBP sites distinct from the P/CAF/E1A binding sites, MyoD interacts with an overlapping CBP fragment called the CH3 region (60,64,65). To understand how P/CAF, p300/CBP and MyoD associate, the CBP sites important for MyoD binding were mapped more precisely. Consistent with previous reports (60,64,65), the CBP fragment spanning residues 1801-2000 (fragment B) bound MyoD. Moreover, deletion of residues 1801 to 1850 within fragment B completely abolished interaction with MyoD, which is similar to the results obtained with P/CAF and E1A. Importantly, an internal deletion of residues 1850-1878 abolished the MyoD interaction with CBP, while it did not affect binding of E1A or P/CAF (43). These results suggest that MyoD and P/CAF bind to distinct sites of p300/CBP, albeit the binding sites may overlap. Moreover, a direct interaction was observed between MyoD and P/CAF, which may contribute to stabilization of the multimeric complex.

These data show that E1A prevents not only p300/CBP-interaction with P/CAF but also that with MyoD in vivo. To obtain evidence that this inhibition is due to the direct action by E1A, competition experiments were performed in vitro. Importantly, the interaction between CBP and MyoD was strongly inhibited by addition of E1A, implicating that E1A inhibits myogenic transcription by disrupting multiple interactions.

Although the present process has been described with reference to specific details of certain embodiments thereof, it is not intended that such details should be



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regarded as limitations upon the scope of the invention except as and to the extent that they are included in the accompanying claims.

Throughout this application various publications are referenced by numbers

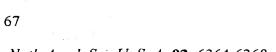
within parentheses. Full citations for these publications are as follows. The disclosures
of these publications in their entireties are hereby incorporated by reference into this
application in order to more fully describe the state of the art to which this invention
pertains.

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#### SEQUENCE LISTING

- (1) GENERAL INFORMATION
- (i) APPLICANT: The United States of America, as repesented by the Secretary, Department of Health and Human Services, c/o National Institutes of Health, Office of Technology Transfer, 6011 Executive Boulevard, Suite 325, Rockville, Maryland 20842
- (ii) TITLE OF THE INVENTION: METHODS AND COMPOSITIONS FOR p300/CBP-ASSOCIATED TRANSCRIPTIONAL CO-FACTOR P/CAF
- (iii) NUMBER OF SEQUENCES: 18
- (iv) CORRESPONDENCE ADDRESS:
  - (A) ADDRESSEE: NEEDLE & ROSENBERG, P.C.
  - (B) STREET: Suite 1200, 127 Peachtree Street, NE
  - (C) CITY: Atlanta
  - (D) STATE: GA
  - (E) COUNTRY: USA
  - (F) ZIP: 30303
- (v) COMPUTER READABLE FORM:
  - (A) MEDIUM TYPE: Diskette
  - (B) COMPUTER: IBM Compatible
  - (C) OPERATING SYSTEM: DOS
  - (D) SOFTWARE: FastSEQ for Windows Version 2.0
- (vi) CURRENT APPLICATION DATA:
  - (A) APPLICATION NUMBER:
  - (B) FILING DATE: 23-JUL-1997
  - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
  - (A) APPLICATION NUMBER: Corresponding U.S. Serial No. 60/022,273
  - (B) FILING DATE: 23-July-1996
- (viii) ATTORNEY/AGENT INFORMATION:
  - (A) NAME: Miller, Mary L
  - (B) REGISTRATION NUMBER: 39,303
  - (C) REFERENCE/DOCKET NUMBER: 14014.0238/P
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  - (A) TELEPHONE: 404/688-0770
  - (B) TELEFAX: 404/688-9880
  - (C) TELEX:
  - (2) INFORMATION FOR SEQ ID NO:1:
- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 832 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None





# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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1				5					10					Gly 15	
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Pro	Pro	Ala 35	Pro	Pro	Gln	Gly	Ser 40	Pro	Cys	Ala	Ala	Ala 45	Ala	Gly	Gly
Ser	Gly		Cys	Gly	Pro	Ala 55	Thr	Ala	Val	Ala	Ala 60	Ala	Gly	Thr	Ala
Glu 65	Gly	Pro	Gly	Gly	Gly 70		Ser	Ala	Arg	Ile 75	Ala	Val	Lys	Lys	Ala 80
Gln	Leu	Arg	Ser	Ala 85	Pro	Arg	Ala	Lys	Lys 90	Leu	Glu	Lys	Leu	Gly 95	Val
Tyr	Ser	Ala	Cys 100	Lys	Ala	Glu	Glu	Ser 105	Cys	Lys	Cys	Asn	Gly 110	Trp	Lys
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Ile	Val 130		Leu	Thr	Glu	Ser 135	Cys	Arg	Ser	Cys	Ser 140	His	Ala	Leu	Ala
Ala 145	His	Val	Ser	His	Leu 150	Glu	Asn	Val	Ser	Glu 155	Glu	Glu	Met.	Așn	Arg 160
Leu	Leu	Gly	Ile	Val 165	Leu	Asp	Val	Glu	Tyr 170	Leu	Phe	Thr	Cys	Val 175	His
Lys	Glu	Glu	Asp 180	Ala	Asp	Thr	Lys	Gln 185	Val	Tyr	Phe	Tyr	Leu 190	Phe	Lys
		195	_				200					205		Glu	
	210		-			215					220			Gln	
225					230					235				Lys	240
-				245					250					Ile 255	
-	•		260					265					270	Pro	
_	-	275					280					285		Cys	
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305					310					315				Met	320
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		355					360					365		Gln	
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				405					410					Cys 415	
			420					425					430		
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	450					455					460			Asp	
Ala 465		Met	. Leu	Gly	Pro 470		Thr	Asn	Phe	Leu 475		Ala	His	Ser	Ala 480



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### (2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 481 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:



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	_			85		Glu			90					95	•
Asp	Ile	Pro	Met 100	Glu	Leu	Ile	Asn	Glu 105	Val	Met	Ser	Thr	Ile 110	Thr	Asp
		115			_	Pro	120					125			
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		_		165		Leu			170					175	
			180			Ile		185					190		į.
-		195				Lys	200					205			
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225					230	Val				235					240
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-		-	260	_		Ile		265					270		
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-				325		Gln			330					335	
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_	370					Asp 375					380				
385					390	His				395	•				400
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	_		420			Ala		425					430		
	_	435				Glu	440					445			
-	450					455 Ser					460				
Leu 465 Lys	GIU	гÀг	rne	FILE	470	ser	пуз	TIG	пур	475	VIG	Gry			480

# (2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 203 amino acids

  - (B) TYPE: amino acid
    (C) STRANDEDNESS: single



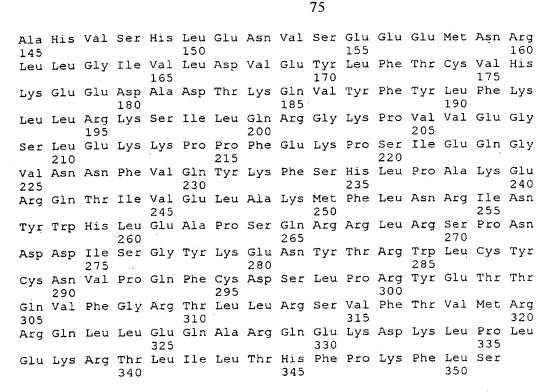
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Arg Val Val Gln His Thr Lys Gly Cys Lys Arg Lys Thr Asn Gly Gly 10 Cys Pro Ile Cys Lys Gln Leu Ile Ala Leu Cys Cys Tyr His Ala Lys 20 His Cys Gln Glu Asn Lys Cys Pro Val Pro Phe Cys Leu Asn Ile Lys 40 Gln Lys Leu Arg Gln Gln Gln Leu Gln His Arg Leu Gln Gln Ala Gln 55 Met Leu Arg Arg Arg Met Ala Ser Met Arg Thr Gly Val Val Gly Gln 75 70 Gln Gln Gly Leu Pro Ser Pro Thr Pro Ala Thr Pro Thr Thr Pro Thr 90 85 Gly Gln Gln Pro Thr Thr Pro Gln Thr Pro Gln Pro Thr Ser Gln Pro 105 100 Gln Pro Thr Pro Pro Asn Ser Met Pro Pro Tyr Leu Pro Arg Thr Gln 125 120 Ala Ala Gly Pro Val Ser Gln Gly Lys Ala Ala Gly Gln Val Thr Pro 140 135 Pro Thr Pro Pro Gln Thr Ala Gln Pro Pro Leu Pro Gly Pro Pro 155 150 Thr Ala Val Glu Met Ala Met Gln Ile Gln Arg Ala Ala Glu Thr Gln 170 165 Arg Gln Met Ala His Val Gln Ile Phe Gln Arg Pro Ile Gln His Gln 180 185 Met Pro Pro Met Thr Pro Met Ala Pro Met Gly 200

- (2) INFORMATION FOR SEQ ID NO:4:
- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 351 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Ser Glu Ala Gly Gly Ala Gly Pro Gly Gly Cys Gly Ala Gly Ala 10 Gly Ala Gly Ala Gly Pro Gly Ala Leu Pro Pro Gln Pro Ala Ala Leu 25 Pro Pro Ala Pro Pro Gln Gly Ser Pro Cys Ala Ala Ala Gly Gly Ser Gly Ala Cys Gly Pro Ala Thr Ala Val Ala Ala Ala Gly Thr Ala ~5.5 Glu Gly Pro Gly Gly Gly Ser Ala Arg Ile Ala Val Lys Lys Ala 75 Gln Leu Arg Ser Ala Pro Arg Ala Lys Lys Leu Glu Lys Leu Gly Val 90 Tyr Ser Ala Cys Lys Ala Glu Glu Ser Cys Lys Cys Asn Gly Trp Lys 100 105 Asn Pro Asn Pro Ser Pro Thr Pro Pro Arg Ala Asp Leu Gln Gln Ile 120 125 Ile Val Ser Leu Thr Glu Ser Cys Arg Ser Cys Ser His Ala Leu Ala 135





#### (2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 476 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

#### (ii) MOLECULE TYPE: None

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

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Ile Lys Asp Gly Arg Val Ile Gly Gly Ile Cys Phe Arg Met Phe Pro 200 195 Thr Gln Gly Phe Thr Glu Ile Val Phe Cys Ala Val Thr Ser Asn Glu 215 220 Gln Val Lys Gly Tyr Gly Thr His Leu Met Asn His Leu Lys Glu Tyr 230 235 His Ile Lys His Asn Ile Leu Tyr Phe Leu Thr Tyr Ala Asp Glu Tyr 250 245 Ala Ile Gly Tyr Phe Lys Lys Gln Gly Phe Ser Lys Asp Ile Lys Val 265 270 Pro Lys Ser Arg Tyr Leu Gly Tyr Ile Lys Asp Tyr Glu Gly Ala Thr 285 280 275 Leu Met Glu Cys Glu Leu Asn Pro Arg Ile Pro Tyr Thr Glu Leu Ser 300 295 His Ile Ile Lys Lys Gln Lys Glu Ile Ile Lys Lys Leu Ile Glu Arg 310 315 Lys Gln Ala Gln Ile Arg Lys Val Tyr Pro Gly Leu Ser Cys Phe Lys 325 330 Glu Gly Val Arg Gln Ile Pro Val Glu Ser Val Pro Gly Ile Arg Glu 345 350 Thr Gly Trp Lys Pro Leu Gly Lys Glu Lys Gly Lys Glu Leu Lys Asp 365 360 355 Pro Asp Gln Leu Tyr Thr Thr Leu Lys Asn Leu Leu Ala Gln Ile Lys 375 Ser His Pro Ser Ala Trp Pro Phe Met Glu Pro Val Lys Lys Ser Glu 390 395 Ala Pro Asp Tyr Tyr Glu Val Ile Arg Phe Pro Ile Asp Leu Lys Thr 405 410 Met Thr Glu Arg Leu Arg Ser Arg Tyr Tyr Val Thr Arg Lys Leu Phe 425 430 420 Val Ala Asp Leu Gln Arg Val Ile Ala Asn Cys Arg Glu Tyr Asn Pro 445 440 Pro Asp Ser Glu Tyr Cys Arg Cys Ala Ser Ala Leu Glu Lys Phe Phe 455 450 Tyr Phe Lys Leu Lys Glu Gly Gly Leu Ile Asp Lys 470

#### (2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2414 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:



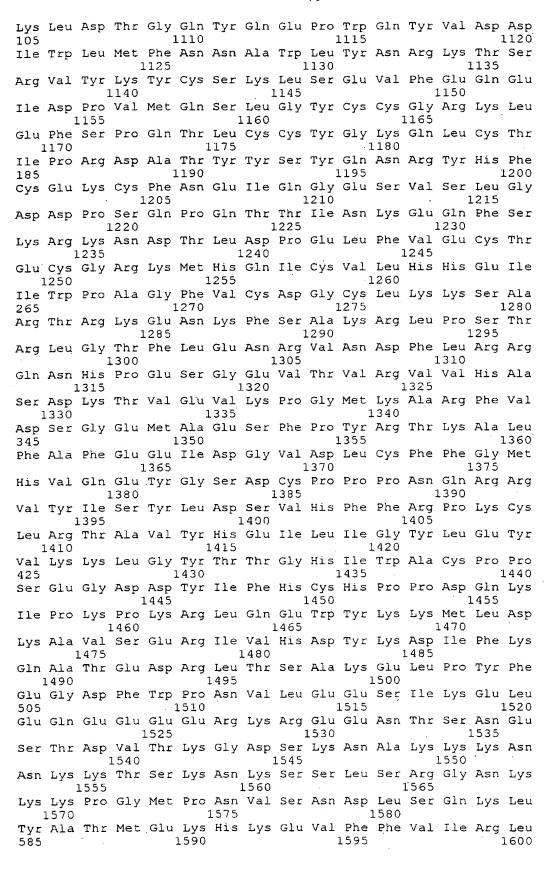
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Gln 145		Pro	Thr	Gln	Ser. 150	Thr	Gly	Met	Met	Asn 155	Ser	Pro	Val	Asn	Gln 160
	Ala	Met	Gly	Met 165	Asn	Thr	Gly	Thr	Asn 170	Aļa	Gly	Met	Asn	Pro 175	Gly
Met	Leu	Ala	Ala 180	Gly	Asn	Gly	Gln	Gly 185	Ile	Met	Pro	Asn	Gln 190	Val	Met
		195					200					205		Туr	
	210	-	•			215					220			Leu	
225	-				230	_				235				Pro	240
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-			260					265					270	Gly	
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	_		_	325					330					Arg 335	
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-	-	355					360					365		Gln	
-	37Ō			_		375					380			Ile	
385	-		=		390					395					400
				405					410					Leu 415	
	-		420					425					430	Thr	
		435	_				440					445		Gln	
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465	_		_		470					475				Gln	480
	•			485					490					Gln 495	
_			500					505					510	Ala	
		515			_	_	520	_				525		Leu	
	530					535					540			Met	
545					550					555				Ala	560
				565					570					11e 575	
			580					585					590	Ile	
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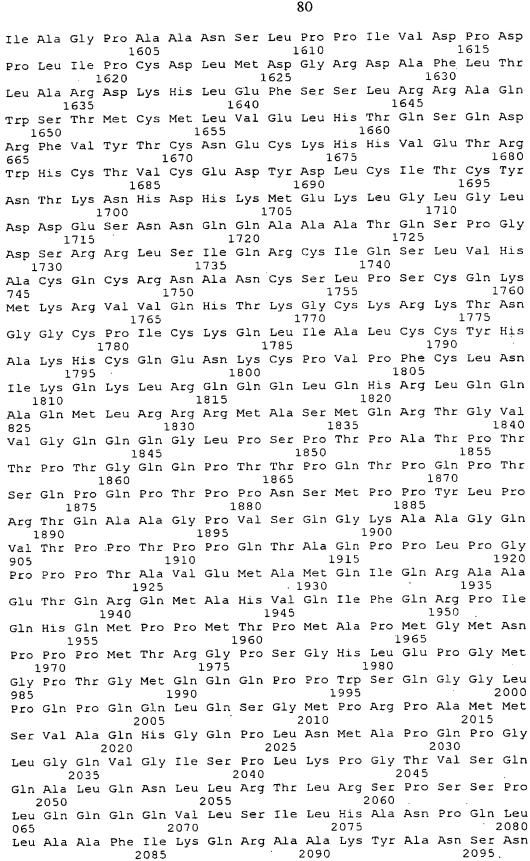




	610	-	Ala			615					620				
Asn 625	Arg	Ala	Glu	Tyr	Tyr 630	His	Leu	Leu	Ala	Glu 635	Lys	Ile	Tyr	Lys	Ile 640
Gln	Lys	Glu	Leu	Glu 645	Glu	Lys	Arg	Arg	Thr 650	Arg	Leu	Gln	Lys	Gln 655	Asn
Met	Leu	Pro	Asn 660	Ala	Ala	Gly	Met	Val 665		Val	Ser	Met	Asn 670	Pro	Gly
Pro	Asn	Met 675	Gly	Gln	Pro	Gln	Pro 680		Met	Thr	Ser	Asn 685	Gly	Pro	Leu
Pro	Asp 690		Ser	Met	Ile	Arg 695		Ser	Val	Pro	Asn 700	Gln	Met	Met <sup>.</sup>	Pro
Arg 705		Thr	Pro	Gln	Ser 710		Leu	Asn	Gln	Phe 715	Gly	Gln	Met	Ser	Met 720
Ala	Gln	Pro	Pro	Ile 725		Pro	Arg	Gln	Thr 730	Pro	Pro	Leu	Gln	His 735	His
Gly	Gln	Leu	Ala 740	Gln	Pro	Gly	Ala	Leu 745	Asn	Pro	Pro	Met	Gly 750	Tyr	Gly
		755	Gln				760					765			
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			Ser	965					970					975	
_			Ala 980					985					990		
		995					1000					1005			
	1010		Thr			1015					1020				
025			Asp		1030					1035					1040
			Ser	1045					1050					1055	
			Pro 1060					1065					1070		
		1075					1080					1085			
Tyr	1090	_	Ile	val	гуs	Ser 1095		Met	. Asp	ьeu	1100		тте	гÀг	Arg









Pro Gln Pro Ile Pro Gly Gln Pro Gly Met Pro Gln Gly Gln Pro Gly 2100 2105 Leu Gln Pro Pro Thr Met Pro Gly Gln Gln Gly Val His Ser Asn Pro 2115 2120 2125 Ala Met Gln Asn Met Asn Pro Met Gln Ala Gly Val Gln Arg Ala Gly 2130 2135 2140 Leu Pro Gln Gln Gln Pro Gln Gln Gln Leu Gln Pro Pro Met Gly Gly 2150 2155 2160 Met Ser Pro Gln Ala Gln Gln Met Asn Met Asn His Asn Thr Met Pro . 2165 2170 2175 Ser Gln Phe Arg Asp Ile Leu Arg Arg Gln Gln Met Met Gln Gln 2180 2185 2190 Gln Gln Gly Ala Gly Pro Gly Ile Gly Pro Gly Met Ala Asn His 2195 2200 2205 Asn Gln Phe Gln Gln Pro Gln Gly Val Gly Tyr Pro Pro Gln Pro Gln 2210 2215 2220 Gln Arg Met Gln His His Met Gln Gln Met Gln Gln Gly Asn Met Gly 2230 2235 Gln Ile Gly Gln Leu Pro Gln Ala Leu Gly Ala Glu Ala Gly Ala Ser 2245 2250 Leu Gln Ala Tyr Gln Gln Arg Leu Leu Gln Gln Met Gly Ser Pro 2260 2265 2270 Val Gln Pro Asn Pro Met Ser Pro Gln Gln His Met Leu Pro Asn Gln 2275 2280 2285 Ala Gln Ser Pro His Leu Gln Gly Gln Gln Ile Pro Asn Ser Leu Ser 2290 2295 2300 Asn Gln Val Arg Ser Pro Gln Pro Val Pro Ser Pro Arg Pro Gln Ser 305 2310 2315 2320 Gln Pro Pro His Ser Ser Pro Ser Pro Arg Met Gln Pro Gln Pro Ser 2325 2330 2335 Pro His His Val Ser Pro Gln Thr Ser Ser Pro His Pro Gly Leu Val 2340 2345 2350 Ala Ala Gln Ala Asn Pro Met Glu Gln Gly His Phe Ala Ser Pro Asp 2355 2360 2365 Gln Asn Ser Met Leu Ser Gln Leu Ala Ser Asn Pro Gly Met Ala Asn 2370 2375 2380 Leu His Gly Ala Ser Ala Thr Asp Leu Gly Leu Ser Thr Asp Asn Ser 385 2390 2395 2400 Asp Leu Asn Ser Asn Leu Ser Gln Ser Thr Leu Asp Ile His

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#### (2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

2405

- (A) LENGTH: 2441 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

 Met Ala Glu Asn Leu Leu Asp Gly Pro Pro Asn Pro Lys Arg Ala Lys 1

 Leu Ser Ser Pro Gly Phe Ser Ala Asn Asp Asn Thr Asp Phe Gly Ser 20

 Leu Phe Asp Leu Glu Asn Asp Leu Pro Asp Glu Leu Ile Pro Asn Gly 35

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 Lys His Lys Gln Leu Ser Glu Leu Leu Arg Gly Gly Ser Gly Ser Ser 70

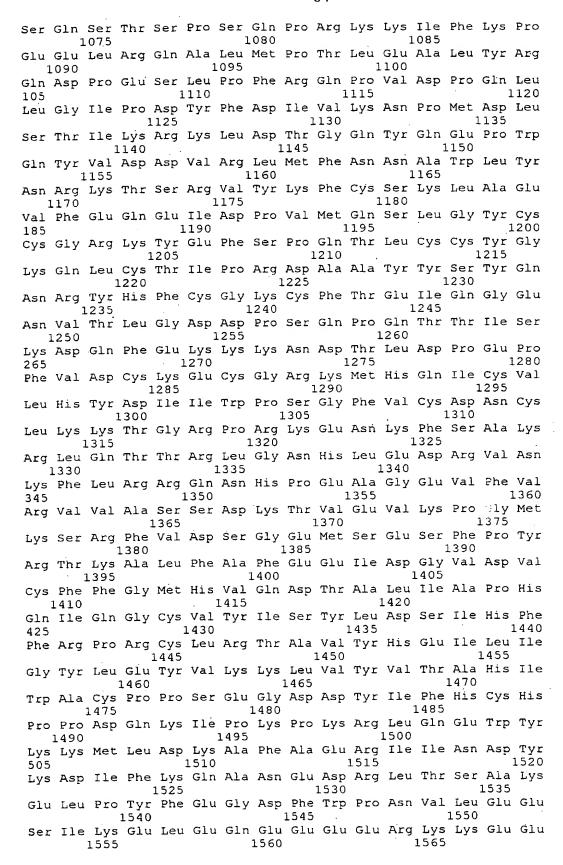


Ile	Asn	Pro	Gly	Ile 85	Gly	Asn	Val	Ser	Ala 90	Ser	Ser	Pro	Val	Gln 95	Gln
Gly	Leu	Gly	Gly 100	Gln	Ala	Gln	Gly	Gln 105	Pro	Asn	Ser	Thr	Asn 110	Met	Ala
Ser	Leu	Gly 115	Ala	Met	Gly	Lys	Ser 120	Pro	Leu	Asn	Gln	Gly 125	Asp	Ser	Ser
Thr	Pro 130	Asn	Leu	Pro	Lys	Gln 135	Ala	Ala	Ser	Thr	Ser 140	Gly	Pro	Thr	Pro
145	Ala				150					155					160
	Thr			165					170					175	
	Ala		180					185					190		
_	His	195					200					205			
	Gly 210					215					220				
225	Pro		•		230					235					240
	Leu			245					250					255	
	Ala		260					265					270		
	Pro	275	_				280					285			
	Thr 290	_				295					300				
305	Leu				310					315					320
	Pro			325					330					335	
	Ala Gln		340					345					350		
	Glu	355					360					365			
-	370 Thr					375					380				
385	Ala		_		390					395					400
-	Trp			405					410					415	
	Asn	_	420	_				425					430		
-	Ser	435					440					445			
	450 Ala	_				455					460				
465	Arg				470					475					480
	Gln			485					490					495	
	His		500					505					510		
	Val	515					520					525			
	530 Ser	•		-	-	535			_		540				
545					550					555					560
uec	M311	ush	GT À	565		JGI	OΙY	. 1311	570		JCL	<u> </u>	JGL	575	-16

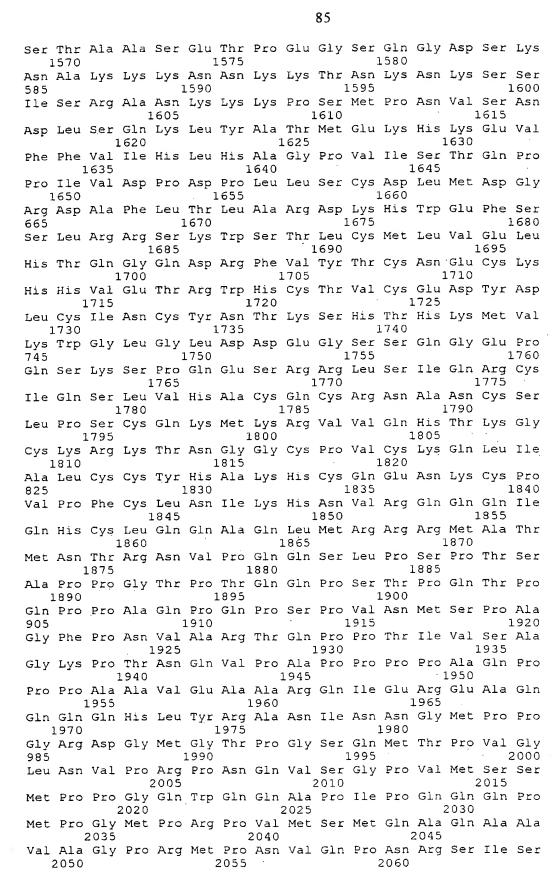


										_		_		-	
Pro	Thr	Ala	Ala 580	Pro	Pro	Ser	Ser	Thr 585	Gly	Val	Arg	Lys	Gly 590	Trp	His
Glu	His	Val 595	Thr	Gln	Asp	Leu	Arg 600	Ser	His	Leu	Val	His 605	Lys	Leu	Val
Gln	Ala 610		Phe	Pro	Thr	Pro 615	Asp	Pro	Ala	Ala	Leu 620	Lys	Asp	Arg	Arg
Met 625		Asn	Leu	Val	Ala 630		Ala	Lys	Lys	Val 635		Gly	Asp	Met	Tyr. 640
	Ser	Ala	Asn	Ser 645		Asp	Glu	Tyr	Tyr 650		Leu	Leu	Ala	Glu 655	
Ile	Tyr	Lys	Ile 660		Lys	Glu	Leu	G1 ú 665		Lys	Arg	Arg	Thr 670		Leu
His	Lys			Ile	Leu	Gly	Asn		Pro	Ala	Leu	Pro 685		Ser.	Gly
Ala		675 Pro	Pro	Val	Ile		680 Pro	Ala	Gln	Ser	Val 700		Pro	Pro	Asn
	690 Pro	Leu	Pro	Leu		695 Val	Asn	Arg	Met	Gln 715		Ser	Gln	Gly	Met 720
705 <b>As</b> n	Ser	Phe	Asn		710 Met	Ser	Leu	Gly.			Gln	Leu	Pro	Gln 735	
Pro	Met	Gly		725 Arg	Ala	Ala	ser		730 Met	Asn	His	Ser	Val 750		Met
Asn	Ser	Met 755	740 Ala	Ser	Val	Pro	Gly 760	745 Met	Ala	Ile	Ser	Pro 765		Arg	Met
Pro	Gln 770		Pro	Asn	Met	Met 775	Gly	Thr	His	Ala	Asn 780		Ile	Met	Ala
		Pro	Thr	Gln	Asn 790		Phe	Leu	Pro	Gln 795		Gln	Phe	Pro	Ser 800
785 Ser	Ser	Gly	Ala	Met 805		Val	Asn	ser	Val 810		Met	Gly	Gln	Pro 815	
Ala	Gln	Ala	Gly 820		Ser	Glņ	Gly	Gln 825		Pro	Gly	Àla	Ala 830		Pro
Asn	Pro	Leu 835		Met	Leu	Ala	Pro 840		Ala	Ser	Gln	Leu 845		Cys	Pro
Pro	Val 850		Gln	Ser	Pro	Leu 855	His	Pro	Thr	Pro	Pro 860		Ala	Ser	Thr
Ala 865		Gly	Met	Pro	Ser 870		Gln	His	Pro	Thr 875		Pro	Gly	Met	Thr 880
	Pro	Gln	Pro	Ala 885		Pro	Thr	Gln	Pro 890		Thr	Pro	Val	Ser 895	
Gly	Gln	Thr	Pro 900	-	Pro	Thr	Pro	Gly 905		Val	Pro	Ser	Ala 910	Ala	Gln
Thr	Gln	Ser 915		Pro	Thr	Val	Gln 920		Ala	Ala	Gln	Ala 925	Gln	Val	Thr
Pro	Gln 930		Gln	Thr	Pro	Val 935	Gln	Pro	Pro	Ser	Val 940	Ala	Thr	Pro	Gln
ser 945		Gln	Gln	Gln	Pro 950		Pro	Val	His	Thr 955	Gln	Pro	Pro	Gly	Thr 960
	Leu	Ser	Gln	Ala 965	Ala	Ala	Ser	Ile	Asp 970	Asn	Arg	Val	Pro	Thr 975	Pro
Ser	Thr	Val	Thr 980	Ser	Ala	Glu	Thr	Ser 985	Ser	Gln	Gln	Pro	Gly 990	Pro	Asp
Val	Pro	Met 995	Leu	Glu	Met	-	Thr 1000	Glu	Val	Gln		Asp 1005	Asp	Ala	Glu
	Glu 1010		T,h r	Glu			Gly	Glu	Pro	_	Ser 1020	Glu	Met	Met	Glu
		Leu	Gln				Gln	Val				Thr	Asp		Thr 1040
	Gln	Lys				Met	Glu				Lys	Lys			
Lys	Val		Ala 1060	Lys	Glu	Glu	Glu			ser	Ser		Asp 1070	Thr	Ala





WO 98/03652 PCT/US97/12877







Pro Ser Ala Leu Gln Asp Leu Leu Arg Thr Leu Lys Ser Pro Ser Ser 2075 2080 2070 Pro Gln Gln Gln Gln Val Leu Asn Ile Leu Lys Ser Asn Pro Gln 2085 2090 2095 Leu Met Ala Ala Phe Ile Lys Gln Arg Thr Ala Lys Tyr Val Ala Asn 2100 2105 2110 Gln Pro Gly Met Gln Pro Gln Pro Gly Leu Gln Ser Gln Pro Gly Met 2115 2120 2125 Gln Pro Gln Pro Gly Met His Gln Gln Pro Ser Leu Gln Asn Leu Asn 2135 2140 2130 Ala Met Gln Ala Gly Val Pro Arg Pro Gly Val Pro Pro Pro Gln Pro 145 2150 2155 2160 Ala Met Gly Gly Leu Asn Pro Gln Gly Gln Ala Leu Asn Ile Met Asn 2165 2170 2175 Pro Gly His Asn Pro Asn Met Thr Asn Met Asn Pro Gln Tyr Arg Glu 2180 2185 2190 Met Val Arg Arg Gln Leu Leu Gln His Gln Gln Gln Gln Gln Gln 2195 2200 2205 Gln Gln Gln Gln Gln Gln Asn Ser Ala Ser Leu Ala Gly Gly 2210 . 2215 2220 Met Ala Gly His Ser Gln Phe Gln Gln Pro Gln Gly Pro Gly Gly Tyr 2230 2235 Ala Pro Ala Met Gln Gln Gln Arg Met Gln Gln His Leu Pro Ile Gln 2245 2250 2255 Gly Ser Ser Met Gly Gln Met Ala Ala Pro Met Gly Gln Leu Gly Gln 2260 2265 2270 Met Gly Gln Pro Gly Leu Gly Ala Asp Ser Thr Pro Asn Ile Gln Gln 2275 2280 2285 Ala Leu Gln Gln Arg Ile Leu Gln Gln Gln Gln Met Lys Gln Gln Ile 2290 2295 2300 Gly Ser Pro Gly Gln Pro Asn Pro Met Ser Pro Gln Gln His Met Leu 305 2310 2315 2320 Ser Gly Gln Pro Gln Ala Ser His Leu Pro Gly Gln Gln Ile Ala Thr 2325 2330 2335 Ser Leu Ser Asn Gln Val Arg Ser Pro Ala Pro Val Gln Ser Pro Arg 2340 2345 2350 Pro Gln Ser Gln Pro Pro His Ser Ser Pro Ser Pro Arg Ile Gln Pro 2360 2365 Gln Pro Ser Pro His His Val Ser Pro Gln Thr Gly Thr Pro His Pro 2370 2375 2380 Gly Leu Ala Val Thr Met Ala Ser Ser Met Asp Gln Gly His Leu Gly 385 2390 2395 2400 Asn Pro Glu Gln Ser Ala Met Leu Pro Gln Leu Asn Thr Pro Asn Arg 2405 2410 2415 Ser Ala Leu Ser Ser Glu Leu Ser Leu Val Gly Asp Thr Thr Gly Asp 2420 2425 Thr Leu Glu Lys Phe Val Glu Gly Leu 2435 2440

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### (2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 813 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: None
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met Ala Glu Ala Gly Gly Ala Gly Ser Pro Ala Leu Pro Pro Ala Pro 1 10 15

Pro	His	Gļy	Ser 20	Pro	Arg	Thr	Leu	Ala 25	Thr	Ala	Ala	Gly	Ser 30	Ser.	Ala
Ser	Cys	Gly 35	Pro	Ala	Thr	Pro	Val 40	Ala	Ala	Ala	Gly	Thr 45	Ala	Glu	Gly
	50	-				55	Arg		,	•	60	·			
Arg 65	Ser	Ala	Pro	Arg	Ala 70	Lys	Lys	Leu	Glu	Lys 75	Leu	Gly.	Val	Tyr	Ser 80
	-	=		85			Cys		90					95	
			100				Arg	105					110		
		115					Ser 120					125			
	130	·				135	Ser				140				
145			* *		150		Tyr			155					160
				165			Val		170					175	_
_	-		180				Gly	185					190		Leu
	-	195					Lys 200 Ser					205			
	210					215	Met				220				
225					230		Arg			235					240
				245			Tyr		250					255	
		_	260				Leu	265					270		
		275					280 Ser					285			
	290					295					300				
305					310		Lys			315	•				320
_				325			Phe Ser		330					335	
			340				Pro	345					350		
		355					360					365			
	370					375	Leu				380				
385					390		Thr			395					400
_				405			Glu		410					415	
			420				Ser	425					430		
		435					Ser 440					445			
	450					455					460				
465					470		Arg			475					480
_				485			Pro		490					495	
Val	Gly	Leu	Gln 500	Asn	Val	Phe	Ser			Leu	Pro	Arg	Met 510	 	ràs,





Glu Tyr Ile Thr Arg Leu Val Phe Asp Pro Lys His Lys Thr Leu Ala 520 525 515 Leu Ile Lys Asp Gly Arg Val Ile Gly Gly Ile Cys Phe Arg Met Phe 535 540 Pro Ser Gln Gly Phe Thr Glu Ile Val Phe Cys Ala Val Thr Ser Asn 555 550 Glu Gln Val Lys Gly Tyr Gly Thr His Leu Met Asn His Leu Lys Glu 570 575 565 Tyr His Ile Lys His Glu Ile Leu Asn Phe Leu Thr Tyr Ala Asp Glu 585 Tyr Ala Ile Gly Tyr Phe Lys Lys Gln Gly Phe Ser Lys Glu Ile Lys 600 595 Ile Pro Lys Thr Lys Tyr Val Gly Tyr Ile Lys Asp Tyr Glu Gly Ala 615 ( 620 Thr Leu Met Gly Cys Glu Leu Asn Pro Gln Ile Pro Tyr Thr Glu Phe 630 635 Ser Val Ile Ile Lys Lys Gln Lys Glu Ile Ile Lys Lys Leu Ile Glu 650 645 Arg Lys Gln Ala Gln Ile Arg Lys Val Tyr Pro Gly Leu Ser Cys Phe 665 670 Lys Asp Gly Val Arg Gln Ile Pro Ile Glu Ser Ile Pro Gly Ile Arg 685 675 680 Glu Thr Gly Trp Lys Pro Ser Gly Lys Glu Lys Ser Lys Glu Pro Lys 700 695 Asp Pro Glu His Val Tyr Ser Thr Leu Lys Asn Ile Leu Gln Gln Val 710 **7**15 Lys Asn His Pro Asn Ala Trp Pro Phe Met Glu Pro Val Lys Arg Thr 735 725 730 Glu Ala Pro Gly Tyr Tyr Glu Val Ile Arg Phe Pro Met Asp Leu Lys 745 Thr Met Ser Glu Arg Leu Arg Asn Arg Tyr Tyr Val Ser Lys Lys Leu 760 Phe Met Ala Asp Leu Gln Arg Val Phe Thr Asn Cys Lys Glu Tyr Asn 775 780 Pro Pro Glu Ser Glu Tyr Tyr Lys Cys Ala Ser Ile Leu Glu Lys Phe 790 795 Phe Phe-Ser Lys Ile Lys Glu Ala Gly Leu Ile Asp Lys

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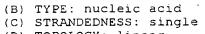
# (2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 50 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

# (2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 2204 base pairs



(D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

					,	
	CCCAGAGCCG				AATCCTGTCG	60
GAGTTGTAGC	CATGCCCTAG	CTGCTCATGT	TTCCCACCTG	GAGAATGTGT	CAGAGGAAGA	120
AATGAACAGA	CTCCTGGGAA	TAGTATTGGA	TGTGGAATAT	CTCTTTACCT	GTGTCCACAA	180
GGAAGAAGAT	GCAGATACCA	AACAAGTTTA	TTTCTATCTA	TTTAAGCTCT	TGAGAAAGTC	240
TATTTTACAA	AGAGGAAAAC	CTGTGGTTGG	AAGGCTCTTT	GGAAAAGAAA	CCCCCATTTG	300
AAAAACCTAG	CATTGAACAG				AGTCACCTGC	360
CAGCAAAAAG	AAAGGCAAAC	CAATAGTTGA	GTTGGCAAAA	ATGTTCCTAA	ACCGCATCAC	420
CTATTGGCAT	CTGGAGGCAC	CATCTCAACG	AGACTGCGAT	CTCCAATGAT	GATATTCTGG	480
ATACAAAGAG	AACTACACAA	GGTGGCTGTG	TTACTGCAAC	GTGCCACAGT	TCTGCGACAG	540
TCTACCTCGG	TACGAAACCA		TGGGAGAACA	TCGTTCGCTC	GGTCTTCACT	600
GTTATGAGGC	GACAACTCCT	GGAACAAGCA	AGACAGGAAA	AAGATAAACT	GCCTCTTGAA	660
AAACGAACTC	TAATCCTCAC	TCATTTCCCA	AAATTTCTGT	CCATGCTAGA	AGAAGAAGTA	-720
TATAGTCAAA	ACTCTCCCAT	CTGGGATCAC	CATTTTCTCT	CAGCCTCTTC	CAGAACCAGC	.780
CAGCTAGGCA	TCCAAACAGT	TATCAATCAC	CTCCTGTGGC	TGGGACAATT	TCATACAATT	840
CAACCTCATC	TTCCCTTGAG	CAGCCAAACG	CAGGGAGCAG	CAGTCCTGCC	TGCAAAGCCT	900
CTTCTGGACT	TGAGGCAAAC	CCAGGAGAAA	AGAGGAAAAT	GACTGATTCT	CATGTTCTGG	960
AGGAGGCCAA	GAAACCCCGA	GTTATGGGGG	ATATTCCGAT	GGAATTAATC	AACGAGGTTA	1020
TGTCTACCAT	CACGGACCCT	GCAGCAATGC	TTGGACCAGA	GACCAATTTT	CTGTCAGCAC	1080
ACTCGGCCAG	GGATGAGGCG	GCAAGGTTGG	AAGAGCGCAG	GGGTGTAATT	GAATTTCACG	1140
TGGTTGGCAA	TTCCCTCAAC	CAGAAACCAA	ACAAGAAGAT	CCTGATGTGG	CTGGTTGGCC	1200
TACAGAACGT	TTTCTCCCAC	CAGCTGCCCC	GAATGCCAAA	AGAATACATC	ACACGGCTCG	1260
TCTTTGACCC	GAAACACAAA	ACCCTTGCTT	TAATTAAAGA	TGGCCGTGTT	ATTGGTGGTA .	1320
TCTGTTTCCG	TATGTTCCCA	TCTCAAGGAT	TCACAGAGAT	TGTCTTCTGT	GCTGTAACCT	1380
CAAATGAGCA	AGTCAAGGGC	TATGGAACAC	ACCTGATGAA	TCATTTGAAA	GAATATCACA	1440
TAAAGCATGA	CATCCTGAAC	TTCCTCACAT	ATGCAGATGA	ATATGCAATT	GGATACTTTA	1500
AGAAACAGGG	TTTCTCCAAA	GAAATTAAAA	TACCTAAAAC	CAAATATGTT	GGCTATATCA	1560
AGGATTATGA	AGGAGCCACT	TTAATGGGAT	GTGAGCTAAA	TCCACGGATC	CCGTACACAG	1620
AATTTTCTGT	CATCATTAAA	AAGCAGAAGG	AGATAATTAA	AAAACTGATT	GAAAGAAAAC	1680
AGGCACAAAT	TCGAAAAGTT	TACCCTGGAC	TTTCATGTTT	TAAAGATGGA	GTTCGACAGA	1740
TTCCTATAGA	AAGCATTCCT	GGAATTAGAG	AGACAGGCTG	GAAACCGAGT	GGAAAAGAGA	1800
AAAGTAAAGA	GCCCAGAGAC	CCTGACCAGC	TTTACAGCAC	GCTCAAGAGC	ATCCTCCAGC	1860
AGGTGAAGAG	CCATCAAAGC	GCTTGGCCCT	TCATGGAACC	TGTGAAGAGA	ACAGAAGCTC	1920
CAGGATATTA	TGAAGTTATA	AGGTCCCCCA	TGGATCTCAA	AACCATGAGT	GAACGCCTCA	1980
AGAATAGGTA	CTACGTGTCT	AAGAAATTAT	TCATGGCAGA	CTTACAGCGA	GTCTTTACCA	2040
ATTGCAAAGA	GTACAACGCC	CCTGAGAGTG	AATACTACAA		ATCCTGGAGA	2100
AATTCTTCTT	CAGTAAAATT	AAGGAAGCTG	GATTAATTGA	CAAGTGATTT	TTTTTCCCCC	2160
TCTGCTTCTT	AGAAACTCAC	CAAGCAGTGT	GCCTAAAGCA	AGGT		2204

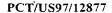
#### (2) INFORMATION FOR SEQ ID NO:11:

# (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2093 base pairs
- (B) TYPE: nucleic acid(C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

GAATTCCGGC	GAAACCACTC	ATGTCTTTGG	GCGAAGCCTT	CTCCGGTCCA	TTTTCACCGT	60
TACCCGCCGG	CAGCTGCTGG	AAAAGTTCCG	AGTGGAGAAG	GACAAATTGG	TGCCCGAGAA	120
GAGGACCCTC	ATCCTCACTC	ACTTCCCCAA	GTAAGGCTCC	TTCTGGCCTA	CCAGGATTTG	180
GCCCCAAGTT	CACATCCTCC	CTGTTGTCCC	CTTTTTTCCA	GGAAGGCTTC	CTGGATTGGT	240
CCCTCCTCTC	CCTCCATGGG	CCTTTTGGGA	TCTGGGCGTC	TACCTGGCAG	ACTTGCCCAT	300
GGCCCAGAAG	CAACTTGCTA	GTACTAGTCT	GGGGATGGCA	GATTCCTGTC	CATGCTGGAG	360
GAGGAGATCT	ATGGGGCAAA	CTCTCCAATC	TGGGAGTCAG	GCTTCACCAT	GCCACCCTCA	420









GAGGGGACAC AGCTGGTTCC CCGGCCAGCT TCAGTCAGTG CAGCGGTTGT TCCCAGCACC CCCATCTTCA GCCCCAGCAT GGGTGGGGGC AGCAACAGCT CCCTGAGTCT GGATTCTGCA GGGGCCGAGC CTATGCCAGG CGAGAAGAGG ACGCTCCCAG AGAACCTGAC CCTGGAGGAT 600 GCCAAGCGGC TCCGTGTGAT GGGTGACATC CCCATGGAGC TGGTCAATGA GGTCATGCTG 660 ACCATCACTG ACCCTGCTGC CATGCTGGGG CCTGAGACGA GCCTGCTTTC GGCCAATGCG 720 GCCCGGGATG AGACAGCCCG CCTGGAGGAG CGCCGCGGCA TCATCGAGTT CCATGTCATC GGCAACTCAC TGACGCCCAA GGCCAACCGG CGGGTGTTGC TGTGGCTCGT GGGGCTGCAG AATGTCTTTT CCCACCAGCT GCCGCGCATG CCTAAGGAGT ATATCGCCCG CCTCGTCTTT 900 GACCCGAAGC ACAAGACTCT GGCCTTGATC AAGGATGGGC GGGTCATCGG TGGCATCTGC 960 TTCCGCATGT TTCCCACCCA GGGCTTCACG GAGATTGTCT TCTGTGCTGT CACCTCGAAT 1020 GAGCAGGTCA AGGGTTATGG GACCCACCTG ATGAACCACC TGAAGGAGTA TCACATCAAG 1080 CACAACATTC TCTACTTCCT CACCTACGCC GACGAGTACG CCATCGGCTA CTTCAAAAAG 1140 CAGGGTTTCT CCAAGGACAT CAAGGTGCCC AAGAGCCGCT ACCTGGGCTA CATCAAGGAC 1200 TACGAGGGAG CGACGCTGAT GGAGTGTGAG CTGAATCCCC GCATCCCCTA CACGGAGCTG 1260 TCCCACATCA TCAAGAAGCA GAAAGAGATC ATCAAGAAGC TGATTGAGCG CAAACAGGCC 1320 1380 CAGATCCGCA AGGTCTACCC GGGGCTCAGC TGCTTCAAGG AGGGCGTGAG GCAGATCCCT GTGGAGAGCG TTCCTGGCAT TCGAGAGACA GGCTGGAAGC CATTGGGGAA GGAGAAGGGG 1440 AAGGAGCTGA AGGACCCCGA CCAGCTCTAC ACAACCCTCA AAAACCTGCT GGCCCAAATC AAGTCTCACC CCAGTGCCTG GCCCTTCATG GAGCCTGTGA AGAAGTCGGA GGCCCCTGAC 1560 TACTACGAGG TCATCCGCTT CCCCATTGAC CTGAAGACCA TGACTGAGCG GCTGCGAAGC 1620 CGCTACTACG TGACCCGGAA GCTCTTTGTG GCCGACCTGC AGCGGGTCAT CGCCAACTGT 1680 CGCGAGTACA ACCCCCGGA CAGCGAGTAC TGCCGCTGTG CCAGCGCCCT GGAGAAGTTC TTCTACTTCA AGCTCAAGGA GGGAGGCCTC ATTGACAAGT AGGCCCATCT TTGGGCCGCA 1800 CCCACGGACC CGACTCAGCT TGAGACACTC CAGCCAAGGG TCCTCCGGAC CCGATCCTGC 1920 AGCTCTTTCT GGACCTTCAG GCACCCCCAA GCGTGCAGCT CTGTCCCAGC CTTCACTGTG 1980 TGTGAGAGGT CTCCTGGGTT GGGGCCCAGC CCCTCTAGAG TAGCTGGTGG CCAGGGATGA 2040 ACCTTGCCCA GCCGTGGTGG CCCCCAGGCC TGGTCCCCAA GAGCCCGGAA TTC

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#### (2) INFORMATION FOR SEQ ID NO:12:

#### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 9046 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

CCTTGTTTGT	GTGCTAGGCT	GGGGGGAGA	GAGGGCGAGA	GAGAGCGGGC	GAGAGTGGGC	60
AAGCAGGACG	CCGGGCTGAG	TGCTAACTGC	GGGACGCAGA	GAGTGCGGAG	GGGAGTCGGG	120
TCGGAGAGAG	GCGGCAGGGG	CCAGAACAGT	GGCAGGGGGC	CCGGGGCGCA	CGGGCTGAGG	180
CGACCCCCAG	CCCCTCCCG	TCCGCACACA	CCCCCACCGC	GGTCCAGCAG	CCGGGCCGGC	240
GTCGACGCTA	GGGGGGACCA	TTACATAACC	CGCGCCCCGG	CCGTCTTCTC	CCGCCGCCGC	300
GGCGCCCGAA	CTGAGCCCGG	GGCGGGCGCT	CCAGCACTGG	CCGCCGGCGT	GGGGCGŢAGC	360
AGCGGCCGTA	TTATTATTTC	GCGGAAAGGA	AGGCGAAGGA	GGGGAGCGCC	GGCGCGAGGA	420
GGGGCCGCCT	GCGCCGCCG	CCGGAGCGGG	GCCTCCTCGG	TGGGCTCCGC	GTCGGCGCGG	480
GCGTGCGGGC	GGCGCTGCTC	GGCCCGGCCC	CCTCGGCCCT	CTGGTCCGGC	CAGCTCCGCT	540
CCCGGCGTCC	TTGCCGCGCC	TCCGCCGGCC	GCCGCGCGAT	GTGAGGCGGC	GGCGCCAGCC	600
TGGCTCTCGG	CTCGGGCGAG	TTCTCTGCGG	CCATTAGGGG	CCGGTGCGGC	GGCGGCGG	660
AGCGCGGCGG	CAGGAGGAGG	GTTCGGAGGG	TGGGGGCGCA	GGCCCGGGAG	GGGGCACCGG	720
GAGGAGGTGA	GTGTCTCTTG	TCGCCTCCTC	CTCTCCCCCC	TTTTCGCCCC	CGCCTCCTTG	780
TGGCGATGAG	AAGGAGGAGG	ACAGCGCCGA	GGAGGAAGAG	GTTGATGGCG	GCGGCGGAGC	840
TCCGAGAGAC	. CTCGGCTGGG	'CAGGGGCCGG	CCGTGGCGGG	CCGGGGACTG	CGCCTCTAGA	900
GCCGCGAGTT	CTCGGGAATT	CGCCGCAGCG	GACCGGCCTC	GGCGAATTTG	TGCTCTTGTG	960
CCCTCCTCCG	GGCTTGGGCC	AGGCCGGCCC	CTCGCACTTG	CCCTTACCTT	TTCTATCGAG	1020
TCCGCATCCC	TCTCCAGCCA	CTGCGACCCG	GCGAAGAGAA	AAAGGAACTT	CCCCCACCCC	1080
CTCGGGTGCC	GTCGGAGCCC	CCCAGCCCAC	CCCTGGGTGC	GGCGCGGGGA	CCCCGGGCCG	1140
AAGAAGAGAT	TTCCTGAGGA	TTCTGGTTTT	CCTCGCTTGT	ATCTCCGAAA	GAATTAAAAA	1200
TGGCCGAGAA	TGTGGTGGAA	CCGGGGCCGC	CTTCAGCCAA	GCGGCCTAAA	CTCTCATCTC	1260
CGGCCCTCTC	GGCGTCCGCC	AGCGATGGCA	CAGATTTTGG	CTCTCTATTT	GACTTGGAGC	1320
ACGACTTACC	AGATGAATTA	ATCAACTCTA	CAGAATTGGG	ACTAACCAAT	GGTGGTGATA	1380





TGTCAGAATT GCTGCGATCT GGTAGTTCCC CTAACCTCAA TATGGGAGTT GGTGGCCCAG GTCAAGTCAT GGCCAGCCAG GCCCAACAGA GCAGTCCTGG ATTAGGTTTG ATAAATAGCA TGGTCAAAAG CCCAATGACA CAGGCAGGCT TGACTTCTCC CAACATGGGG ATGGGCACTA GTGGACCAAA TCAGGGTCCT ACGCAGTCAA CAGGTATGAT GAACAGTCCA GTAAATCAGC 1680 CTGCCATGGG AATGAACACA GGGACGAATG CGGGCATGAA TCCTGGAATG TTGGCTGCAG . 1740 GCAATGGACA AGGGATAATG CCTAATCAAG TCATGAACGG TTCAATTGGA GCAGGCCGAG 1800 GGCGACAGGA TATGCAGTAC CCAAACCCAG GCATGGGAAG TGCTGGCAAC TTACTGACTG AGCCTCTTCA GCAGGGCTCT CCCCAGATGG GAGGACAAAC AGGATTGAGA GGCCCCCAGC CTCTTAAGAT GGGAATGATG AACAACCCCA ATCCTTATGG TTCACCATAT ACTCAGAATC CTGGACAGCA GATTGGAGCC AGTGGCCTTG GTCTCCAGAT TCAGACAAAA ACTGTACTAT 2040 CAAATAACTT ATCTCCATTT GCTATGGACA AAAAGGCAGT TCCTGGTGGA GGAATGCCCA 2100 ACATGGGTCA ACAGCCAGCC CCGCAGGTCC AGCAGCCAGG TCTGGTGACT CCAGTTGCCC 2160 AAGGGATGGG TTCTGGAGCA CATACAGCTG ATCCAGAGAA GCGCAAGCTC ATCCAGCAGC 2220 AGCTTGTTCT CCTTTTGCAT GCTCACAAGT GCCAGCGCCG GGAACAGGCC AATGGGGAAG 2280 TGAGGCAGTG CAACCTTCCC CACTGTCGCA CAATGAAGAA TGTCCTAAAC CACATGACAC 2340 ACTGCCAGTC AGGCAAGTCT TGCCAAGTGG CACACTGTGC ATCTTCTCGA CAAATCATTT CACACTGGAA GAATTGTACA AGACATGATT GTCCTGTGTG TCTCCCCCTC AAAAATGCTG GTGATAAGAG AAATCAACAG CCAATTTTGA CTGGAGCACC CGTTGGACTT GGAAATCCTA GCTCTCTAGG GGTGGGTCAA CAGTCTGCCC CCAACCTAAG CACTGTTAGT CAGATTGATC CCAGCTCCAT AGAAAGAGCC TATGCAGCTC TTGGACTACC CTATCAAGTA AATCAGATGC CGACACACC CCAGGTGCAA GCAAAGAACC AGCAGAATCA GCAGCCTGGG CAGTCTCCCC AAGGCATGCG GCCCATGAGC AACATGAGTG CTAGTCCTAT GGGAGTAAAT GGAGGTGTAG GAGTTCAAAC GCCGAGTCTT CTTTCTGACT CAATGTTGCA TTCAGCCATA AATTCTCAAA 2820 ACCCAATGAT GAGTGAAAAT GCCAGTGTGC CCTCCCTGGG TCCTATGCCA ACAGCAGCTC 2880 AACCATCCAC TACTGGAATT CGGAAACAGT GGCACGAAGA TATTACTCAG GATCTTCGAA 2940 ATCATCTTGT TCACAAACTC GTCCAAGCCA TATTTCCTAC GCCGGATCCT GCTGCTTTAA AAGACAGACG GATGGAAAAC CTAGTTGCAT ATGCTCGGAA AGTTGAAGGG GACATGTATG AATCTGCAAA CAATCGAGCG GAATACTACC ACCTTCTAGC TGAGAAAATC TATAAGATCC 3120 AGAAAGAACT AGAAGAAAAA CGAAGGACCA GACTACAGAA GCAGAACATG CTACCAAATG 3180 CTGCAGGCAT GGTTCCAGTT TCCATGAATC CAGGGCCTAA CATGGGACAG CCGCAACCAG GAATGACTTC TAATGGCCCT CTACCTGACC CAAGTATGAT CCGTGGCAGT GTGCCAAACC 3240 AGATGATGCC TCGAATAACT CCACAATCTG GTTTGAATCA ATTTGGCCAG ATGAGCATGG 3360 CCCAGCCCC TATTGTACCC CGGCAAACCC CTCCTCTCA GCACCATGGA CAGTTGGCTC AACCTGGAGC TCTCAACCCG CCTATGGGCT ATGGGCCTCG TATGCAACAG CCTTCCAACC 3420 3480 AGGGCCAGTT CCTTCCTCAG ACTCAGTTCC CATCACAGGG AATGAATGTA ACAAATATCC CTTTGGCTCC GTCCAGCGGT CAAGCTCCAG TGTCTCAAGC ACAAATGTCT AGTTCTTCCT GCCCGGTGAA CTCTCCTATA ATGCCTCCAG GGTCTCAGGG GAGCCACATT CACTGTCCCC AGCTTCCTCA ACCAGCTCTT CATCAGAATT CACCCTCGCC TGTACCTAGT CGTACCCCCA CCCCTCACCA TACTCCCCCA AGCATAGGGG CTCAGCAGCC ACCAGCAACA ACAATTCCAG 3780 CCCCTGTTCC TACACCACCA GCCATGCCAC CTGGGCCACA GTCCCAGGCT CTACATCCCC CTCCAAGGCA GACACCTACA CCACCAACAA CACAACTTCC CCAACAAGTG CAGCCTTCAC 3900 TTCCTGCTGC ACCTTCTGCT GACCAGCCC AGCAGCAGCC TCGCTCACAG CAGAGCACAG CAGCGTCTGT TCCTACCCCA AACGCACCGC TGCTTCCTCC GCAGCCTGCA ACTCCACTTT CCCAGCCAGC TGTAAGCATT GAAGGACAGG TATCAAATCC TCCATCTACT AGTAGCACAG 3960 4020 4080 AAGTGAATTC TCAGGCCATT GCTGAGAAGC AGCCTTCCCA GGAAGTGAAG ATGGAGGCCA 4140 AAATGGAAGT GGATCAACCA GAACCAGCAG ATACGCAGCC GGAGGATATT TCAGAGTCTA 4200 AAGTGGAAGA CTGTAAAATG GAATCTACCG AAACAGAAGA GAGAAGCACT GAGTTAAAAA 4260 CTGAAATAAA AGAGGAGGAA GACCAGCCAA GTACTTCAGC TACCCAGTCA TCTCCGGCTC 4320 CAGGACAGTC AAAGAAAAG ATTTTCAAAC CAGAAGAACT ACGACAGGCA CTGATGCCAA 4380 CATTGGAGGC ACTTTACCGT CAGGATCCAG AATCCCTTCC CTTTCGTCAA CCTGTGGACC 4440 CTCAGCTTTT AGGAATCCCT GATTACTTTG ATATTGTGAA GAGCCCCATG GATCTTTCTA 4500 CCATTAAGAG GAAGTTAGAC ACTGGACAGT ATCAGGAGCC CTGGCAGTAT GTCGATGATA 4560 TTTGGCTTAT GTTCAATAAT GCCTGGTTAT ATAACCGGAA AACATCACGG GTATACAAAT 4620 ACTGCTCCAA GCTCTCTGAG GTCTTTGAAC AAGAAATTGA CCCAGTGATG CAAAGCCTTG 4680 GATACTGTTG TGGCAGAAG TTGGAGTTCT CTCCACAGAC ACTGTGTTGC TACGGCAAAC AGTTGTGCAC AATACCTCGT GATGCCACTT ATTACAGTTA CCAGAACAGG TATCATTTCT 4800 GTGAGAAGTG TTTCAATGAG ATCCAAGGGG AGAGCGTTTC TTTGGGGGAT GACCCTTCCC 4860 AGCCTCAAAC TACAATAAAT AAAGAACAAT TTTCCAAGAG AAAAAATGAC ACACTGGATC 4980 CTGAACTGTT TGTTGAATGT ACAGAGTGCG GAAGAAAGAT GCATCAGATC TGTGTCCTTC ACCATGAGAT CATCTGGCCT GCTGGATTCG TCTGTGATGG CTGTTTAAAG AAAAGTGCAC 5040 GAACTAGGAA AGAAAATAAG TTTTCTGCTA AAAGGTTGCC ATCTACCAGA CTTGGCACCT 5100 TTCTAGAGAA TCGTGTGAAT GACTTTCTGA GGCGACAGAA TCACCCTGAG TCAGGAGAGG 5160

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TTAATCAGCT TCAGACAAGT CTTGGCATGG TACAAGATGC AGCTTCTAAA CATAAACAGC



	AGTAGTTCAT					5220
CAAGGTTTGT	GGACAGTGGA	GAGATGGCAG	AATCCTTTCC	ATACCGAACC	AAAGCCCTCT	5280
TTGCCTTTGA	AGAAATTGAT	GGTGTTGACC	TGTGCTTCTT	TGGCATGCAT	GTTCAAGAGT	5340
	CTGCCCTCCA			CATATCTTAC	CTCGATAGTG	5400
	CCGTCCTAAA					5460
	TGTCAAGAAA					5520
	TGATTATATC					5580
						5640
	GGAATGGTAC			TGTATCAGAG		
	GGATATTTTT					5700
	CGAGGGTGAT					5760
	AGAAGAGAGA					5820
CCAAGGGAGA	CAGCAAAAAT	GCTAAAAAGA	AGAATAATAA	GAAAACCAGC	AAAAATAAGA	5880
GCAGCCTGAG	TAGGGGCAAC	AAGAAGAAAC	CCGGGATGCC	CAATGTATCT	AACGACCTCT	5940
CACAGAAACT	ATATGCCACC	ATGGAGAAGC	ATAAAGAGGT	CTTCTTTGTG	ATCCGCCTCA	6000
TTGCTGGCCC	TGCTGCCAAC			TCCTGATCCT		6060
	GGATGGTCGG			AAGGGACAAG		6120
TCTCTTCACT				GCTGGTGGAG		6180
	CCGCTTTGTC					6240
	TGTCTGTGAG					6300
	AATGGAGAAA					6360
	CCAGAGCCCA					6420
	TGCTTGCCAG				TGCCAGAAGA	6480
	TGTGCAGCAT					6540
TCTGCAAGCA	GCTCATTGCC	CTCTGCTGCT	ACCATGCCAA	GCACTGCCAG	GAGAACAAAT	6600
GCCCGGTGCC	GTTCTGCCTA	AACATCAAGC	AGAAGCTCCG	GCAGCAACAG	CTGCAGCACC	6660
	GGCCCAAATG					6720
	ACAGGGCCTC					6780
	CACCCGCAG			GCCTCAGCCT		6840
	ACCCTACTTG					6900
	GGTGACCCCT					6960
						7020
	AGCAGTGGAA				•	•
	CGTGCAAATT			CCAGATGCCC		7080
	CATGGGTATG					7140
	GGGACCGACA					7200
	GCAACTACAG					7260
ATGGTCAACC.	TTTGAACATG	GCTCCACAAC	CAGGATTGGG	CCAGGTAGGT	ATCAGCCCAC	7320
TCAAACCAGG	CACTGTGTCT	CAACAAGCCT	TACAAAACCT	TTTGCGGACT	CTCAGGTCTC	7380
CCAGCTCTCC	CCTGCAGCAG	CAACAGGTGC	TTAGTATCCT	TCACGCCAAC	CCCCAGCTGT	7440
TGGCTGCATT	CATCAAGCAG	CGGGCTGCCA	AGTATGCCAA	CTCTAATCCA	CAACCCATCC	7500
	TGGCATGCCC					7560
	CCACTCCAAT					7620
	CCTGCCCCAG			CCAGCCACCC		7680
	GGCTCAGCAG					7740
	ACGACAGCAA					7800
	AATGGCCAAC					7860
	GCAGCGGATG					7920
	GCTTCCCCAG					7980
	CCTTCAGCAA					8040
	GCTCCCAAAT					8100
					CCACAGTCCC	8160
	CTCCAGTCCT					8220
CCCCACAGAC	AAGTTCCCCA	CATCCTGGAC	TGGTAGCTGC	CCAGGCCAAC	CCCATGGAAC	8280
AAGGGCATTT	TGCCAGCCCG	GACCAGAATT	CAATGCTTTC	TCAGCTTGCT	AGCAATCCAG	8340
	CCTCCATGGT					8400
	AAACCTCTCA					8460
					ATTTTTTGA	8520
	GCCTAAAAGA					8580
	AAGCAAACAT					8640
	GGGCTGGTTA					
						8700
	GGAGGCTGAG					8760
	ATTATTTTTT					8820
	TTATTATTTA					8880
TTATGGAAGA	GTTAAAACAT	TTCTAAACCA	GAGGACAAAA	GGGGTTAATG	TTACTTTGAA	8940



ATTACATTCT ATATATAT AAATATAT AAATATAT TAAAATACCA GTTTTTTTTC 9000 TCTGGGTGCA AAGATGTTCA TTCTTTTAAA AAATGTTTAA AAAAAA 9046

# (2) INFORMATION FOR SEQ ID NO:13:

# (i) SEQUENCE CHARACTERISTICS:

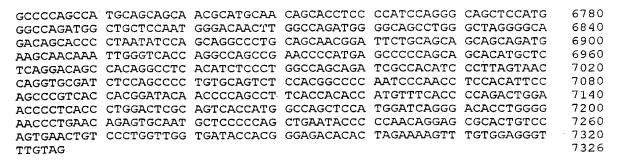
- (A) LENGTH: 7326 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

ATGGCCGAGA	ACTTGCTGGA	CGGACCGCCC	AACCCCAAAC	GAGCCAAACT	CAGCTCGCCC	60
GGCTTCTCCG	CGAATGACAA	CACAGATTTT	GGATCATTGT	TTGACTTGGA	AAATGACCTT	120
		TGGAGAATTA				180
		ACAACTGTCA				240
		TGTGAGTGCC				300
		CAGTACAAAC				360
		ATCAACACCC				420
GGGCCCACTC	CCCCTGCCTC	CCAAGCACTG				480
	GTCCTGCCAC			TCTGCATGAA		540
		TCTCAATAGT				600
		CATGAATGGA				660
		TCCAGCCATG				720
		ACAAATGGCT				780
		AATGACTGGT				840
		GGGAGCCACT			CAGCAAACAG	900
		TGCTTTTCCT				960
		GCAAACATCA				1020
		TGAAAAACGC				1080
		GAGACGAGAG				1140
CTCCCACACT	GTCGAACCAT	GAAAAACGTT	TTGAATCACA	TGACACATTG	TCAGGCTCCC	1200
	AAGTTGCCCA			TCATCTCTCA		1260
TGCACACGAC	ATGACTGTCC	TGTTTGCCTC	CCTTTGAAAA	ATGCCAGTGA	CAAGCGAAAC	1320
CAACAAACCA	TCCTGGGATC	TCCAGCTAGT	GGAATTCAAA	ACACAATTGG	TTCTGTTGGT	1380
GCAGGGCAAC	AGAATGCCAC	TTCCTTAAGT	AACCCAAATC	CCATAGACCC	CAGTTCCATG	1440
		AGGACTCCCC				1500
CCTCAGGTTC	CTGGCCAGCA	ACCAGCACAG	CCTCCAGCCC	ACCAGCAGAT	GAGGACTCTC	1560
AATGCCCTAG	GAAACAACCC	CATGAGTGTC	CCAGCAGGAG	GAATAACAAC	AGATCAACAG	1620
CCACCAAACT	TGATTTCAGA	ATCAGCTCTT	CCAACTTCCT	TGGGGGCTAC	CAATCCACTG	1680
ATGAATGATG	GTTCAAACTC	TGGTAACATT	GGAAGCCTCA	GCACGATACC	TACAGCAGCG	1740
CCTCCTTCCA	GCACTGGTGT	TCGAAAAGGC	TGGCATGAAC	ATGTGACTCA	GGACCTACGG	1800
AGTCATCTAG	TCCATAAACT	CGTTCAAGCC	ATCTTCCCAA	CTCCAGACCC	TGCAGCTCTG	1860
AAAGATCGCC	GCATGGAGAA	CCTGGTTGCC	TATGCTAAGA	AAGTGGAGGG	AGACATGTAT	1920
GAGTCTGCTA	ATAGCAGGGA	TGAATACTAT	CATTTATTAG	CAGAGAAAAT	CTATAAAATA	1980
CAAAAAGAAC	TAGAAGAAAA	GCGGAGGACA	CGTTTACATA	AGCAAGGCAT	CCTGGGTAAC	2040
CAGCCAGCTT	TACCAGCTTC	TGGGGCTCAG	CCCCCTGTGA	TTCCACCAGC	CCAGTCTGTA	2100
AGACCTCCAA	ATGGGCCCCT	GCCTTTGCCA	GTGAATCGCA	TGCAGGTTTC	TCAAGGGATG	2160
AATTCATTTA	ACCCAATGTC	CCTGGGAAAC	GTCCAGTTGC	CACAGGCACC	CATGGGACCT	2220
CGTGCAGCCT	CCCCTATGAA	CCACTCTGTG	CAGATGAACA	GCATGGCCTC	AGTTCCGGGT	2280
ATGGCCATTT	CTCCTTCACG	GATGCCTCAG	CCTCCAAATA	TGATGGGCAC	TCATGCCAAC	2340
AACATTATGG	CCCAGGCACC	TACTCAGAAC	CAGTTTCTGC	CACAGAACCA	GTTTCCATCA	2400
TCCAGTGGGG	CAATGAGTGT	GAACAGTGTG	GGCATGGGGC	AACCAGCAGC	CCAGGCAGGT	2460
GTTTCACAGG	GTCAGGAACC	TGGAGCTGCT	CTCCCTAACC	CTCTGAACAT	GCTGGCACCC	2520
		CCCACCAGTG			GACTCCACCT	25.80
CCTGCTTCCA	CAGCTGCTGG	CATGCCCTCT	CTCCAACATC	CAACGGCACC	AGGAATGACC	2640
CCTCCTCAGC	CAGCAGCTCC	CACTCAGCCA	TCTACTCCTG	TGTCATCTGG	GCAGACTCCT	27.00
		GCCCAGCGCT			TACAGTCCAG	2760
		GACTCCACAG				2820
		'GCAGCAACCA				2880
CCGCTTTCTC	AGGCAGCAGC	CAGCATTGAT	AATAGAGTCC	CTACTCCCTC	CACTGTGACC	2940



AGTGCTGAAA	CCAGTTCCCA	GCAGCCAGGA	CCCGATGTGC	CCATGCTGGA	AATGAAGACA	. 3000
GAGGTGCAGA	CAGATGATGC	TGAGCCTGAA	CCTACTGAAT	CCAAGGGGGA	ACCTCGGTCT.	3060
CACATGATGG	AAGAGGATTT	ACAAGGTTCT	TCCCAAGTAA	AAGAAGAGAC	AGATACGACA	3120
GAGAIGAIGG	CAGAGCCAAT	CCANCTACAA	CDDDDCGDDC	СТСДДСТДДД	AGTGGAAGCT	3180
GAGCAGAAGI	AAGAGAACAG	MMCCD ACCAC	DATE COUNTY	AATCAACATC	TCCTTCCCNG	3240
AAAGAGGAAG	AAGAGAACAG	TTCGAACGAC	ACAGCCTCAC	AAICAACAIC	P. C.	
CCACGCAAAA	AAATCTTTAA	ACCCGAGGAG	CTACGCCAGG	CACTTATGCC	AACTCTAGAA	3300
GCACTCTATC	GACAGGACCC	AGAGTCTTTG	CCTTTTCGTC	AGCCTGTAGA	TCCTCAGCTC	3360
CTAGGAATCC	CAGATTATTT	TGATATAGTG	AAGAATCCTA	TGGACCTTTC	TACCATCAAA	3420
CGAAAGCTGG	ACACAGGGCA	ATATCAAGAA	CCCTGGCAGT	ATGTGGATGA	TGTCAGGCTT	3480
AUCUUCA ACA	ATGCGTGGCT	ATATAATCGT	AAAACGTCCC	GTGTATATAA	ATTTTGCAGT	3540
AIGIICAACA	AGGTCTTTGA	ACAACAAATT	CACCCTGTCA	TGCAGTCTCT	TGGATATTGC	3600
AAACTIGCAG	AGTATGAGTT	CTCCCCACAC	A CERTECT CE	CTTACCCAAA	CCACCTCTCT	3660
TGTGGACGAA	AGTATGAGTT	CICCCCACAG	ACTITIGIGET	CCMAMCAMM	CTCTCCCAAC	3720
ACAATTCCTC	GTGATGCAGC	CTACTACAGC	TATCAGAATA	DOGRACATTI	CIGIGGGAAG	3780
TGTTTCACAG	AGATCCAGGG	CGAGAATGTG	ACCCTGGGTG	ACGACCCTIC	CCAACCICAG	
ACGACAATTT	CCAAGGATCA	ATTTGAAAAG	AAGAAAAA'I'G	ATACCTTAGA	TCCTGAACCT	3840
TTTGTTGACT	GCAAAGAGTG	TGGCCGGAAG	ATGCATCAGA	TTTGTGTTCT	ACACTATGAC	3900
ATCATTTGGC	CTTCAGGTTT	TGTGTGTGAC	AACTGTTTGA	AGAAAACTGG	CAGACCTCGG	3960
AAAGAAAACA	AATTCAGTGC	TAAGAGGCTG	CAGACCACAC	GATTGGGAAA	CCACTTAGAA	4020
GACAGAGTGA	ATAAGTTTTT	GCGGCGCCAG	AATCACCCTG	AAGCTGGGGA	GGTTTTTGTC	4080
DCDCTCCTCG	CCAGCTCAGA	CAAGACTGTG	GAGGTCAAGC	CGGGAATGAA	GTCAAGGTTT	4140
AGAGIGGIGG	GAGAGATGTC	CCDATCTTTC	CCATATCGTA	CCAAAGCACT	CTTTGCTTTT	4200
GTGGATICIG	ATGGAGTCGA	MCMCMCCMMM	TTTTCCCDTCC	ATGTGCAAGA	TACGGCTCTG	4260
GAGGAGATCG	ATGGAGTCGA	TGTGTGCTTT	TITGGGATGC	RIGIGCAAGA	TACGGCICIG	4320
ATTGCCCCCC	ACCAAATACA	AGGCTGTGTA	TACATATCTT	ATCTGGACAG	TATTCATTIC	
TTCCGGCCCC	GCTGCCTCCG	GACAGCTGTT	TACCATGAGA	TCCTCATCGG	ATATCTCGAG	4380
TATGTGAAGA	AATTGGTGTA	TGTGACAGCA	CATATTTGGG	CCTGTCCCCC	AAGTGAAGGA	4440
GATGACTATA	TCTTTCATTG	CCACCCCCT	GACCAGAAAA	TCCCCAAACC	AAAACGACTA	4500
CAGGAGTGGT	ACAAGAAGAT	GCTGGACAAG	GCGTTTGCAG	AGAGGATCAT	TAACGACTAT	4560
AAGGACATCT	TCAAACAAGC	GAACGAAGAC	AGGCTCACGA	GTGCCAAGGA	GTTGCCCTAT	4620
TTTCAACCAC	ATTTCTGGCC	ΤΑΑΤΩΤΩΤΤΩ	GAAGAAAGCA	TTAAGGAACT	AGAACAAGAA	4680
CANCAACAA	GGAAAAAAGA	ACACACTACT	GCAGCGAGTG	AGACTCCTGA	GGGCAGTCAG	4740
GAAGAAGAAA	AAAATGCGAA	CDADADIACI	DACABOOMOTO	CCAACAAAAA	CANANGCAGC	4800
GGTGACAGCA	AAAATGCGAA	GAAAAAGAAC	AACAAGAAGA	mmmcca a cca	COMOTOCO CO	4860
ATTAGCCGCG	CCAACAAGAA	GAAGCCCAGC	AIGCCCAAIG	TTTCCAACGA	TCTGTCGCAG	4920
AAGCTGTATG	CCACCATGGA	GAAGCACAAG	GAGGTATTCT	TTGTGATTCA	TCTGCATGCT	
GGGCCTGTTA	TCAGCACTCA	GCCCCCCATC	GTGGACCCTG	ATCCTCTGCT	TAGCTGTGAC	4980
CTCATGGATG	GGCGAGATGC	CTTCCTCACC	CTGGCCAGAG	ACAAGCACTG	GGAATTCTCT	5040
TCCTTACGCC	GCTCCAAATG	GTCCACTCTG	TGCATGCTGG	TGGAGCTGCA	CACACAGGGC	5100
CAGGACCGCT	TTGTTTATAC	CTGCAATGAG	TGCAAACACC	ATGTGGAAAC	ACGCTGGCAC	5160
TGCACTGTGT	GTGAGGACTA	TGACCTTTGT	ATCAATTGCT	ACAACACAAA	GAGCCACACC	5220
CATAAGATGG	TGAAGTGGGG	GCTAGGCCTA	GATGATGAGG	GCAGCAGTCA	GGGTGAGCCA	5280
CAIAAGAIGG	GCCCCAGGA	ATCCCGGCGT	CTCAGCATCC	AGCGCTGCAT	CCAGTCCCTG	5340
	GCCAGTGTCG	CARECCOROC	TECTENCTEC	CGTCTTGCCA	CAACATGAAG	5400
GTGCATGCCT	GCCAGIGICG	CAAIGCCAAC	CCCARCACTO	AMECA CCAME	CCCACTCTCC	5460
CGAGTCGTGC	AGCACACCAA	GGGCTGCAAG	CGCAAGACIA	AIGGAGGAIG	TRANSCOCK.	
AAGCAGCTCA	TTGCTCTTTG	CTGCTACCAC	GCCAAACACT	GCCAAGAAAA	TAAATGCCCT	5520
GTGCCCTTCT	GCCTCAACAT	CAAACATAAC	GTCCGCCAGC	AGCAGATCCA	GCACTGCCTG	5580
CAGCAGGCTC	AGCTCATGCG	CCGGCGAATG	GCAACCATGA	ACACCCGCAA	TGTGCCTCAG	5640
CAGAGTTTGC	CTTCTCCTAC	CTCAGCACCA	CCCGGGACTC	CTACACAGCA	GCCCAGCACA	5700
CCCCAAACAC	CACAGCCCCC	AGCCCAGCCT	CAGCCTTCAC	CTGTTAACAT	GTCACCAGCA	5760
GGCTTCCCTA	ATGTAGCCCG	GACTCAGCCC	CCAACAATAG	TGTCTGCTGG	GAAGCCTACC	5820
AACCAGGTGC	CAGCTCCCCC	ACCCCCTGCC	CAGCCCCCAC	CTGCAGCAGT	AGAAGCAGCC	5880
CCCCNNNTTC	AACGTGAGGC	CCAGCAGCAG	CAGCACCTAT	ACCGAGCAAA	CATCAACAAT	5940
CCCAMCCCC	CAGGACGTGA	CGGTNTGGGG	ACCCCAGGAA	GCCDADTGAC	TCCTGTGGGC	6000
GGCATGCCCC	CCCGTCCCAA	CCDDCMCACM	CCCCCTCTCA	TGTCTAGTAT	GCCACCTGGG	6060
CTGAATGTGC	CCCGTCCCAA	CCAAGICAGI	GGGCCIGICA	CACCCAMCCC	CACCCCCCC	6120
CAGTGGCAGC	AGGCACCCAT	CCCTCAGCAG	CAGCCGATGC	CAGGCATGCC	CAGGCCIGIA	
ATGTCCATGC	AGGCCCAGGC	AGCAGTGGCT	GGGCCACGGA	TGCCCAATGT	GCAGCCAAAC	6180
AGGAGCATCT	CGCCAAGTGC	CCTGCAAGAC	CTGCTACGGA	CCCTAAAGTC	ACCCAGCTCT	6240
CCTCAGCAGC	AGCAGCAGGT	GCTGAACATC	CTTAAATCAA	ACCCACAGCT	AATGGCAGCT	6300
TTCATCAAAC	AGCGCACAGC	CAAGTATGTG	GCCAATCAGC	CTGGCATGCA	GCCCCAGCCC	6360
GGACTTCAAT	CCCAGCCTGG	TATGCAGCCC	CAGCCTGGCA	TGCACCAGCA	GCCTAGTTTG	6420
CAAAACCTGA	ACGCAATGCA	AGCTGGTGTG	·CCACGGCCTG	GTGTGCCTCC	ACCACAACCA	6480
GCAATGGGAG	GCCTGAATCC	CCAGGGACAA	GCTCTGAACA	TCATGAACCC	AGGAÇACAAC	6540
CCCANCATGA	CAAACATGAA	TCCACAGTAC	CGAGAAATGG	TGAGGAGACA	GCTGCTACAG	6600
COCMACAIGA	AGCAGCAGCA	DCDCCDCCAC	CAGCAGCAGC	DACABCABAA	TAGTGCCAGC	6660
TACCAGCAGC	GCATGGCGGG	ACAGCAGCAG	TTTCCAGCAGC		TCCACCAGO	6720
TTGGCCGGGG	- GCATGGCGGG	ACACAGCCAG	DDAJDAJII	CACAAGGACC	1 GGAGG1 1A1	0/20



#### (2) INFORMATION FOR SEQ ID NO:14:

#### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2499 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

TCACTTGTCA ATTAATCCAG CTTCCTTAAT TTTACTGAAG AAGAATTTCT CCAGGATATT GGCACATTTG TAGTATTCAC TCTCAGGGGC GTTGTACTCT TTGCAATTGG TAAAGACTCG 120 CTGTAAGTCT GCCATGAATA ATTTCTTAGA CACGTAGTAC CTATTCTTGA GGCGTTCACT CATGGTTTTG AGATCCATGG GGGACCTTAT AACTTCATAA TATCCTGGAG CTTCTGTTCT 240 CTTCACAGGT TCCATGAAGG GCCAAGCGCT TTGATGGCTC TTCACCTGCT GGAGGATGCT 300 CTTGAGCGTG CTGTAAAGCT GGTCAGGGTC TCTGGGCTCT TTACTTTTCT CTTTTCCACT 360 420 CGGTTTCCAG CCTGTCTCTC TAATTCCAGG AATGCTTTCT ATAGGAATCT GTCGAACTCC
ATCTTTAAAA CATGAAAGTC CAGGGTAAAC TTTTCGAATT TGTGCCTGTT TTCTTTCAAT
CAGTTTTTTA ATTATCTCCT TCTGCTTTTT AATGATGACA GAAAATTCTG TGTACGGGAT
CCGTGGATTT AGCTCACATC CCATTAAAGT GGCTCCTTCA TAATCCTTGA TATAGCCAAC 540 600 ATATTTGGTT TTAGGTATTT TAATTTCTTT GGAGAAACCC TGTTTCTTAA AGTATCCAAT TGCATATCA TCTGCATATG TGAGGAAGTT CAGGATGTCA TGCTTTATGT GATATTCTTT 660 720 CAAATGATTC ATCAGGTGTG TTCCATAGCC CTTGACTTGC TCATTTGAGG TTACAGCACA GAAGACAATC TCTGTGAATC CTTGAGATGG GAACATACGG AAACAGATAC CACCAATAAC 840 ACGGCCATCT TTAATTAAAG CAAGGGTTTT GTGTTTCGGG TCAAAGACGA GCCGTGTGAT 900 GTATTCTTTT GGCATTCGGG GCAGCTGGTG GGAGAAACG TTCTGTAGGC CAACCAGCCA
CATCAGGATC TTCTTGTTTG GTTTCTGGTT GAGGGAATTG CCAACCACGT GAAATTCAAT 960 1020 TACACCCCTG CGCTCTTCCA ACCTTGCCGC CTCATCCCTG GCCGAGTGTG CTGACAGAAA ATTGGTCTCT GGTCCAAGCA TTGCTGCAGG GTCCGTGATG GTAGACATAA CCTCGTTGAT 1140
TAATTCCATC GGAATATCCC CCATAACTCG GGGTTTCTTG GCCTCCTCCA GAACATGAGA 1200 ATCAGTCATT TTCCTCTTTT CTCCTGGGTT TGCCTCAAGT CCAGAAGAGG CTTTGCAGGC 1260 AGGACTGCTG CTCCCTGCGT TTGGCTGCTC AAGGGAAGAT GAGGTTGAAT TGTATGAAAT 1320 TGTCCCAGCC ACAGGAGGTG GATTGATAAC TGTTTGGATG CCTAGCTGGC TGGTTCTGGA 1380 AGAGGCTGAG AGAAAATCCT GATCCCAGAT GGGAGAGTTT TGACTATATA CTTCTTCTTC 1440 TAGCATGGAC AGAAATTTTG GGAAATGAGT GAGGATTAGA GTTCGTTTTT CAAGAGGCAG 1500 TTTATCTTTT TCCTGTCTTG CTTGTTCCAG GAGTTGTCGC CTCATAACAG TGAAGACCGA GCGAAGCAAT GTTCTCCCAA ACACCTGTGT GGTTTCGTAC CGAGGTAGAC TGTCGCAGAA 1560 CTGTGGCACG TTGCAGTAAC ACAGCCACCT TGTGTAGTTC TCTTTGTATC CAGAAATATC 1680 ATCATTGGGA GATCGCAGTC TTCGTTGAGA TGGTGCCTCC AGATGCCAAT AGTTGATGCG 1740 GTTTAGGAAC ATTTTTGCCA ACTCAACTAT TGTTTGCCTT TCTTTTGCTG GCAGGTGACT 1800 AAATTTGTAC TGCACAAAGT TATTCACACC CTGTTCAATG CTAGGTTTTT CAAATGGGGG 1860 TTTCTTTCC AAAGAGCCTT CAACCACAGG TTTTCCTCTT TGTAAAATAG ACTTTCTCAA 1920 GAGCTTAAAT AGATAGAAAT AAACTTGTTT GGTATCTGCA TCTTCTTCCT TGTGGACACA 1980 GGTAAAGAGA TATTCCACAT CCAATACTAT TCCCAGGAGT CTGTTCATTT CTTCCTCTGA 2040 CACATTCTCC AGGTGGGAAA CATGAGCAGC TAGGGCATGG CTACAACTCC GACAGGATTC 2100 TGTTAGACTG ACAATTATTT GCTGCAGGTC GGCTCTGGGG GGAGTGGGTG AGGGGTTAGG 2160
GTTTTTCCAG CCATTACATT TACAAGACTC CTCGGCCTTG CAGGCGGAGT ACACTCCGAG 2220
TTTCTCCAGT TTCTTGGCCC GCGGAGCGGA GCGTAGTTGC GCTTTCTTCA CGGCGATTCG 2280 GGCCGAGCCA CCGCCTCCCG GTCCTTCGGC CGTGCCCGCT GCAGCCACTG CCGTCGCCGG 2340 ACCGCAGGCG CCCGAGCCCC CGGCGGCAGC GGCGCAGGGG GAGCCCTGCG GGGGCGCGGG 2400





CGGAAGCGC GCAGGCTGCG GGGGCAGCGC CCCGGGCCCG GCCCCTGCCC CGGCTCCTGC 2460 CCCGCAGCCG CCCGCCAGC CTCGGACAT 2499

# (2) INFORMATION FOR SEQ ID NO:15:

# (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2442 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

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TTGCAAGTCC	GCCATGAATA	ACTTCTTAGA	CACATAGTAC	CTGTTCCTGA	GGCGTTCACT	180
CATGGTTTTC	AGATCCATGG	GGAACCTTAT	AACTTCATAA	TATCCCGGAG	CTTCTGTTCT	240
CTTCACTGGT	TCCATGAAAG	GCCAAGCATT	TGGATGGTTC	TTCACCTGCT	GCAGGATGTT	300
CTTGAGGGTG	CTGTAAACGT	GCTCAGGGTC	TTTGGGCTCT	TTACTTTTCT	CTTTTCCACT	360
TGGTTTCCAG	CCTGTCTCTC	TGATTCCAGG	AATGCTTTCT	ATAGGAATCT	GCCGAACTCC	420
ATCTTTGAAA	CACGAAAGTC	CAGGGTAGAC	TTTTCGAATC	TGGGCTTGTT	TTCTTTCTAT	480
CAGCTTTTTA	ATGATCTCCT	TCTGCTTTTT	AATGATGACA	GAGAACTCTG	TGTATGGGAT	540
CTGAGGGTTC	AGCTCACATC	CCATCAAAGT	GGCCCCTTCA	TAATCCTTGA	TGTAGCCAAC	600
ATATTTGGTT	TTAGGTATTT	TGATTTCTTT	GGAGAAACCC	TGCTTCTTGA	AATAGCCGAT	660
GGCATACTCA	TCTGCATATG	TGAGGAAGTT	GAGGATCTCG	TGCTTTATGT	GGTATTCTTT	720
GAGATGGTTC	ATCAGGTGGG	TTCCATAGCC	CTTGACTTGT	TCATTTGAGG	TTACTGCACA	780
GAAAACAATC	TCTGTGAATC	CCTGGGATGG	AAACATCCGG	AAACAGATAC	CACCAATGAC	840
ACGGCCATCT	TTAATTAAAG	CAAGGGTTTT	GTGTTTCGGG	TCAAAGACGA	GCCGTGTGAT	900
GTACTCTTTG	GGCATTCTGG	GCAGCTGGTG	GGAAAACACA		CCACGAGCCA	960
CATCAGGATC	TTCTTGTTTG	GTTTCTGGTT	CAGGGAGTTG	CCCACCACGT		1020
GACACCCCTG	CGTTCTTCCA	GCCGTGCCGC	CTCATCTCTG	GCCGAATGGG	CTGACAGAAA	1080
ATTGGTCTCT	GGTCCAAGCA	TCCCTGCAGG	GTCTGTGATG	GTAGACATGA	CCTCATTGAT	1140
CAATTCCACG	GGAATATCCC	CCATCACTCG	AGATCTCTTG	GCCTCCTCGG	GAGCATGAGA	1200
GTTGTTCATT	TTCCTCTTTT	CTCCCGGGTT	TGCTTCAAGC	CCAGAAGAGC	CTCTGCATCC	1260
AGGACTTGTT	CTCCCTCCAT	TGATCTGCTC	ATGGGAAGTT	GAATTTGAAC	TGAACAATGC	1320
TGTCCCAGTA	ACAGGAGGAC	TGATTACTGT	TTGGATTCCT	AGCGGGCTGG	TTCTGGAAGA	1380
GGCTGAGAGA	AAATCCTGAT	CCCAGATAGG	AGAATŢTTGA	CTATACACTT	CTTCTTCCAA	1440
CATGGACAGA	AACTTTGGGA	AATGTGTGAG	GATAAGCGTG	CGTTTCTCAA	GAGGCAGTTT	1500
GTCTTTTTTC	TGTCTGGCTT	GTTCCAAGAG	CTGTCGTCTC	ATGATGGTGA	AGACCGAGCG	1560
AAGCAATGTT	CTCCCAAACA	CCTTTGTGGT	TTCGTACCGA	GGTAAGCTGT	CACAGAACTG	1620
CGGTACATTG	CAGTAGCACA	ACCACCTTGT	GTAGTTTTCC	TTGTATCCAG	AGATGTCATC	1680
ATTGGGAGAC	CGTAGTCTCC	GCTGAGATGG	AGCCTCCAGA	TGCCAGTAGT	TGATGCGGTT	1740
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CTTGAATAGG	TAGAAGTACA	CTTGTTTGGT	ATCTGCATCT	TCTTCTTTGT	GGACGCAGGT	1980
GAAGAGGTAC	TCCACATCCA	ACACAATTCC	CAGGAGTCTG	TCCATCTCTT	CCTCTGACAC	2040
ATTCTCCAAG	TGAGAAACGT	GAGCAGCAAG	GGCATGGCTA			2100
CAAACTGACA	ATTATCTGCT	GGAGGTCTCC	TCTTGGTGGA			2160
CTTCCAGCCA	TTGCATTTAC	AGGACTCCTC	TGCCTTGCAG	-	CGCCGAGTTT	2220
CTCCAGCTTC	TTCGCCCGCG	GAGCAGAGCG			CGATCCGGGC	2280
CGAGCCGCCT	•			GCCACCGGCG		2340
GCAGGAAGCA		CAGCGGTGGC			GCGGGGGGCGC	2400
GGGAGGCAGT	GCTGGGGACC	CGGCCCCGCC	AGCCTCGGCC	AT.		2442

# (2) INFORMATION FOR SEQ ID NO:16:

#### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

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# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16: CCCGCCAGCC TCGGACATGC (2) INFORMATION FOR SEQ ID NO:17:

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

#### CCCGCCAGCC TCGGCCATGC

20

20

#### (2) INFORMATION FOR SEQ ID NO:18:

#### (i) SEQUENCE CHARACTERISTICS:

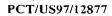
(A) LENGTH: 2442 base pairs

(B) TYPE: nucleic acid(C) STRANDEDNESS: single(D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

ATGGCCGAGG CTGGCGGGGC CGGGTCCCCA GCACTGCCTC CCGCGCCCCC, GCACGGTTCC. CCCCGGACCC TGGCCACCGC TGCCGGGAGC TCTGCTTCCT GCGGGCCAGC GACGCCGGTG 120 GCCGCGGCGG GCACCGCCGA GGGACCGGGA GGAGGCGGCT CGCCCGGAT CGCCGTGAAG 180 AAGGCGCAGT TGCGCTCTGC TCCGCGGGCG AAGAAGCTGG AGAAACTCGG CGTGTACTCC 240 GCCTGCAAGG CAGAGGAGTC CTGTAAATGC AATGGCTGGA AGAACCCTAA CCCCTCTCCT 300 ACTCCACCAA GAGGAGACCT CCAGCAGATA ATTGTCAGTT TGACAGAATC CTGTCGAAGC TGTAGCCATG CCCTTGCTGC TCACGTTTCT CACTTGGAGA ATGTGTCAGA GGAAGAGATG 360 420 GACAGACTCC TGGGAATTGT GTTGGATGTG GAGTACCTCT TCACCTGCGT CCACAAAGAA 480 GAAGATGCAG ATACCAAACA AGTGTACTTC TACCTATTCA AGCTCTTGAG AAAGTCAATT 540 TTACAAAGAG GAAAACCTGT GGTTGAAGGC TCCTTGGAGA AGAAGCCGCC ATTTGAGAAG 600 CCCAGTATTG AACAGGGTGT GAACAACTTC GTGCAGTACA AGTTTAGTCA CTTGCCATCG 660 AAAGAGAGGC AGACAACGAT CGAGCTGGCC AAGATGTTTC TGAACCGCAT CAACTACTGG CATCTGGAGG CTCCATCTCA GCGGAGACTA CGGTCTCCCA ATGATGACAT CTCTGGATAC 780 AAGGAAAACT ACACAAGGTG GTTGTGCTAC TGCAATGTAC CGCAGTTCTG TGACAGCTTA 840 CCTCGGTACG AAACCACAAA GGTGTTTGGG AGAACATTGC TTCGCTCGGT CTTCACCATC
ATGAGACGAC AGCTCTTGGA ACAAGCCAGA CAGAAAAAAG ACAAACTGCC TCTTGAGAAA
CGCACGCTTA TCCTCACACA TTTCCCAAAG TTTCTCTCAG TGTTGGAAGA AGAAGTGTAT
AGTCAAAATT CTCCTATCTG GGATCAGGAT TTTCTCTCAG CCTCTTCCAG AACCAGCCCG 900 1080 CTAGGAATCC AAACAGTAAT CAGTCCTCCT GTTACTGGGA CAGCATTGTT CAGTTCAAAT 1140 TCAACTTCCC ATGAGCAGAT CAATGGAGGG AGAACAAGTC CTGGATGCAG AGGCTCTTCT 1200 GGGCTTGAAG CAAACCCGGG AGAAAAGAGG AAAATGAACA ACTCTCATGC TCCCGAGGAG 1260 GCCAAGAGAT CTCGAGTGAT GGGGGATATT CCCGTGGAAT TGATCAATGA GGTCATGTCT 1320 ACCATCACAG ACCCTGCAGG GATGCTTGGA CCAGAGACCA ATTTTCTGTC AGCCCATTCG 1380 GCCAGAGATG AGGCGGCACG GCTGGAAGAA CGCAGGGGTG TCATTGAATT CCACGTGGTG 1440 GGCAACTCCC TGAACCAGAA ACCAAACAAG AAGATCCTGA TGTGGCTCGT GGGCCTCCAG 1500 AATGTGTTTT CCCACCAGCT GCCCAGAATG CCCAAAGAGT ACATCACACG GCTCGTCTTT 1560
GACCCGAAAC ACAAAACCCT TGCTTTAATT AAAGATGGCC GTGTCATTGG TGGTATCTGT 1620 TTCCGGATGT TTCCATCCCA GGGATTCACA GAGATTGTTT TCTGTGCAGT AACCTCAAAT GAACAAGTCA AGGGCTATGG AACCCACCTG ATGAACCATC TCAAAGAATA CCACATAAAG 1740 CACGAGATCC TCAACTTCCT CACATATGCA GATGAGTATG CCATCGGCTA TTTCAAGAAG 1800 CAGGGTTTCT CCAAAGAAT CAAAATACCT AAAACCAAAT ATGTTGGCTA CATCAAGGAT 1860 TATGAAGGGG CCACTTTGAT GGGATGTGAG CTGAACCCTC AGATCCCATA CACAGAGTTC 1920 TCTGTCATCA TTAAAAAGCA GAAGGAGATC ATTAAAAAGC TGATAGAAAG AAAACAAGCC 1980 CAGATTCGAA AAGTCTACCC TGGACTTTCG TGTTTCAAAG ATGGAGTTCG GCAGATTCCT 2040







ATAGAAAGCA	TTCCTGGAAT	CAGAGAGACA	GGCTGGAAAC	CAAGTGGAAA	AGAGAAAAGT	2100
AAAGAGCCCA	AAGACCCTGA	GCACGTTTAC	AGCACCCTCA	AGAACATCCT	GCAGCAGGTG	2160
AAGAACCATC	CAAATGCTTG	GCCTTTCATG	GAACCAGTGA	AGAGAACAGA	AGCTCCGGGA	2220
TATTATGAAG	TTATAAGGTT	CCCCATGGAT	CTGAAAACCA	TGAGTGAACG	CCTCAGGAAC	2280
AGGTACTATG	TGTCTAAGAA	GTTATTCATG	GCGGACTTGC	AACGAGTGTT	CACCAACTGC	2340
AAGGAGTACA	ACCCTCCCGA	GAGCGAGTAC	TACAAATGCG	CCAGCATCCT	GGAGAAGTTC	2400
TTCTTCAGTA	AAATTAAGGA	AGCAGGGTTG	ATTGACAAGT	GA		2442



### What is claimed is:

- 1. A purified protein designated P/CAF having a molecular weight of about 93,000 daltons as determined by sodium dodecyl sulfate polyacrylamide gel electrophoresis under reducing conditions and which acetylates histones.
- 2. The protein of claim 1 consisting of the amino acid sequence of SEQ ID NO:1.
- 3. The protein of claim 1 comprising the amino acid sequence of SEQ ID NO:2.
- 4. The protein of claim 1, which also binds to the amino acid sequence of SEQ ID NO:3 on a p300 cellular protein and to amino acid residues 1805-1854 of a CBP cellular protein (SEQ ID NO:9).
- 5. A fragment of the protein of claim 1 having histone acetyltransferase activity.
- 6. A polypeptide consisting of the amino acid sequence of SEQ ID NO: 2.
- 7. A fragment of the protein of claim 1 which binds to the amino acid sequence of SEQ ID NO: 3 on the p300 cellular protein and the amino acid sequence of SEQ ID NO:9 on the CBP cellular protein.
- 8. A polypeptide consisting of the amino acid sequence of SEQ ID NO:4.
- 9. A nucleic acid consisting of the nucleotide sequence of SEQ ID NO:10.
- 10. A nucleic acid having a nucleotide sequence which encodes the protein of claim
- 1.

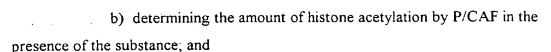


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- A nucleic acid having a nucleotide sequence which encodes the protein of claim 11. 2.
- A nucleic acid having a nucleotide sequence which encodes the protein of claim 12. 3.
- A nucleic acid consisting of the nucleotide sequence which encodes the protein 13. of claim 4.
- A nucleic acid complementary to and which selectively hybridizes with the 14. nucleic acid of claim 11 under stringent hybridization conditions
- A fragment of the nucleic acid of claim 9, which encodes a polypeptide that 15. acetylates histones.
- A fragment of the nucleic acid of claim 9, which encodes a polypeptide which 16. binds to the amino acid sequence of SEQ ID NO:3 on the p300 cellular protein and the amino acid sequence of SEQ ID NO:9 on the CBP cellular protein.
- A purified antibody which specifically binds the protein of claim 1. 17.
- A purified antibody which specifically binds the protein of claim 2. 18.
- A purified antibody which specifically binds the protein of claim 3. 19.
- A purified antibody which specifically binds the protein of claim 4. 20.
- An assay for screening substances for the ability to inhibit or stimulate the 21. histone acetyltransferase activity of P/CAF comprising:
- a) contacting the substance with a system in which histone acetylation by P/CAF can be determined;





- c) comparing the amount of histone acetylation by P/CAF in the presence of the substance with the amount of histone acetylation by P/CAF in the absence of the substance, a decreased or increased amount of histone acetylation by P/CAF in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of P/CAF.
- 22. An assay for screening substances for the ability to inhibit binding of P/CAF to p300/CBP comprising:
- a) contacting the substance with a system in which the P/CAF binding of P300/CBP can be determined;
- b) determining the amount of P/CAF binding of p300/CBP in the presence of the substance; and
- c) comparing the amount of binding of P/CAF to p300/CBP in the presence of the substance with the amount of binding of P/CAF to p300/CBP in the absence of the substance, a decreased amount of binding of P/CAF to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of P/CAF to p300/CBP.
  - The method of claim 22, wherein the system consists of a cell free reaction mixture comprising a fragment of the p300 protein comprising amino acid residues 1767-1816 (SEQ ID NO:3) and the protein of claim 4.
  - The method of claim 22, wherein the system consists of a cell free reaction mixture comprising a fragment of the CBP protein comprising amino acid residues 1805-1854 (SEQ ID NO.9) and the protein of claim 4.
  - The method of claim 22, wherein the system consists of a cell extract produced from cells producing both p300 and P/CAF.





- An assay for screening substances for the ability to inhibit or stimulate the histone acetyltransferase activity of p300/CBP comprising:
- a) contacting the substance with a system in which histone acetylation by p300/CBP can be determined;

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- b) determining the amount of histone acetylation by p300/CBP in the presence of the substance; and
- c) comparing the amount of histone acetylation by p300/CBP in the presence of the substance with the amount of histone acetylation by p300/CBP in the absence of the substance, a decreased or increased amount of histone acetylation by p300/CBP in the presence of the substance indicating a substance that can inhibit or stimulate, respectively, the histone acetyltransferase activity of p300/CBP.
  - 27. An assay for screening substances for the ability to inhibit binding of a DNA-binding transcription factor to p300/CBP comprising:
  - a) contacting the substance with a system in which the DNA-binding transcription factor binding of P300/CBP can be determined;
  - b) determining the amount of DNA-binding transcription factor binding of p300/CBP in the presence of the substance; and
- c) comparing the amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance with the amount of binding of DNA-binding transcription factor to p300/CBP in the absence of the substance, a decreased amount of binding of DNA-binding transcription factor to p300/CBP in the presence of the substance indicating a substance that can inhibit the ability to inhibit binding of DNA-binding transcription factor to p300/CBP.
  - 28. The method of claim 27, wherein the system consists of a cell free reaction mixture comprising a DNA-binding transcription factor and p300/CBP.
  - 29. The method of claim 27, wherein the system consists of a cell free reaction mixture comprising a fragment of the CBP protein comprising a DNA-binding transcription factor and p300/CBP.





- 30. The method of claim 27, wherein the system consists of a cell extract produced from cells producing both a DNA-binding transcription factor and p300/CBP.
- The method of claim 27, wherein the DNA-binding transcription factor is selected from the group consisting of a nuclear hormone receptor, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YY1, Sap-1a, c-Fos, MyoD and SRC-1
- 32. A method for inhibiting the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.
- The method of claim 32, wherein the substance can inhibit the transcription modulating activity of P/CAF by preventing the binding of P/CAF to p300/CBP.
- A method for stimulating the transcription modulating activity of P/CAF in a subject, comprising administering to the subject a transcription modulating activity stimulating amount of a substance in a pharmaceutically acceptable carrier.
- 35. The method of claim 34, wherein the substance can stimulate the transcription modulating activity of P/CAF by promoting the binding of P/CAF to p300/CBP.
- 36. The method of claim 34, wherein the substance can stimulate the transcription modulating activity of P/CAF by stimulating the histone acetlytransferase activity of P/CAF.
- 37. A method for inhibiting the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity inhibiting amount of a substance in a pharmaceutically acceptable carrier.

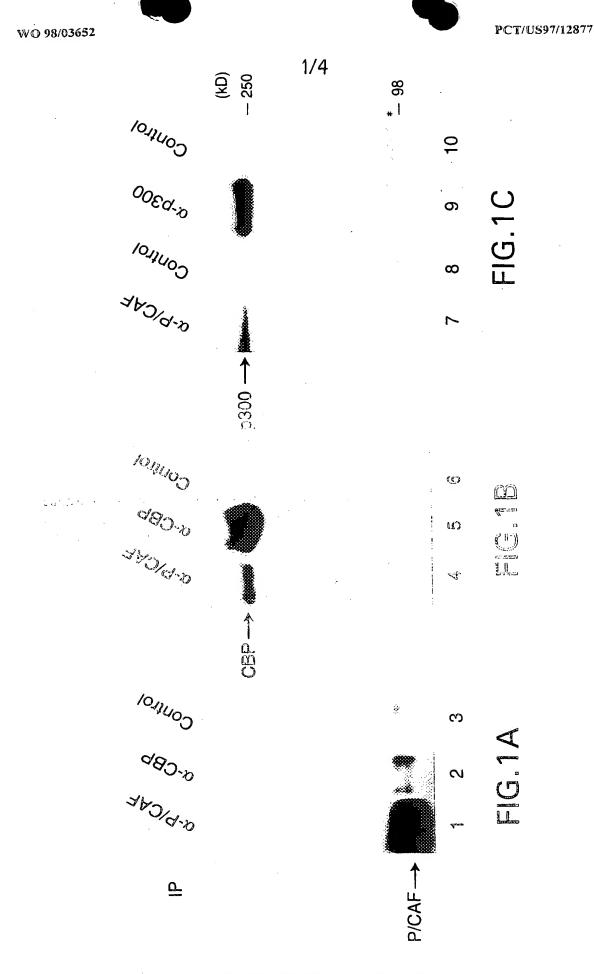




38. The method of claim 37, wherein the substance can inhibit the transcription modulating activity of p300/CBP by preventing the binding of a DNA-binding transcription factor to p300/CBP.

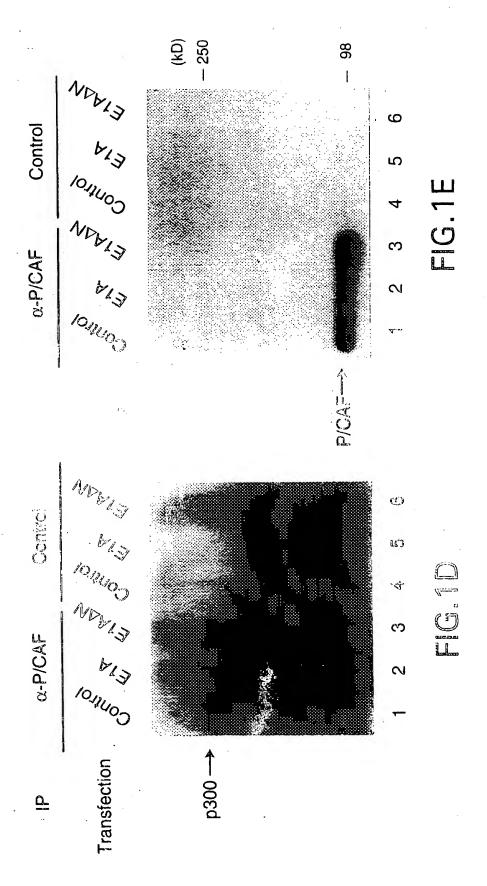
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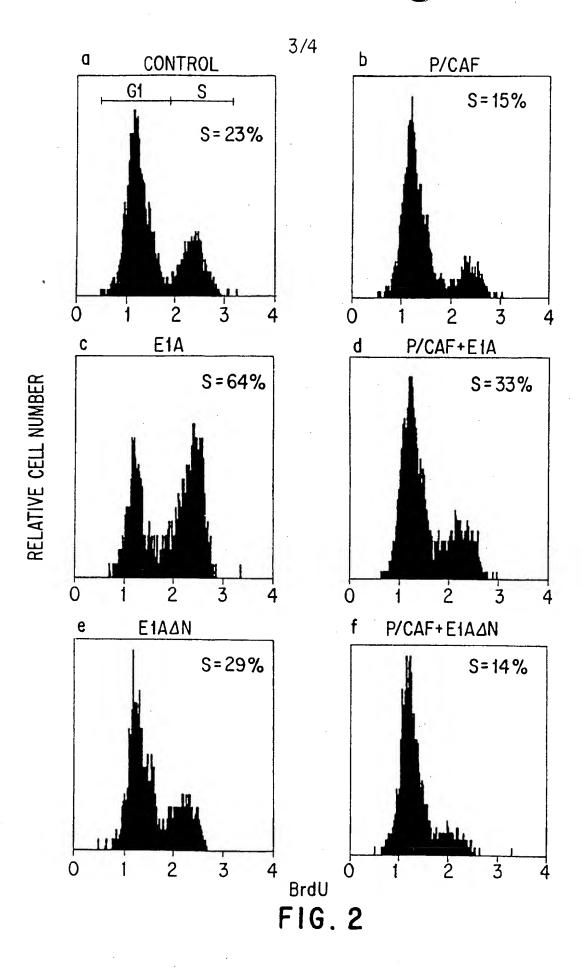
- The method of claim 38, wherein the DNA-binding transcription factor is selected from the group consisting of a nuclear hormone receptor, CREB, c-Jun/v-Jun, c-Myb/v-Myb, YY1, Sap-1a, c-Fos, MyoD and SRC-1
- The method of claim 37, wherein the substance is an antibody which binds p300/CBP.
- A method for stimulating the histone acetyltransferase activity of p300/CBP in a subject, comprising administering to the subject a histone acetyltransferase activity stimulating amount of a substance in a pharmaceutically acceptable carrier.
- The method of claim 41, wherein the substance can stimulate the histone acetyltransferase activity of p300/CBP by promoting the binding of a DNA-binding transcription factor to p300/CBP.



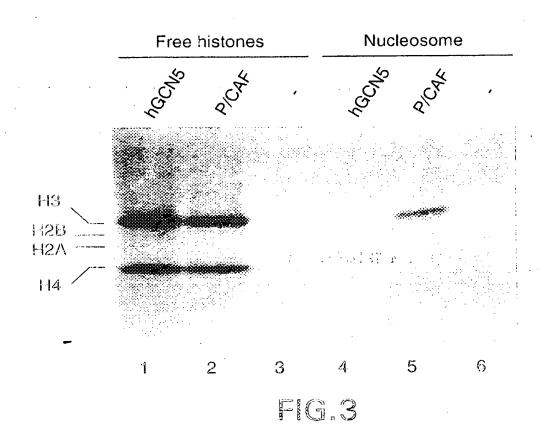
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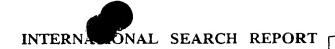




Intern .nal Application No PCT/US 97/12877

A. CLASSIF	C12N15/12 C07K14/47 G01N33/5	60 A61K38/17		
According to	International Patent Classification (IPC) or to both national classifica	tion and IPC		
B. FIELDS				
	cumentation searched (classification system followed by classification	on symbols)		
IPC 6	C07K			
Documentat	ion searched other than minimum documentation to the extent that su	uch documents are included in the fields sear	ched	
Electronic da	ata base consulted during the international search (name of data bas	se and, where practical, search terms used)		
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C. DOCUME	ENTS CONSIDERED TO BE RELEVANT			
Category °	Citation of document, with indication, where appropriate, of the rele	evant passages	Relevant to claim No.	
	- 1 N20500		1	
Α	EMBL EST, Accession numberN39522 Sequence no yv27b08.sl Homo sapi	ens cDNA	1	
	clone 243927 3'			
	25 January 1996			
	XP002050402 see the whole document			
		7.44		
Α	GEORGAKOPOULOS, T. & THIREOS, G.	:	1	
	EMBO JOURNAL., vol. 11, 1992, EYNSHAM, OXFORD	GB,		
	pages 4145-4152, XP002050399			
	see the whole document	·		
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	her documents are listed in the continuation of box C.	Patent laminy members are instead in		
	ategories of cited documents :	"T" later document published after the interr or priority date and not in conflict with the	ne application but	
*A* document defining the general state of the art which is not cited to understand the principle or theory underlying the considered to be of particular relevance invention				
*E* earlier document but published on or after the international filling date  *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to				
"L" document which may throw doubts on priority claim(s) or involve an inventive step when the document is taken alone which is cited to establish the publication date of another "Y" document of particular relevance; the claimed invention			aimed invention	
O docum	citation or other special reason (as specified)  Cannot be considered to involve an inventive step when the document referring to an oral disclosure, use, exhibition or document is combined with one or more other such docu-			
*P* docum	means ent published prior to the international filing date but	ments, such combination being obvious in the art.		
	han the priority date claimed	*&" document member of the same patent for  Date of mailing of the international search		
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1	7 December 1997	1 T. UI. JU		
Name and mailing address of the ISA  Authorized officer				
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	ation) DOCUMENTS CONSIDERED TO BE RELEVANT  Citation of document, with indication, where appropriate, of the relevant passages	. Relevant to claim No.
Category °	Citation of document, with indication, where appropriate, or the relevant passages	
P,X	YANG, X.Y. ET AL.: "A p300-CBP-associated factor that competes with the adenoviral oncoprotein E1A" NATURE., vol. 382, no. 8589, 25 July 1996, LONDON GB, pages 319-324, XP002050400 see the whole document	1
P,X	OGRYZKO, V.V. ET AL.: "The transcriptional coactivators p300 and CBP are histone acetyltransferases" CELL, vol. 87, no. 5, November 1996, NA US, pages 953-959, XP002050401 see the whole document	1
P,X	EMBL EST, Accession number U57316, Sequence reference human GCN5 (hGCN5) complete cds. 26 august 1996 XP002050403 see the whole document	



# INTERNATIONAL SEARCH REPORT

PCT/US 97/12877

BoxI	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This Inte	ernational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:  see FURTHER INFORMATION sheet PCT/ISA/210
2.	Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Inte	ernational Searching Authority found multiple inventions in this international application, as follows:
-	
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4.	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remar	The additional search fees were accompanied by the applicant's protest.  No protest accompanied the payment of additional search fees.



# FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Remark: Although claims 32 to 42 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the composition.